

**Taxonomic Revision and Molecular Studies  
of  
*Garcinia* Section *Garcinia* (Guttiferae)**

**M. Nazre Saleh**

**Doctor of Philosophy  
The University of Edinburgh  
&  
Royal Botanic Garden, Edinburgh  
2006**

**Declaration**

I hereby declare that this thesis has been composed and the work contained within,  
unless otherwise acknowledged and cited, is my own.

## Abstract

*Garcinia* section *Garcinia* is one of 14 sections of the species-rich pantropical genus *Garcinia* (Guttiferae/Clusiaceae). In its most recent circumscription the section comprised 43 species, mostly of rain forest understorey trees, distributed from eastern India to Fiji, and in Madagascar (Jones, 1980: unpublished Phd. Thesis, University of Leicester). Its most famous member is the fruit tree, mangosteen (*G. mangostana*).

Taxonomic revision of section *Garcinia* reduces the number of species from 43 to 16 with five varieties, with distribution from eastern India to Malesia. A total of nine species are excluded, 19 species are reduced to synonyms and five species are insufficiently known to be classified. Within sect. *Garcinia*, *G. acuticosta*, *G. discoidea*, *G. exiguous*, *G. ochraceus* and *G. sanguisanguis* are newly described, *G. diospyrifolia* var. *arborea*, *G. diospyrifolia* var. *minor* and *G. malaccensis* var. *pseudomangostana* are new varieties, and *G. cataractalis*, previously unassigned to any section by Jones (1980) is newly included. Species limits are defined on the basis of combinations of characters such as shape of stamens and presence of pistillode, fruit type, and leaf characters such as shape, size, venation pattern and type of glandular lines.

A total of 83 accessions representing 42 *Garcinia* species were sequenced for the nuclear ribosomal internal transcribed spacer (ITS) region, while 50 accessions (30 spp.) and 24 accessions (16 spp.) were sequenced for the chloroplast non-coding regions *trnS-G* and *trnD-T* respectively. Of these, 30 accessions or ten spp. (ITS), 23 accessions or nine spp. (*trnS-G*) and 17 accessions or nine spp. (*trnD-T*) belong to sect. *Garcinia*. Parsimony and Bayesian analyses show that the ITS data are more phylogenetically informative and provide better resolution for sectional and species relationships compared to *trnS-G* and *trnD-T*. Most of the sections delimited by Jones (1980) proved to be monophyletic but sect. *Garcinia* is not. *Garcinia maingayi*, *G. trianaii* and *G. costata*, assigned to sect. *Garcinia* by Jones, are in a separate clade that is strongly supported as sister to sect. *Brindonia*. These species have significantly different morphological characters from sect. *Garcinia*, and should be excluded from it.

Characters that are important for sectional delimitation are inflorescences that are simple cymes, stamen bundles that are 4-angled or 4-lobed, and fruits with a smooth surface. Four major clades in sect. *Garcinia* are supported by the shape of the stamen bundles, the shape of the fruit and the stigma, and fruit wall characters. ITS trees are significantly incongruent with plastid trees because of the placement of *G. rigida*, which could reflect a hybrid origin. Another hybrid species, the cultivated mangosteen, could be the product of hybridisation events between varieties of *G. malaccensis* if mangosteen is proven to be an obligate agamosperm. However, if mangosteen is a facultative agamosperm, *G. malaccensis* is likely to be the female parent, but any other *Garcinia* species could be the putative paternal species.

Phylogenetic trees of ITS show that most *Garcinia* species from east of Wallace's Line are nested within species from the west. This might reflect dispersal of species across this biogeographic division from west to east when the Sahul and Sunda shelves converged (c. 20 MY). This hypothesis is supported by the estimated divergence of accessions of *G. rigida* from the east of Wallace's Line, from their most recent common ancestor to the west of the line, not later than 21.58±2.90 MY.

## Acknowledgements

My special thanks to my supervisors, Toby Pennington and Mark Newman, for their advice and support throughout the period of my study in RBGE and I would also like to thank to James Richardson, David Middleton and Kwiton Jong for their valuable advice and help in preparing this thesis.

In RBGE, I am grateful to Martin Pullan for helping me with the PADME database, Michelle Hollingsworth and Alex Clark for all their assistance in the lab, David Harris, Adele Smith, and Helen Hoy for helping me to manage my herbarium specimens, Robert Mill, Mai-Lan Krantiz, Helene Citerne and Daniella Schill for the translation works, Mark Hughes for digital image and DIVA-GIS and many other people especially in the library for their direct and indirect help.

I would also like to thanks to the curator of these herbaria; K, SING, UC, A, L, P, and SAR for the loan materials. Curator and staff of the following herbaria during my visit: K, BM, KEP, KINA, MARDI, SAR and SAN.

During the course of my visit and fieldwork, I would personally thankful to LIPI and Bogor Botanic Gardens especially to Hendrian, Dadi Darnadi, Rismita Sari and Irawaty; Sarawak Forestry Corporation, especially to Nigel Lim, Runi Pungga, Lucy Chong, Bibian Diway, Andela Tipot and Sylvester Tan; Sabah Parks and Sabah Forestry Department especially to Jamili Nais, John Sugau and Bernard Valentine; and in Peninsular Malaysia, the Forestry Department and UPM especially to Prof. Hamami Hj Shaari, Pn. Salmah, Tabingon, Shamsul Khamis, Radhiah Zakaria and Nur Abdullah Psyquay; in MARDI especially to Salma Idris and Masron, and lastly to my good travel companion and tree climber, Jantan Mok (Unyit).

In RBGE there are many friends that I would like to appreciate; Vanessa, Estelle, Bruce, Greg, Sophie, Gemma, Camila, Daniela, Cath, Vimi, Peter, Mai-Lan, Alan, Barbara, Jin, Will, Chris, Kate, Colin, Haja and to all Malaysian friends who have made me and family enjoy our stay in Edinburgh, may Allah bless us all.

My personal grateful and special thank also to my family in Kota Tinggi and Sepang who gave their valuable support for most of the time, to 'emak & abah', I am grateful for your 'doa'.

Last but not least, to my beloved wife Suri and my daughter Sarah, the backbone of my life and works, thank you for the support and understanding.

## Contents

Abstract

Acknowledgements

<b>CHAPTER 1 – INTRODUCTION</b>	1
1.1 General Background	1
1.2 Project Aims and Objectives	3
1.3 Thesis Structure	4
<b>CHAPTER 2 - TAXONOMIC HISTORY</b>	5
2.1 Introduction	5
2.2 The Genus <i>Garcinia</i>	5
2.2.1 <i>Generic Limit</i>	8
2.2.2 <i>Infrageneric Classifications of Garcinia</i>	11
2.3 Section <i>Garcinia</i> sensu Jones	19
<b>CHAPTER 3 - THE ECOLOGY AND BIOLOGY OF SECTION GARCINIA</b>	21
3.1 Introduction	21
3.2 Habit	21
3.3 Distribution and Dispersal	23
3.4 Floral and Fruit Biology	25
3.5 Agamospermy and <i>Garcinia</i>	28
3.6 Chromosome Numbers	32
3.7 Ethnobotanical Uses	33
3.8 Natural Products and Pharmacological Potential	35
<b>CHAPTER 4 – MORPHOLOGICAL CHARACTERS</b>	37
4.1 Introduction	37
4.2 Materials and Methods	37
4.3 Vegetative Characters	39
4.3.1 <i>Leaves</i>	39
4.3.1.1 <i>Colour</i>	40
4.3.1.2 <i>Shape and Size</i>	40
4.3.1.3 <i>Petiole</i>	42
4.3.1.4 <i>Midrib</i>	42
4.3.1.5 <i>Secondary Veins</i>	44
4.3.1.6 <i>Glandular Lines</i>	45
4.4 Floral Characters	48
4.4.1 <i>The Inflorescences</i>	48
4.4.2 <i>Sepals and Petals</i>	50
4.4.3 <i>Male Flowers</i>	51
4.4.3.1 <i>Stamens</i>	51
4.4.3.2 <i>Pistillodes</i>	54
4.4.4 <i>Female Flowers</i>	55
4.4.4.1 <i>Ovary</i>	55
4.4.4.2 <i>Stigma</i>	55

4.4.4.3	<i>Staminodes</i>	57
4.5	Fruits	58
4.5.1	<i>Fruit Wall</i>	58
4.5.2	<i>Colour</i>	60
4.5.3	<i>Shape and Size</i>	60
4.5.4	<i>Stigma</i>	62
4.5.5	<i>Seed</i>	64
<b>CHAPTER 5 – MOLECULAR STUDIES OF SECTION GARCINIA</b>		65
5.1	Introduction	65
5.1.1	<i>Classification of Section Garcinia Sensu Jones</i>	65
5.1.2	<i>Molecular Studies of Garcinia</i>	68
5.1.3	<i>Hybridisation in Garcinia</i>	70
5.1.4	<i>Biogeography of Section Garcinia</i>	71
5.2	Objectives	73
5.3	Materials and Methods	73
5.3.1	<i>Region Chosen</i>	73
5.3.2	<i>Taxonomic Sampling</i>	75
5.3.3	<i>Outgroup Taxa</i>	76
5.3.4	<i>DNA Extraction</i>	77
5.3.5	<i>Amplification</i>	77
5.3.6	<i>Sequencing</i>	79
5.3.7	<i>Sequence Alignment</i>	79
5.3.8	<i>Phylogenetic Analysis</i>	80
5.3.8.1	<i>Choice of Methods for Phylogenetic Inference</i>	80
5.3.8.2	<i>Maximum Parsimony</i>	83
5.3.8.3	<i>Bayesian Analysis</i>	84
5.3.8.4	<i>Incongruence Length Difference Test</i>	85
5.3.8.5	<i>Age Estimation</i>	85
5.4	Results	87
5.4.1	<i>ITS Data</i>	87
5.4.2	<i>TrnS-trnG</i>	97
5.4.3	<i>Combined trnS-trnG and trnD-trnT</i>	101
5.4.4	<i>Congruence Test</i>	104
5.4.5	<i>Age Estimation</i>	105
5.5	Discussion	106
5.5.1	<i>Utility of ITS and cpDNA Sequences</i>	106
5.5.2	<i>Section Garcinia and Morphology</i>	107
5.5.3	<i>Phylogenetic Relationships within Sect. Garcinia</i>	109
5.5.4	<i>Hybridisation and Origin of Mangosteen</i>	115
5.5.5	<i>Distribution and Biogeography of Sect. Garcinia</i>	124
5.6	Conclusion	125

<b>CHAPTER 6 – TAXONOMIC REVISION OF <i>GARCINIA</i> SECTION <i>GARCINIA</i></b>	127
6.1 Introduction	127
6.2 Agamospermy and Species Delimitation	127
6.3 Species Concept Used In This Study	130
6.4 Groups in Section <i>Garcinia</i>	133
6.5 Generic Description	135
6.6 Section Description	137
6.7 Identification Keys	139
6.7.1 <i>Key to Species from Vegetative Characters</i>	139
6.7.2 <i>Key to Species for Fertile Specimens</i>	141
6.8 Species Description	145
6.8.1 <i>Species with Unknown Status</i>	202
6.8.2 <i>Excluded Species</i>	207
6.9 List of Exsiccatae	212
 <b>CHAPTER 7 – GENERAL CONCLUSION AND RECOMMENDATIONS</b>	 220
7.1 Delimitation and taxonomy of Section <i>Garcinia</i>	220
7.2 Molecular Study on Section <i>Garcinia</i>	221
7.3 Future Work and Recommendations	223
 <b>BIBLIOGRAPHY</b>	 228
 <b>APPENDICES</b>	
Appendix I. Scientific names listed by Jones (1980) with associated taxa taken from IPNI; names in bold currently belong to sect. <i>Garcinia</i> as defined in this thesis (Chapter 6) and asterisks denote originally listed by Jones.	
Appendix II. Species, accessions and origin of samples used in molecular analysis for this study.	
Appendix III. List of sequences taken from GENBANK used in this study.	
Appendix IV. Molecular lab protocol.	
Appendix V. Matrix of internal transcribed spacer (ITS) sequences with insertion and deletion characters.	
Appendix VI. Matrix of <i>trnS-trnG</i> sequences and binary gap coded characters.	
Appendix VII. Matrix of <i>trnD-trnT</i> sequences and binary gap coded characters.	
Appendix VIII. Electropherogram of <i>Garcinia rigida</i> Miq. (accession 045) upstream sequence of ITS.	
Appendix IX. Electropherogram of <i>Garcinia mangostana</i> L. (accession 096) upstream sequence of ITS	

List of Tables

List of Figures

List of Maps

## List of Tables

### Chapter 2

Table 2.1. Historical events in the infrageneric classification of genus <i>Garcinia</i> .	14-16
Table 2.2. Groups, subgroups and sections established by Jones (1980).	17
Table 2.3. Comparison of the section names used by Jones, Engler & Pierre.	18-19
Table 2.4. Species number and geographical distribution of 14 sections of <i>Garcinia</i> sensu Jones (1980).	20

### Chapter 3

Table 3.1. Chromosomes numbers of some <i>Garcinia</i> species.	33
Table 3.2. Some natural products extracted from species of sect. <i>Garcinia</i> .	35
Table 3.3. Species with pharmacological uses in sect. <i>Garcinia</i> .	36

### Chapter 5

Table 5.1. Summary of the accession and sequence numbers used in this study.	76
Table 5.2. Primers used for amplification and sequencing.	78
Table 5.3. Summary of sequences information, analysis parameters and tree characteristics from Maximum Parsimony and Bayesian Inference analyses. CI = Consistency Index, RC = Rescaled Consistency Index, HI = Homoplasy Index, RI = Retention Index, hLRT = Hierarchical Likelihood Ratio Test, GTR+I+G = General Time Reversible with Invariant sites Estimated and a Gamma distribution of rates, HKY +G= Hasegawa, Kishino & Yano and a Gamma distribution of rates.	88
Table 5.4. Estimated time of divergence of <i>Garcinia</i> based on the published ITS data.	106

## List of Figures

### Chapter 1

- Figure 1.1 Exudates from *Garcinia* trees. A: *G. penangiana* Pierre; B: *G. discoidea* Nazre 1

### Chapter 3

- Figure 3.1. Worldwide distribution of sect. *Garcinia*, showing Wallace's Line 25

### Chapter 4

- Figure 4.1. Leaf shape and size. — **a.** *Garcinia exiguus* Nazre (Coode *et al.* 7886, A) scale bar=1 cm; **b.** *G. mangostana* L (Salma & Sohaima 787, MARDI); **c.** *G. harmandii* Pierre (Thorel 1018., K); **d.** *G. cataractalis* Whitmore (Whitmore FRI20350, K); **e.** *G. diospyrifolia* var. *minor* Ng ex Nazre (Wong 32216, A). Scale bar = 1 cm. 41
- Figure 4.2. Petiole and ligule. — **a.** *G. venulosa* (Blanco) Choisy (Cumming 2114, K) showing ligule (arrow); **b.** *Garcinia celebica* L. (Anon 115, K) showing striae (arrow). 42
- Figure 4.3. Abaxial surfaces of the midribs (above) and cross sections (below). — **a.** *Garcinia nitida* Pierre (Argent 9318, E); **b.** *G. celebica* L. (Harreveld *s.n.*, L); **c.** *G. moselleyana* Pierre (Moseley *s.n.*, K). 43
- Figure 4.4. Secondary venation; a – b: brochidodromous, c-d: cladodromous. — **a.** *Garcinia mangostana* L. (Shamsudin *s.n.*, MARDI), arrow highlight two intramarginal veins; **b.** *G. penangiana* Pierre (Henderson 21689, K), arrow highlight inter-secondary veins; **c.** *G. moselleyana* Pierre (Wenzel 3036, K); **d.** *G. harmandii* Pierre (Thorel 1018, K). 45
- Figure 4.5 Glandular line patterns, arrows highlight glandular lines. — **a.** *Garcinia penangiana* Pierre (Henderson 21689, K); **b.** *G. exiguus* Nazre (Coode 7886, A); **c.** *G. nitida* Pierre (Haviland 2339, K); **d.** *G. sangudsangud* Nazre (Aban SAN32977, L). 47
- Figure 4.6. Inflorescences in section *Garcinia* (a – b) and omitted species from sect. *Garcinia* (c). — **a.** *Garcinia celebica* L. (Hamilton 1113, A); **b.** *G. mangostana* L. (Shamsudin *s.n.*, MARDI); **c.** *G. maingayi* Hk. f. (Curtis 1610, K). 49
- Figure 4.7. Variation of male flowers; Type 1=a, Type 2=b and Type 3= c to g. — **a.** *Garcinia celebica* L. (Nazre 062, E); **b.** *G. sangudsangud* Nazre (Aban SAN 32914, L); **c.** & **d.** *G. malaccensis* var. *malaccensis* (c. Ashton BRUN3392, SAR; d. Maingay 149, K); **e.** *G. mangostana* L. (Shamsudin *s.n.*, MARDI); **f.** *G. penangiana* Pierre (Whitmore FRI20347, L); **g.** *Garcinia malaccensis* var. *pseudomangostana* Nazre (Madani 50574, L). Scale bar = 1 cm. Drawn by Anna Dorward. 52
- Figure 4.8. Comparison of male flowers in section *Garcinia* (a) and omitted species (b). — **a.** *G. celebica* L.; **b.** *G. costata* Hemsl. ex King. 53
- Figure 4.9. Variations of pistillode (a–d), bar and arrow highlight the pistillode; species without a pistillode (c–f.) — **a.** *Garcinia celebica* L. (Nazre 062, E); **b.** *G. mangostana* L. (Shamsudin *s.n.*, MARDI); **c.** & **d.** *G. malaccensis* var. *malaccensis* (c. Ashton BRUN3392, SAR; d. Cockburn FRI18181, KEP); **e.** *G. venulosa* (Blanco) Choisy (Alvarez 18513, K); **f.** *G. penangiana* Pierre (Burbridge *s.n.*, K). 54

- Figure 4.10. Variation in ovaries and stigmas, bars indicate ovary and arrows indicate stigma; a–c: ovoid or broadly ovate ovary with fungiform cap-shaped stigma; d: globose ovary with skull cap-shaped stigma; e: stigma in 4-bundles with corrugated surface. — **a.** *G. diospyrifolia* var. *diospyrifolia* (Zainal AA914, A); **b.** *G. nitida* Pierre (Haviland 2339, K); **c.** *G. celebica* L. (Anon 115, K) **d.** *Garcinia moselleyana* Pierre (Elmer 7187, K); **e.** *G. penangiana* Pierre (Chai S32334, L). 56
- Figure 4.11. The position of free staminodes in female flowers of *Garcinia mangostana* L. Arrow highlights staminodes. 57
- Figure 4.12. Fruit wall type; a. thick and b. thin and easily ruptured. — **a.** *G. celebica* L. (Blume 2026, L); **b.** *G. diospyrifolia* var. *minor* Ng ex Nazre (Wong 32216, A). 59
- Figure 4.13. Comparison of fruit wall surfaces in section *Garcinia* (a) with omitted species from the section (b, c). — **a.** *Garcinia discoidea* Nazre with smooth wall without stripe; **b.** *G. costata* Hemsl. ex King with ribbed wall; **c.** *G. trianii* Pierre with striped fruit wall. 60
- Figure 4.14. Variations of fruits, bars highlight elongation of stigma. — **a.** *Garcinia discoidea* Nazre (Shamsul SA02-10, E); **b.** *G. cataractalis* Whitmore (Whitmore FRI 20350, K); **c. & d.** *G. celebica* L. (c: Burkill HMB259, SING; Yahya SFN21419, SING); **e.** *G. diospyrifolia* var. *diospyrifolia* (Soengeng 47, L); **f.** *G. harmandii* Pierre (Pierre 1371, K); **g.** *G. exiguus* Nazre (Chai S31750, SAR); **h.** *G. nitida* Pierre (Argent & Amiril 9318, K); **i.** *G. malaccensis* var. *malaccensis* (Whitmore FRI 8986, L). Drawn by Anna Dorward. 61
- Figure 4.15. Variations of the fruit stigmas; distinctly lobed (a – c); weakly lobed (d – i); and without lobes (j – m). — **a.** *Garcinia mangostana* L. (Sibat S23041, L.); **b.** *G. malaccensis* var. *malaccensis* (Whitmore FRI 8986, L); **c.** *G. penangiana* Pierre (Sinclair SFN 40794, SING); **d.** *G. discoidea* Nazre (Henderson 19654, L); **e.** *G. cataractalis* Whitmore (Whitmore FRI 20350, K); **f.** *G. celebica* L. (Burkill HMB259, SING); **g.** *G. diospyrifolia* var. *diospyrifolia* (Kochummen 77731, L); **h.** *G. discoidea* Nazre (Shamsul SA01-10, E); **i.** *G. exiguus* Nazre (Chai S31750, SAR); **j.** *G. harmandii* Pierre (Thorel 1018, K); **k.** *G. nitida* Pierre (Argent & Amiril 9318, K); **l.** *G. ochraceus* Nazre (Streiman & Martin 52890, E); **m.** *G. sangudsangud* Nazre (Muin Chai SAN 33449, L). Scale bar = 1 cm. 63

## Chapter 5

- Figure 5.1. Hypothetical relationships of sections in *Garcinia* sensu Jones (1980). Tree originally drawn by Jones with some modification showing characters for her artificial groupings and subgroups. *Garcinia* was firstly divided into informal groups (characters 15-17) and subgroups (characters 18-22) then were divided to form sections (characters 1-14). 67
- Figure 5.2. The position of Wallace's Line and Huxley's Line. 72
- Figure 5.3. One of 10, 000 most parsimonious trees from the analysis of the ITS data. Numbers above branches indicate bootstrap values (shown for values above 60% only). The grey branches indicate the topology of the strict consensus tree. Tree length = 998 steps, CI = 0.50, RI = 0.88 and HI = 0.51. The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). 93-94

- Figure 5.4. Fifty percent Majority Rule consensus tree resulting from the Bayesian analysis of ITS. Numbers above branches indicate posterior probabilities (shown for values above 60% only). The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). Origin of samples collected; CEL=Sulawesi, JAV=Java, KAL=Kalimantan, LAO=Laos; MAL=Maluku; MY=Peninsular Malaysia, NWG=New Guinea, S-AM=South America, SBH=Sabah, SRL=Sri Lanka, SUM=Sumatra, TH=Thailand, Vn=Vietnam; Purple=Sri Lanka, Blue=Indochina & Thailand, Yellow=Peninsular Malaysia, Red=Malay Islands, Orange=Australia, Grey=South America and White=unknown origin. 95-96
- Figure 5.5. One of 10, 000 most parsimonious trees from the analysis of the *trnS-G* region. Numbers above branches indicate bootstrap values (shown for values above 60% only). The grey branches indicate the topology of the strict consensus tree. Tree length=525 steps. CI=0.81, RI=0.76 and HI=0.19. The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). 99
- Figure 5.6. Fifty percent Majority Rule consensus tree resulting from Bayesian analysis of the *trnS-G* region. Numbers above branches indicate posterior probabilities (shown for values above 60% only). The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). 100
- Figure 5.7. One of 10, 000 most parsimonious trees from the analysis of the *trnS-G* and *trnD-T* regions combined. Numbers above branches indicate bootstrap values (shown for values above 60% only). The grey branches indicate the strict consensus tree. Tree length=514 steps. CI=0.87, RI=0.79 and HI=0.14. The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). 102
- Figure 5.8. Fifty percent Majority Rule consensus tree resulting from Bayesian analysis of combined *trnS-G* and *trnD-T* sequences. Numbers above branches indicate posterior probabilities (shown for values above 60% only). The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). 103
- Figure 5.9. Groups within section *Garcinia* inferred from Bayesian analysis of ITS. Origin of samples collected; CEL=Sulawesi, JAV=Java, KAL=Kalimantan, LAO=Laos, MAL=Maluku, MY=Peninsular Malaysia, SBH=Sabah, SUM=Sumatra, TH=Thailand and VN=Vietnam. 111

## Chapter 6

- Figure 6.1. *Garcinia acuticosta* Nazre. **A.** habit (male); **B.** glandular lines; **C.** fruit and stigma [Nazre BB01, E]. 147
- Figure 6.2. *Garcinia cataractalis* Whitmore. **A.** habit (male); **B.** lamina and glandular lines; **C, D, E.** fruits and stigma. [Whitmore FRI 20350, K]. 150
- Figure 6.3. *Garcinia celebica* L. **A.** Habit (male); **B.** glandular lines; **C.** male flower; **D.** female flower; **E.** fruit and stigma. [(A & B: Curtis 690, SING); (C: Nazre 062, E); (D: Nazre s.n., E); (Ei:Burkill HMB259, SING); (Eii: Ahmad Shukor AS29, SING); (Eiv: Yahya SFN21419, SING); (Ev: Pierre 700, K)]. 157
- Figure 6.4. *Garcinia diospyrifolia* var. *diospyrifolia*. **A.** habit (female); **B, C.** fruit and stigma; **D.** glandular lines. [(A, B, C, D: Soegeng 47, L); (B: Kochummen 7773, L)]. 162

- Figure 6.5. *Garcinia diospyrifolia* var. *arborea* Nazre. **A.** habit (female); **B.** female flower; **C, D.** fruits and stigma; **E.** glandular lines [(A, B, C, E: *King's Collector 7232*, P); (D: *Pereira JTP65*, E)]. 163
- Figure 6.6. *Garcinia diospyrifolia* var. *minor* Ng ex Nazre. **A.** habit (female); **B, C.** fruit and stigma [(A, B: *Wong 32216*, A); (C: *Rosemary 541*, L)]. 164
- Figure 6.7. *Garcinia discoidea* Nazre. **A.** habit (female); **B.** male flower; **C, D.** fruit and stigma. [(A, C: *Shamsul SA02-10*, E); (B: *Corner s.n.*, SING); (D: *Kadim KN307*, L)]. 167
- Figure 6.8. *Garcinia exiguus* Nazre. **A.** habit (female); **B.** glandular lines; **C:** fruit and stigma [*Chai S31750*, SAR]. 169
- Figure 6.9. *Garcinia harmandii* Pierre. **A.** habit (male); **B:** inflorescence; **C:** fruit and stigma [(A, B: *Pierre 1371*, K); (C: *Thorel 1018*, K)]. 172
- Figure 6.10. *Garcinia malaccensis* var. *malaccensis*. **A.** habit (male); **B.** glandular lines; **C.** male flowers; **D.** female flower; **E, F.** fruit and stigma; **G.** secondary nervations with two intra-marginal veins [(A, B, Cii: *Maingay 149*, K); (Ci: *Ashton BRUN3392*, SAR); (D: *Maingay 149/Barcode 489504*, L); (E: *Wyatt-Smith 80555*, L); (F, G: *Whitmore FRI8986*, L)]. 176
- Figure 6.11. *Garcinia malaccensis* var. *pseudomangostana* Nazre. **A.** habit (male); **B.** fruit and stigma; **C.** glandular lines; **D.** female flower; **E.** secondary nervations with two intra-marginal veins [(A, C: *Madani 50574*, L); (Bi: *Meijer SAN 19236*, L); (Bii: *Fidilis & Sumbing SAN 96472*, L); (D, E: *Sigin & Joseph SAN 97194*, L)]. 177
- Figure 6.12. *Garcinia mangostana* L. **A.** habit; **B:** female flower; **C:** fruit; **D:** male flower [(A, C: *Sibat S23041*, L.); (B: *Anon*, MARDI); (D: *Shamsudin s.n.*, MARDI)]. 181
- Figure 6.13. *Garcinia moselleyana* Pierre. **A.** habit (male); **B:** inflorescences [*Moseley s.n.*, K]. 183
- Figure 6.14. *Garcinia nitida* Pierre. **A.** habit; **B.** glandular lines; **C.** stigma; **D:** fruits and stigma [(A, C, B: *Argent & Amiril 9318*, K); (D: *Ilias 13317*, K)]. 186
- Figure 6.15. *Garcinia ochraceus* Nazre. **A.** habit (male); **B.** male flower; **C, E.** fruits and stigma; **D.** glandular lines [(A, B: *Streiman & Martin 52886*, E); (C, D, E: *Streiman & Martin 52890*, E)]. 189
- Figure 6.16. *Garcinia penangiana* Pierre. **A.** habit (male); **B:** young fruit and stigma; **C:** male flower; **D, E:** mature fruit [(A, C: *Whitmore FRI 20347*, L); (B: *Sibat S25238*, K); (D: *Sibat S23622*, K); (E: *Sinclair SFN 40794*, SING)]. 192
- Figure 6.17. *Garcinia rigida* Miq. **A.** habit (female); **B.** fruit and stigma [*Harmand 4012*, K]. 195
- Figure 6.18. *Garcinia sangudsangud* Nazre. **A.** habit (male); **B.** male flower; **C.** glandular lines; **D.** young fruit; **E.** mature fruits and stigma [(A, B, C: *Aban SAN 32914*, L); (D: *Meijer SAN 22942*, L); (Ei: *Muin Chai SAN 33449*, L); (Eii: *Burley 2660*, E)]. 198
- Figure 6.19. *Garcinia venulosa* (Blanco) Choisy. **A.** habit (female); **B.** glandular lines; **C.** male flower; **D.** fruits and stigma [(A, B: *Cuming 1124*, K); (C: *Alvarez 18513*, K); (D: *Curran 17827*, K)]. 201

## List of Maps

### Chapter 6

Map 6.1. Distribution of <i>Garcinia acuticosta</i> Nazre	146
Map 6.2. Distribution of <i>Garcinia cataractalis</i> Whitmore	149
Map 6.3. Distribution of <i>Garcinia celebica</i> L.	156
Map 6.4. Distribution of <i>Garcinia diospyrifolia</i> Pierre. <i>G. diospyrifolia</i> var. <i>diospyrifolia</i> (circle); <i>G. diospyrifolia</i> var. <i>arborea</i> (triangle); <i>G. diospyrifolia</i> var. <i>minor</i> (square).	161
Map 6.5. Distribution of <i>Garcinia discoidea</i> Nazre	166
Map 6.6. Distribution of <i>Garcinia exiguus</i> . Nazre	170
Map 6.7. Distribution of <i>Garcinia harmandii</i> Pierre.	171
Map 6.8. Distribution of <i>Garcinia malaccensis</i> Hook. f. <i>G. malaccensis</i> var. <i>malaccensis</i> (circle); <i>G. malaccensis</i> var. <i>pseudomangostana</i> (triangle).	178
Map 6.9. Distribution of <i>Garcinia moselleyana</i> Pierre.	184
Map 6.10. Distribution of <i>Garcinia nitida</i> Pierre.	187
Map 6.11. Distribution of <i>Garcinia ochraceus</i> Nazre.	188
Map 6.13. Distribution of <i>Garcinia penangiana</i> Pierre	191
Map 6.14. Distribution of <i>Garcinia rigida</i> Miq.	196
Map 6.16. Distribution of <i>Garcinia sanguisangud</i> Nazre	199
Map 6.17. Distribution of <i>Garcinia venulosa</i> (Blanco) Choisy.	202

## CHAPTER 1 INTRODUCTION

### 1.1 General Background

*Garcinia* L. is one of the biggest genera in Guttiferae (Clusiaceae) comprising an estimated 200 species (Stevens, 2001 onwards). The species are small to large trees or occasionally shrubs found in the tropical rain forest. In Asia, its distribution is centralised in the Malesian region, with some species reaching India and the Micronesian islands, but it is also found in tropical Africa and the Neotropics. The trees are characterised by their monopodial form and yellow, white or cream exudates produced from cut boles and twigs (Figure 1.1), and are well known as fruit trees because most species have edible arillate fruits. One of the most famous tropical fruits from South East Asia, mangosteen (*G. mangostana* L.), which is sometimes dubbed as the “Queen of Fruit”, is the most widely known and cultivated species.



Figure 1.1 Exudates from *Garcinia* trees. A: *G. penangiana* Pierre; B: *G. discoidea* Nazre

Many taxonomic works on *Garcinia* are confined to restricted geographic areas, for example Peninsular Malaysia (Whitmore, 1973) or tropical Africa (Robson, 1961; Bamps et al., 1978). The last world-wide monograph is much older (Engler, 1893, 1925), and based upon other 19<sup>th</sup> century works (Pierre, 1882-1885; Vesque, 1893). Engler's monograph deals with a total of 95 species of *Garcinia* in 34 sections. The number of species since this account was published has increased nearly fourfold, to about 400 species as estimated by Whitmore (1973). However, I would estimate the number of species in *Garcinia* should be around 250 species considering the numerous names in the literature and new species yet to be described.

An unpublished PhD thesis by Jones (1980) proposed a new classification of the sections in *Garcinia*, and suggested that the genus should be divided into 14 sections based mainly on male flower characters. The incorporation of phylogenetic methodology and DNA sequence data in systematic studies allows the evaluation of taxonomic classifications using new methods and data. In *Garcinia*, preliminary molecular phylogenetic work has been started by Rismita-Sari (2000) to test Jones (1980) classifications, and these studies are extended in this thesis, with special emphasis on its main subject, the section *Garcinia*.

In Jones's (1980) classification, sect. *Garcinia* consisted of 43 species distributed in Madagascar, and India to Fiji. However, she stressed the need for a monographic revision of this section because of the poorly defined delimitation of some species. Because of the lack of information for the species on a world wide basis, Whitmore

(1973) left 10 *Garcinia* species unnamed in Peninsular Malaysia despite finding them morphologically different from other Malayan species. He specified that they may have been described elsewhere. Because of this, taxonomic revision in *Garcinia* should be done on a world wide monographic basis to resolve problem of species delimitation.

## **1.2 Project Aims and Objectives**

The core subjects of this project were to evaluate the monophyly of section *Garcinia* sensu Jones (1980) and to prepare a monographic account of section *Garcinia* which will clarify the presently poorly defined species boundaries. Molecular data together with morphological characters will be used to aid the classification and provide a better understanding of the phylogeny and evolution of the section. With this robust phylogeny and comprehensive taxonomic revision, it will be possible to evaluate global patterns and distribution of sect. *Garcinia* with clear concepts of sectional and species limits. It will also allow evaluation of the biogeographical history of sect. *Garcinia* which will contribute to our understanding of the historical construction of South East Asian rain forests.

In summary, the objectives of this study are;

- i. to investigate the monophyly of sect. *Garcinia*.
- ii. to prepare a taxonomic revision and produce detailed descriptions of morphology, accepted names in use and synonymy, to produce an

identification key and geographic distribution for each species in sect.

*Garcinia*.

- iii. to investigate the relationships, phylogeny and biogeography of the species within sect. *Garcinia*.

### **1.3 Thesis structure**

The thesis starts with a brief introduction and objectives of the study in the **Chapter 1**, while **Chapter 2** and **Chapter 3** will review the taxonomic history and the ecology and biology for sect. *Garcinia* from published literature and my own field observations. The morphological investigation is divided into **Chapter 4**, explaining the significance of morphological characters for sectional and species delimitation and **Chapter 6** will provide the morphological treatment for each species, including the distribution of sect. *Garcinia*. **Chapter 5** will cover the molecular systematic study, and the last chapter, **Chapter 7** will conclude the thesis and give some recommendations for further studies.

## **CHAPTER 2**

### **TAXONOMIC HISTORY**

#### **2.1 Introduction**

This chapter will review the taxonomic history of the classifications of the genus *Garcinia* and its sections.

#### **2.2 The Genus *Garcinia***

*Garcinia* honours Laurentius Garcin (1683-1752) who was a Dutch army doctor and naturalist in the Dutch Indies (Indonesia) between the years 1720-1729. During his voyage to the Moluccas (Maluku islands, Indonesia), he examined the fruit bearing tree which the locals called 'mangoustan' (mangosteen) and gave a description of the fruiting female specimen (Garcin, 1733). The species was named *Garcinia mangostana* by Linnaeus (1753), which became the type species for the genus. Mangosteen may be one of the earliest *Garcinia* species to be recorded by Europeans because the name 'mangostan' had been used by the Portuguese in India as mentioned in Garcia da Orta's book whose title translated as "Colloquies on the Simples, Drugs and Meteria Medica of India and some of the fruits found there" published in 1563 (Burkill, 1935). It was thought that the fruit was native in the Moluccas (Garcin, 1733) or Indonesia and Peninsular Malaysia (Garcin, 1733; Rumphius, 1741) but many suggested (e.g.

Whitmore, 1973; Richards, 1990c) that since mangosteen has never been found in the wild, it indicated that this is a hybrid species. The place of origin for mangosteen was suggested as Peninsular Malaysia because closely related species, *G. malaccensis* Hook f. and *G. celebica* L. (syn. *G. hombroniana* Pierre) are found there (Richards, 1990c). Mangosteen is believed to have been cultivated for hundreds of years in the South East Asia region (Ramage et al., 2004).

The family for *Garcinia*, Guttiferae, was created by Jussieu (1789) based on the presence of the exudates secreted from cut stems and leaves. However, Adanson (1763) was the first to correctly group *Garcinia* with the other guttiferous genera (*Calophyllum* L., *Cambogia* L., *Mesua* L. and *Mammea* L.) without providing a name. In addition, he also included the unrelated genera of *Fraxinus* L. (Oleaceae) and *Lawsonia* L. (Lythraceae).

Several years after Linnaeus' monumental work "Species Plantarum" where *Garcinia* had only one species, *G. mangostana*, two more species were added, each by Stickman (Linnaeus, 1754) and Murray (Linnaeus, 1774) specifically distinguished by shape of leaves and the inflorescences but sharing similar fruit characters, i.e. four sepals and petals, eight seeds and a peltate stigma. The number of species then gradually increased during the 19<sup>th</sup> century and some improvements were made to the definition of the genus by adding more characters, particularly from the flower, and also by the discovery of more new species, especially from the Far East. Examples are the works of Planchon & Triana (1860) who included characters from the embryo and seeds and

Bentham (1862) who added some characters from the inflorescence. Anderson (1874) introduced the inclusion of pentamerous flowers species as shown in *G. xanthochymus* although most *Garcinia* are tetramerous flower, and Baillon (1877) was the first to suggest the importance of separating the male and female characters.

Since then, species delimitation in *Garcinia* has always incorporated characters available from fruits, flowers or both. When Pierre (1882-1885) published his 'Forest Flora of Cochinchine' the species delimitation used many characters which included flowers (male and female), fruits and the latex colour. His species delimitation had a narrow concept with regard to the geographic distribution. For example, *G. celebica* (a name currently accepted in this study) was previously known by many synonyms created by Pierre such as *G. jawoera* (Java), *G. rumphii* (Bangka Is.), *G. hombroniana* (Peninsular Malaysia), *G. krawang* (Borneo), *G. kurzii* (Andaman) and *G. benthami* (Indo-china). The most important characters considered by Pierre (1882-1885) were the inflorescences, stamens and fruit characters. However, fruit characters together with some vegetative characters could also be used to recognise species, for example in *G. mangostana*. Pierre's species concept proved stable and was followed by subsequent monographic works such as Vesque (1893) and Engler (1893, 1925). Since Pierre, the importance of using both male and female (or fruit) characters has been consistently acknowledged. However, finding both male and female specimens of the same species is problematic. One solution, as adopted by Whitmore (1973), used species recognition from vegetative (leaf) characters and specific ecological distribution (e.g. montane or lowland). In the latest sectional revision by Jones (1980), no new characters or criteria for species

delimitation were suggested. In this thesis, the characters used for species delimitation are outlined in Chapter 4.

The International Plant Name Index (IPNI; <http://www.ipni.org>) has a total of 652 specific and subspecific names in *Garcinia* but the total number of accepted species for *Garcinia* is not entirely known. The latest estimation of 200 species was by Stevens (2001 onwards) which is smaller than the earlier estimates of 400 species by Whitmore (1973) or Jones (1980). These conflicting numbers reflect problems of poorly understood species in *Garcinia*. Many new species have been created since the last revision by Engler (1893, 1925) mainly from the publication of local geographic floras. These floras have contributed new knowledge on the species diversity of *Garcinia*, but because of their restricted geographic focus, may have resulted in the superfluous description of species that are not globally distinct. Examples of such floristic revisions of *Garcinia* are: Africa (Oliver, 1868; Robson, 1961; Bamps et al., 1978); India (Anderson, 1874; Hooker, 1875; Maheshwari, 1960); Myanmar (Kurz, 1877); Indo-china (Pierre, 1882-1885; Pitard, 1910; Pham-Hoang, 1991); Thailand (Craib, 1931); Peninsular Malaysia (King, 1891; Ridley, 1922; Whitmore, 1973); Indonesia (Miquel, 1859, 1860; Koorders & Valetton, 1903; Backer & Bakhuizen, 1963) and the Philippines (Merrill, 1908; 1923).

### 2.2.1 *Generic Limits*

During Linnaean time, many genera presently considered synonymous with *Garcinia* were described separately. Murray (Linnaeus, 1774) for instance, separated *Cambogia* by its short stamen filaments and fewer stigma rays, but it was later sunk into *Garcinia* by Choisy (1823).

When erecting the family Guttiferae, Jussieu (1789) divided it into three unnamed groups, and *Garcinia* was placed with *Cambogia*, *Clusia*, *Tovomita*, *Quapoya* and *Grias* into Group I, defined by lack of a style. Because of the dependence on fruit characters in *Garcinia* (rather than the flowers) for generic delimitation in the early years, several authors afterward erected many other genera in the family Guttiferae which they considered possibly related to *Garcinia*. The genera include *Stalagmitis* Murray (1789), *Oxycarpus* Loureiro (1890), *Brindonia* Thouars (1805), *Xanthochymus* Roxburgh (1798) *Hebradendron* Graham (1837) and *Discostigma* Hasskarl (1844). As commented by Jones (1980), during the period from 1789 to 1859, the relationship among these genera could not be established because use of the male flower was overshadowed by the ovary and fruit characters in suprageneric classifications.

*Stalagmitis* was recognised until 1861 when Planchon & Triana found that there were similarities of flowers with the specimens treated by Murray (1789) as *Garcinia* L. and *Xanthochymus* Roxb. However, Kurz (1870), when he observed *X. pinctoria* as having both pentamerous and tetramerous flowers suggested that *Xanthochymus* was no

different from other *Garcinia*. It was not until 1875 that Hooker (1875) who agreed with Kurz, sank *Xanthochymus* into *Garcinia*.

As for *Oxycarpus* Lour. and *Brindonia* Thouars, both were sunk into *Garcinia* by Choisy in 1824 under his section *Brindonia*. *Hebradendron cambogiodes* was the only species described by Graham (1837) when he erected this genus but it was reduced to a synonym of *G. morella* Desr. by Planchon & Triana (1860) who retained *Hebradendron* as the name for their new section.

Warburg (1891) erected *Pentaphalangium* based on having five stamen bundles (usually four in *Garcinia*) adnate to the petals, but it was sunk by Kostermans (1976). He considered the different numbers of stamen bundles in *Pentaphalangium* were part of the natural variation observed in *G. wareni*. A recent classification by Stevens (in Gustafsson et al., 2002) also included *Pentaphalangium* along with *Rheedia* and *Tripetalum* into *Garcinia*.

*Rheedia* created by Linnaeus (1753) is, however, not considered part of *Garcinia* by some authors, for example Berg (1979). This reflects its distribution, restricted to Africa and South America, and morphology, the filaments of the stamens being free rather than united as in *Garcinia*. Robson (1958) who did not agree with the stamen character argument, created new *Garcinia* species in Africa. This was also followed by Adams (1970), Borhidi (1980), Liogier (1986), Hammel (1989) and Zappi (1993)

although much earlier Vesque (1893) had already sunk a few *Rheedia* species from Madagascar into *Garcinia*.

*Tripetalum* was created by Schumann & Hollrung (1889) with only two species, one in Africa and another in New Guinea. Turner & Stevens (1999) proposed to transfer *Tripetalum cymosum*, a species found in New Guinea to *Garcinia*. Jones (1980), who revised the sectional classifications and generic limits in *Garcinia*, proposed several changes. She considered that *Garcinia* should include, 1) seven species of *Ochrocarpus* because of their 2-lobed globose anthers, 2) a monospecific *Septogarcinia* Kostermans (1962) which only differs by dehiscent and ribbed fruit, and 3) all species in *Rheedia* by sharing a similar pollen type and free stamens as in *Garcinia*. Jones's suggestion was tested by nuclear ribosomal internal transcribed spacer (ITS) sequence data by Rismita-Sari (2000) which mostly agreed with the unification of the genera mentioned but she further proposed to elevate several sections into four subgenera. As neither work has been published these names changes remain invalid. Jones's sectional classification is being reviewed by Patrick Sweeney (Missouri Botanical Gardens) using morphological and molecular data (pers. comm.) Recent work, including data taken from molecular analysis (Gustafsson et al., 2002), also shows *Garcinia* together with *Pentaphalangium*, *Rheedia* and *Tripetalum* are members of the same monophyletic clade.

### 2.2.2 *Infrageneric Classification of Garcinia*

In the early period of infrageneric classification in *Garcinia*, several disagreements arose because of lack of materials and difficulties in communication. The first effort to establish an infrageneric classification in *Garcinia* was made by Choisy (1823 & 1824), who established two sections, *Mangostana* and *Brindonia*, based on stamen and flower characters (Table 2.1) to account for the nine species then known. However, when more species were found (30 species, Choisy 1851) he then changed this classification by dividing the species into two subgenera where most of the species he had described earlier were grouped together into subgenus I while the rest were placed in subgenus II.

By contrast, Blume (1825) put six species of *Garcinia* found in the Dutch East Indies (Indonesia) into two sections, *Cladogynos* and an unnamed section (*G. mangostana* and related species), based on characters of the stigma on the fruits. Wight (1840), on the other hand, suggested that Indian *Garcinia* should be divided into two subgenera, *Mangostana* and *Oxycarpus* and then further separated into sections by the characters of the anthers. He could not, however, place some of the species he took from Roxburgh's *Hortus Bengalensis* (1814) and Wallich's catalogue (1828-1849).

Planchon & Triana (1860) then published their own classification based on characters of male flowers and supported by the female and fruit characters. *Garcinia* was divided into six sections and many problematic species were resolved. Lanessan (1872)

did not agree with Planchon & Triana, claiming they used too many characters for infrageneric classification. He created five unnamed subgenera based on the characters of the anthers. Only subgenus I was divided into four other sections (Table 2.1).

In his work 'Forest Flora of Cochinchine', Pierre (1882-1885) described 149 *Garcinia* species, many of which were new, and placed these species within 38 sections based mainly on characters of the inflorescences and the anthers. His treatment, however, covered the species not only found in the Indo-china region but also as far as from Africa and New Caledonia, and provided the basis for the next works in the classification of *Garcinia*. He divided the sections into six groups based on the characters of the anthers but did not formally give any taxonomic ranks to these groups.

Vesque's (1893) work is considered to be the first monograph for *Garcinia*. It did not entirely agree with Pierre's sectional classification but concurred on many of Pierre's species. It divided *Garcinia* into three subgenera and nine sections based on flower characters and, in addition, he included stomatal anatomy to justify his classification based upon his anatomical studies (Vesque, 1889) on family Guttiferae. However, the most significant classification for *Garcinia* was by Engler (1893), in which he retained most of Pierre's sectional classification. In Engler's final account (Engler, 1925), *Garcinia* was divided into 34 sections based mainly on characters of the anthers and stamens (Table 2.3).

No major taxonomic work was published after Engler's treatment except for the floristic accounts mentioned above. The most recent revision of *Garcinia* was by Jones (1980) who incorporated a greater range of characters from the inflorescences, flowers, pistillode, fruit and vegetative features. As a result, she divided *Garcinia* into 14 sections (see table 2.2). The important characters were from the male flowers, i.e. the arrangement of the stamens bundles and the presence of the pistillode.

Table 2.1. Historical events in the infrageneric classification of genus *Garcinia*.

Author	Infrageneric	Characters used
Choisy (1823 & 1824)	1. Sect. <i>Mangostana</i> 2. Sect. <i>Brindonia</i>	Flowers monoecious or hermaphrodite & stamens
Blume (1825)	Sect. I. (Unnamed) Sect. II. <i>Cladogynos</i>	Stigma
Wight (1840)	1. Subgenera <i>Mangostana</i> 2. Subgenera <i>Oxycarpus</i>	Anthers
Choisy (1851)	1. Subgenus Primum 2. Subgenus Secundum	Inflorescences: position & flower numbers
Planchon & Triana (1860)	I. Sect. <i>Mangostana</i> II. Sect. <i>Peltostigma</i> III. Sect. <i>Cambogia</i> IV. Sect. <i>Trachycarpus</i> V. Sect. <i>Comarostigma</i> VI. Sect. <i>Hebradendron</i>	Stamen, anther, fruit, stigma, staminodes, ovary
Lanessan (1872)	Subgenera 1 I. Sect. <i>Mangostana</i> II. Sect. <i>Peltostigma</i> III. Sect. <i>Echinocarpa</i> IV. Sect. <i>Cyclandreae</i> V. Sect. <i>Synadelphandreae</i> Subgenera 2 Subgenera 3 Subgenera 4 Subgenera 5	Male: stamens, pistillode, anther; female: stigma, ovary
Hook f. (1875)	Subgenus I. <i>Garcinia</i> Subgenus II. <i>Xanthochymus</i>	Number of sepals & petals

Author	Infrageneric	Characters used
Pierre (1882-1885)	Group 1 – Sects.: I. <i>Teracentrum</i> II. <i>Rheediopsis</i> III. <i>Xanthochymus</i> IV. <i>Discostigma</i> V. <i>Dicrananthera</i> VI. <i>Plinthostigma</i> VII. <i>Mucestigma</i> Group 2 – Sects.: VIII. <i>Depastigma</i> IX. <i>Echinostigma</i> X. <i>Peltostigma</i> XI. <i>Gamodesmis</i> XII. <i>Conocentrum</i> XIII. <i>Sphaerocentrum</i> XIV. <i>Mangostana</i> XV. <i>Gynegonia</i> XVI. <i>Kiras</i> XVII. <i>Tagmanthera</i> XVIII. <i>Collanthera</i> XIX. <i>Stemmandra</i> XX. <i>Simosanthera</i> XXI. <i>Brindonia</i> XXII. <i>Tetradium</i> XXIII. <i>Pachyphyllum</i> XXIV. <i>Papilla</i> XXV. <i>Cambogia</i> XXVI. <i>Echinocarpa</i> XXVII. <i>Cladogynos</i> XXVIII. <i>Anisostigma</i> XXIX. <i>Olostigma</i> Group 3 – Sects.: XXX. <i>Comarostigma</i> XXXI. <i>Didymadenia</i> XXXII. <i>Rhinostigma</i> XXXIII. <i>Tetraclinia</i> XXXIV. <i>Oxycarpus</i> Group 4 – Sect.: XXXV. <i>Hebradendron</i> Group 5 – Sect.: XXXVI. <i>Ripidionanthera</i> Group 6 – Sect.: XXXVII. <i>Daedalanthera</i> XXXVIII. <i>Campylanthera</i>	Inflorescences, anther
Vesque (1893)	1. Subgenus <i>Xanthochymus</i> 1. Sect. <i>Xanthochymus</i> 2. Subgenus <i>Rheediopsis</i>	Inflorescences, sepals and petals, stamen, anther, pistillode, ovary, stigma,

Author	Infrageneric	Characters used
	2. Sect. <i>Rheediopsis</i> 3. Sect. <i>Teracentrum</i> Subgenus <i>Eugarcinia</i> 4. Sect. <i>Mangostana</i> 5. Sect. <i>Cambogia</i> 6. Sect. <i>Oxycarpus</i> 7. Sect. <i>Rhinostigma</i> 8. Sect. <i>Hebradendron</i> 9. Sect. <i>Paragarcinia</i>	stomata
King (1891)	Subgenus 1 <i>Garcinia</i> Sect. 1 Sect. 2 Sect. 3 Sect. 4 Sect. 5 Subgenus 2 <i>Xanthochymus</i>	Sepals & petals; stamen, anther, pistillode

Jones grouped her sections into three (groups 1 to 3) based on the types of the anthers but did not give any ranks to these groups (Table 2.2). The groups are (1) two thecous anthers, ellipsoid, recurved with a central connective: sect. I, II, III, IV, V, VI and VII, (2) four thecous anthers: sect. VIII, IX and X, (3) two thecous anthers, globose and apical slit: sect. XI, XII, XIII, XIV. Within group 1, three subgroups were created (A-C) and within group 3, two more subgroups were created (A and B) based on floral and fruit characters. She also did not give any ranks for subgroups because she believed that the purpose of classification is practical to identify the species, and these groups, are difficult to recognise using herbarium specimens, especially when sterile. Jones's approach of creating the groups with numerical numbers for the sections were influenced by the work of Whitmore (1973) although she felt that relationships could be expressed at several levels and the characters could be used both for herbarium and living material.

Table 2.2. Groups, subgroups and sections established by Jones (1980).

<i>SECTION</i>	<i>CHARACTERS</i>
<b>Group 1</b>	
A. I. <i>Garcinia</i>	Inflorescences various; flower tetramerous (rarely pentamerous); stamens numerous in 4-bundles or in annular mass; anthers 2-theous, ellipsoid; pistillode present or absent; fasciclodes present or absent; staminodes present or absent; ovary 2-10 locules; stigma entire or lobed; fruit various, sometimes with woody wall.
B. II. <i>Rheediopsis</i> III. <i>Teracentrum</i> IV. <i>Rheedia</i>	Inflorescences of cauline axillary glomerules; flower tetramerous except in <i>Rheedia</i> ; stamens fewer than 50, partially united in 4-bundles or free; anthers 2-theous, ellipsoid; fasciclodes present; pistillode absent; ovary 2-4 locules; stigma entire or weakly lobed; fruit skin thin, smooth.
C. V. <i>Macrostigma</i> VI. <i>Tetraphalangium</i> VII. <i>Tripetalum</i>	Inflorescences terminal; flower pentamerous, tetramerous or trimerous; stamens numerous in bundles adnate to the petals; anthers 2-theous, ellipsoid; fasciclodes present or absent; pistillode present or absent; ovary 3-5 locules; stigma lobed; fruit skin thin, smooth.
<b>Group 2</b> VIII. <i>Brindonia</i> IX. <i>Mungotia</i> X. <i>Hebradendron</i>	Inflorescences axillary or terminal or both; flower tetramerous; stamens in a single mass; anthers 4 or 2-theous, erect or peltate; fasciclodes absent; ovary 2-15 locules; stigma in rays, glandular; fruit skin thin, smooth, warty or echinate, sometimes grooved.
<b>Group 3</b>	
A. XI. <i>Xanthochymus</i> XII. <i>Paragarcinia</i>	Inflorescences axillary or terminal or both; flower pentamerous or tetramerous except calyx undivided before anthesis in <i>Paragarcinia</i> ; stamens partially united in bundles; anthers 2-theous, globose; fasciclodes present; pistillode absent; ovary 4-5 locules; stigma lobed, smooth; fruit skin thin or with slightly woody wall, smooth or rough
B. XIII. <i>Discostigma</i> XIV. <i>Tagmanthera</i>	Inflorescences axillary or terminal or both; flower tetramerous; stamens united in 4, rarely 2, strap-shaped bundles; anthers 2-theous, globose; fasciclodes absent; pistillode present; ovary 1-4 locules; stigma entire, smooth; fruit skin thin or with slightly woody wall, smooth.

Table 2.3. Comparison of the section names used by Jones, Engler & Pierre.

Jones (1980)	Engler (1925)	Pierre (1882-1885)
I – <i>Garcinia</i>	<i>Mangostana</i> p.p. <i>Peltostigma</i> <i>Anisostigma</i> p.p. <i>Conocentrum</i> <i>Daedalanthera</i> p.p. <i>Echinostigma</i> p.p. <i>Gamodesmis</i> <i>Simosanthera</i>	<i>Kiras</i>       <i>Sphaerocentrum</i>
II – <i>Rheediopsis</i>	<i>Stemandra</i> <i>Rheediopsis</i>	
III – <i>Teracentrum</i>	<i>Teracentrum</i>	
IV – <i>Rheedia</i>	-	
V – <i>Macrostigma</i>		<i>Macrostigma</i>
VI – <i>Tetraphalangium</i>	<i>Tetraphalangium</i>	
VI - <i>Tripetalum</i>	-	
VIII – <i>Brindonia</i>	<i>Cambogia</i> <i>Comarostigma</i> <i>Echinocarpa</i> [ <i>Anisostigma</i> p.p.] <i>Didymadenia</i> <i>Holostigma</i> <i>Depastigma</i> <i>Oxycarpus</i> <i>Rhinostigma</i>	<i>Brindonia</i> <i>Tetradium</i> <i>Papilla</i> <i>Cladogynos</i>
IX – <i>Mungotia</i>	<i>Tetrachoristemon</i> <i>Mungotia</i> [ <i>Daedalanthera</i> p.p.] <i>Tetraclinia</i> [ <i>Mangostana</i> p.p.]	[ <i>Gynegonia</i> p.p.] <i>Pachyphyllum</i>
X – <i>Hebradendron</i>	<i>Hebradendron</i> [ <i>Daedalanthera</i> p.p.]	
XI – <i>Xanthochymus</i>	<i>Xanthochymus</i> <i>Xanthochymopsis</i>	
XII – <i>Paragarcinia</i>	<i>Paragarcinia</i>	
XIII – <i>Discostigma</i>	<i>Discostigma</i>	
Subsect. I – <i>Discostigma</i>	<i>Campylanthera</i> <i>Mucestigma</i>	

Jones (1980)	Engler (1925)	Pierre (1882-1885)
		<i>Plinthostigma</i>
	[ <i>Mangostana</i> p.p.]	
Subsect. II– <i>Dicrananthera</i>	<i>Dicrananthera</i>	
XIV – <i>Tagmanthera</i>		<i>Tagmanthera</i>
Subsect. I – <i>Tagmanthera</i>		<i>Collanthera</i>
Subsect. II - <i>Bifasciculata</i>		

### 2.3 Section *Garcinia* sensu Jones

Section *Garcinia* is among the biggest sections in *Garcinia* with a total of 43 species (Appendix I), and is typified by *G. mangostana* L. Jones considered sect. *Garcinia* as the most heterogeneous section of *Garcinia* based on the pollen types, but the key character that united species into this section was the stamens united in four bundles or in an annular mass, usually around the central pistillode. By using this key character, Jones (1980) united 11 sections previously described by Pierre (1882-1885), Vesque (1893) and Engler (1893, 1925) into sect. *Garcinia* (Table 2.3). In sect. *Garcinia*, she stressed the need for taxonomic revision and defining the limits of the species. Apart from revising species in sect. *Garcinia*, this study will also test the sectional circumscription of sect. *Garcinia* as established by Jones.

On the relationships and evolution of sect. *Garcinia*, Jones (1980) was more speculative because not all species had been treated in depth for character variation. By the characters of the 2-theous, ellipsoid anthers and basic tricolporate pollen, sect. *Garcinia* was said to be closely related with sect. II – VII, but from the vegetative

morphology, it resembles members from sect. II, III and IV (Jones, 1980). However, this thesis will not cover morphological evolution and relationship of the sections because it is being studied by Patrick Sweeney (Missouri Botanical Garden).

The distribution of sect. *Garcinia* is centralised in the Malesian region, and goes up to the north-west of Indo-China, Myanmar, eastern India with one species in Madagascar. To the east, with the inclusion of *G. pseudo-guttiferae*, the distribution of sect. *Garcinia* reaches the Fijian and Tonga islands (Table 2.4). In total, there were eight species studied by Jones and in addition, she listed 35 more species (excluding three invalid names) that should be included in sect. *Garcinia* (Appendix I).

Table 2.4. Species number and geographical distribution of 14 sections of *Garcinia* sensu Jones (1980).

Section	Number of species	Distribution
I. <i>Garcinia</i>	43	Madagascar, India, Indo-china, Malesia, New Guinea & Fiji.
II. <i>Rheediopsis</i>	20	Tropical Africa.
III. <i>Teracentrum</i>	4	Tropical Africa.
IV. <i>Rheedia</i>	30	Central & South America, Madagascar.
V. <i>Macrostigma</i>	10	Java, Borneo, N. Guinea, Philippines, Caroline Is.
VI. <i>Tetraphalangium</i>	2	Central & West tropical Africa.
VII. <i>Tripetalum</i>	1	New Guinea.
VIII. <i>Brindonia</i>	77	India, Sri Lanka, Indo-china, P. Malaysia, New Guinea, Philippines.
IX. <i>Mungotia</i>	8	Fiji Is. & New Caledonia.
X. <i>Hebradendron</i>	40	India, Indo-china, Thailand, P. Malaysia, Java, Philippines & N. Guinea.
XI. <i>Xanthochymus</i>	42	Trop. Africa, Madagascar, India, Nepal, S.E. Asia & N.
XII. <i>Paragarcinia</i>	10	Guinea.
XIII. <i>Discostigma</i>	54	Madagascar.
XIV. <i>Tagmanthera</i>	18	Tropical Africa, India, S.E. Asia, Fiji & Tonga Is. & Taiwan

## CHAPTER 3

### THE ECOLOGY AND BIOLOGY OF SECTION *GARCINIA*

#### 3.1 Introduction

Information on the ecology and biology of *Garcinia* is scarce as only a few species have been studied. The cultivated species *G. mangostana* (mangosteen) is an exception, as it has been extensively studied for a long time because of the economic importance of the fruit. The biology of mangosteen has been investigated since the early 20<sup>th</sup> century (e.g. Treub, 1911; Sprecher, 1919; Hume & Cobin, 1946) with much recent work (e.g. Lim, 1984; Verheij, 1991; Normah, 2000; Cruz, 2001). This chapter will, however, try to place more emphasis on species from the wild. Scientific names follow the taxonomic treatment in chapter 6. This has resulted in some nomenclatural changes in comparison with previously published work. For example, *G. malaccensis* Hook. f. in Pasoh Forest Reserve is a misidentification; the correct name is *G. penangiiana* Pierre. However, some old synonymous names are provided when necessary.

#### 3.2 Habit

Members of sect. *Garcinia* are generally small to medium sized trees reaching 10-35 cm diameter at breast height (dbh) and 5-25 m tall when mature, and rarely reaching 50 cm dbh and 30 m tall. This means that most *Garcinia* trees are found in the forest

under-storey though a few species can reach the main canopy (Appanah & Weinland, 1993). In the tropical forest, under-storey trees do not generally exceed 20 m tall while main canopy trees do not exceed 30 m tall (Kochummen et al., 1990).

An example of an under-storey tree from sect. *Garcinia* is *G. penangiana*, one of the most common species distributed in Sumatra, Peninsular Malaysia and Borneo. Studies in the 50-ha plot of lowland forest in Pasoh Forest Reserve show that of 899 individuals of *G. penangiana*, only three reached 15-20 cm dbh (Nazre, 2000). A similar pattern of diameter was observed in the 52-ha plot in Sarawak where only four of 47 trees of *G. penangiana* reached 10-19 cm dbh (Lee et al., 2002).

Nevertheless, a few species of sect. *Garcinia* in Malaysia, for example *G. celebica* L, *G. malaccensis* Hook f. and *G. nitida* Pierre, have been recorded to reach more than 50 cm dbh and may be logged for timber. In Malaysia, timber of *Garcinia* species is classified based on their common names, and is known as 'Kandis', a medium hardwood which is said to be moderately durable and difficult to work. Whitmore (1973) claimed that the woods of *Garcinia* are not considered to be of commercial value because of the few species that reach desirable timber size in comparison with other timber groups, e.g. the Dipterocarps. Other species in sect. *Garcinia*, namely *G. harmandii* Pierre, *G. cataractalis* Whitmore and *G. diospyrifolia* var. *minor* Nazre var. nov. are bushy trees with dense crowns and *G. diospyrifolia* var. *diospyrifolia* Nazre var. nov. is small and of slender stature.

The crown of *Garcinia* trees was described by Whitmore (1973) as monopodial, dense and conical (pagoda like). This growth form is quite common in tropical forest, and found in families such as Annonaceae or Myristicaceae, but *Garcinia* can easily be differentiated from these groups by its opposite leaves. Monopodial branching is a growth model in which secondary shoots or branches arise behind the growing point but remain subsidiary to the main stem, which continues to grow indefinitely (Tootil, 1984).

### **3.3 Distribution and Dispersal**

*Garcinia* trees are almost always confined to the interior of the forest or to the shade (Whitmore, 1973). Even species commonly found in coastal forest such as *G. celebica* are shade tolerant and do not grow in the open sunlight but rather among other species within the coastal forest. Ecologically, most species of sect. *Garcinia*, such as *G. penangiana*, *G. malaccensis* and *G. diospyrifolia* are found in the lowland or hill forest (less than 900 m altitude) but some other *Garcinia* species, such as *G. clusiaefolia* are restricted to montane areas (Whitmore, 1973). *Garcinia celebica*, the most widely distributed species of sect. *Garcinia*, found from South East Asia to India, can be present in different ecological and altitudinal conditions from lowland to montane forest, though it is found more commonly in coastal forest. Another species in sect. *Garcinia* that is found at higher altitude (to 1200 m) is *G. sangudsangud*, while *G. cataractalis* is the only rheophyte, restricted to wet riverine areas.

In lowland forest of Peninsular Malaysia, studies show that half of the 14 species of *Garcinia* in Pasoh Forest Reserve have a high preference for well-drained alluvial soil on flat land rather than on hillsides (Nazre, 2000). Pasoh forest is classified as mixed Dipterocarp forest with two relatively dry periods each year and annual rainfall is below any other tropical rain forest near the equator (Kochummen et al., 1990; Wolseley et al., 1996). The two common species of sect. *Garcinia* found in Pasoh, *G. penangiana* and *G. diospyrifolia* var. *minor* strongly follow the distribution pattern of preferring well-drained and flat land. It is, however, not fully understood how other factors such as soil nutrients affect the distribution of *Garcinia* species though it is known that the composition of the rain forest flora varies dramatically in relation to soil nutrients (Ashton, 1988c).

Geographically, sect. *Garcinia* is as defined in this thesis distributed in the South East Asian region though it spreads to the east to New Guinea while to the west, it reaches the east of India (Figure 3.1). The number of species shows a significant decline towards the east and west. For example, across Wallace's Line to the east, three species are found (*G. celebica*, *G. rigida*, *G. ochraceus*) while to the west of Wallace's Line, a total of 15 species of sect. *Garcinia* are found. Species numbers again decrease as it crosses Indo-china (3 species) towards India (1 species).

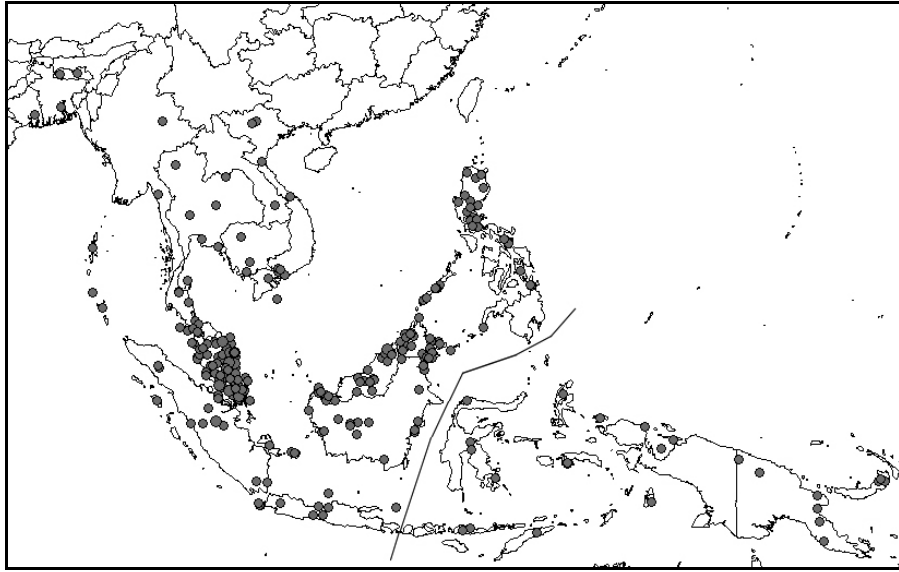


Figure 3.1. Worldwide distribution of sect. *Garcinia*, showing Wallace's Line

### 3.4 Floral and Fruit Biology

The approximate period of time needed for *Garcinia* trees to reach maturity and first start to produce fruits or flowers is not known except in mangosteen. Based on cultivated trees of mangosteen (Lim, 1984), *Garcinia* species are thought to be slow-growing species and take probably around 10-15 years to start producing flowers.

Each species has a different and irregular flowering pattern. For example, one of the most common *Garcinia* (also sect. *Garcinia*) species in Peninsular Malaysia, *G. penangiana*, flowers only once in a three year period (Ha et al., 1988). This is a perfectly normal pattern for tropical rain forest trees (Whitmore, 1975; Appanah, 1985) and is one reason why *Garcinia* is a difficult subject for study of floral and fruit biology. Richards

(1990b), claimed that *G. celebica* (syn. *G. hombroniana*) is able to flower throughout the year with a peak of flowering from January to June but this observation was based on cultivated trees planted in parks and arboreta where growing conditions differ from natural forest. When comparison was made between cultivated and natural populations of *G. celebica* by Richards (1990b), he found that there were no correlations in flowering time.

During flowering, as observed in *G. celebica* (Richards, 1990b), the anthesis of female flowers occurs when the petals start to reflex and is followed by the loss of the petals. The sepals persist even when the fruit is mature, and the time from flowering to fruit maturation is between 100 and 110 days (Richards, 1990b). For the male flowers, no petals are lost when flowers reach maturity, and the flowering time is much longer than the female (Richards, 1990b). This pattern is typical for most dioecious plants (Bawa, 1980). For many *Garcinia* species, male inflorescences usually contain more flowers than female inflorescences. For example, in *G. celebica* up to 16 flowers are found in male inflorescences but usually solitary, paired or triad flowers in female inflorescences. These factors are considered to increase the chances of sexual pollination. Morphologically, the male flowers are smaller. Thomas (1997) and Ha et al. (1988) found that *G. scortechinii* (non sect. *Garcinia*) has a female based-ratio with only one male tree recorded in Peninsular Malaysia. The same pattern is observed in *G. penangiana* where one population had 31 males compared to 48 females (Thomas, 1997).

The data on pollinators of *Garcinia* is limited but generally for dioecious species, bees are the most common pollinators in tropical rain forest (Renner & Feil, 1993). Bees of the genus *Trigona* may be pollinators as observed in *G. celebica* (Richards, 1990b), attracted to the flowers by their nectar and pollen. The colour of the petals might also help to attract pollinators and in sect. *Garcinia*, species such as *G. diospyrifolia* and *G. malaccensis* have red or pinkish petals. However, in the absence of the pollinators, *Garcinia* may survive and successfully reproduce because of its capacity to produce fertile seed from an asexual breeding system known as agamospermy (see below for more discussion).

Fruits produced in *Garcinia* are edible in almost all of the species. The edible part is the pulp, usually white in colour, surrounding the seeds and protected by an indehiscent fruit wall. In the Indo-Malayan forest, fruit such as this is mostly consumed by primates (Leighton & Leighton, 1983). Some other small mammals might also be attracted to *Garcinia* fruits because the fruits are generally green and yellow at maturity (Corlett, 1993). Fruits eaten largely by mammals and primates are typically duller-coloured than fruits taken mostly by birds (Leighton & Leighton, 1983; Lambert & Marshall, 1991) because mammals and primates have special visual adaptation to make it easier for them to detect fruits against a background of leaves (Osario & Vorobyev, 1996). Some species, however, such as *G. celebica*, *G. diospyrifolia* and *G. nitida*, produce an attractive bright red fruit which might attract large birds such as Bucerotidae (Hornbill). Corlett (1993) also reported that *Garcinia* fruit had been found to be consumed by species of lizards. In

general, the dispersal agents for *Garcinia* are most likely primates and other small mammals such as Tupaiidae (tree shrews).

The germination period for different species is variable. From three species of *Garcinia* studied in Pasoh Forest Reserve, Ha et al. (1988) found that the germination of the seed of *Garcinia* was variable and slow, taking from six to eight months and that nearly half of the seeds were sterile. In total, three species from sect. *Garcinia* have been observed; in the well studied *G. mangostana*, germination took 17-64 days; 55-189 days for *G. diospyrifolia*; and 14-35 days for *G. penangiana* (Ng, 1992). Germination is hypogeal where the shoots develop from one end of the seed, and the primary root from the other end, and the primary root is replaced by adventitious roots arising from the base of the shoot (Ng, 1992).

### **3.5 Agamospermy and *Garcinia***

Agamospermy is a type of apomixis in which seed production is by asexual processes where fertilisation is not required for the normal development of fruit and seed (Gustafson, 1946-1947). Some authors use the term apomixis instead of agamospermy as they consider both synonymous (Richards, 1983), while others use apomixis to describe all forms of asexual reproduction in plants.

Apomixis is considered as a strategy for plant evolutionary success and survival because it maintains genetic heterozygosity inherited from ancestral outcrossers (Allem, 2004). There are two types of agamospermy, obligate and facultative. Obligate agamospermy is rare (Asker & Jerling, 1992), such as in subarctic Poaceae and Cyperaceae (Richards, 2003), perhaps because species capable of facultative agamospermy can increase their heterozygosity by sexual outbreeding. In angiosperms, the change to apomixis is probably influenced by genetics, physiological and environment factors (Clausen, 1954; Allem, 2004).

In theory, agamospermous progenies will have similar morphology to their mother trees and can be regarded as 'seed clones' (Richards, 2003). If one (or more) siblings reproduce sexually, the progeny will show some variation via increased heterozygosity. The role of agamospermy in *Garcinia* is not fully understood as breeding and cytology have been studied in insufficient species. In *G. celebica*, Richards (1990b) observed that two populations, coastal and montane, varied in the size of the pollen, with the coastal populations having a smaller size pollen grain. It is unclear whether this variation reflects agamospermy or environmental factors.

In temperate regions, agamospermy is a common and well-studied phenomenon, for example in *Hieracium*, *Taraxacum* and *Rubus* (Richards, 2003). In contrast, it is poorly studied in tropical forest tree species. Agamospermy has been long known in the tropics, but the first observations were on fruit trees such as mangosteen (e.g. Treub,

1911). Kaur et al. (1978) postulated that agamospermy might be widespread in the tropical rain forest based on findings in Dipterocarp species. Carman (1978) recorded 33 agamospermous angiosperm families and Allem (2004) increased this number to 86. Richards (1983) estimated about 330 plant genera are agamospermous. Few data are available for tree genera in Asian tropical rain forest that are capable of agamospermy, but confirmed agamosperms include *Garcinia* and *Calophyllum* (Guttiferae), *Lansium* (Meliaceae), *Syzygium* (Myrtaceae), *Diospyros* (Ebenaceae), *Memecylon* (Melastomataceae), *Mangifera* (Anacardiaceae), and *Shorea* and *Hopea* (Dipterocarpaceae) (Ashton, 1988b; Richards, 1990a; Lughadha & Proenca, 1996; Allem, 2004).

*Garcinia* is the only agamospermous genus in which both sporophytic agamospermy (adventitious embryony) and gametophytic agamospermy (probably a mitotic diplospory) coexist, having evolved independently within the same genus (Ha et al., 1988). In the presence of male plants, a substantial proportion of sexual seed formation will occur in most species (facultative agamospermy) and only in *G. mangostana* has it been suggested that there is obligate agamospermy where males are always absent (Richards, 1990a). In order to test the occurrence of agamospermy in *Garcinia*, simple experiments have been done by bagging the flower buds at a similar stage of development either with the anthers (if present) removed or not (Lim, 1984). Another method involves monitoring the embryological development when no viable pollen is produced and the embryo is non-zygotic (Ha et al., 1988; Soepadmo, 1989).

However, for *Garcinia*, Richards (1990a) outlined five criteria to determine presence of agamospermy; a) the occurrence of germinable seed in the absence of pollen or after bagging flowers; b) the precocious development of the embryo, i.e. before anthesis; c) the occurrence of adventitious proembryos, budding vegetatively from nucellar or integumental tissues; d) multiple seedling production from a single seed; e) a rarity or absence of males. Following these indications, Richards (1990a) listed 10 species of *Garcinia* that he considered agamospermous by possessing either one or more of these criteria. Of these, three species are in sect. *Garcinia* and their embryology has been studied: *G. mangostana* (Lim, 1984; Richards, 1990a), *G. penangiana* (mistakenly identified as *G. malaccensis*; Ha *et al.*, 1988) and *G. celebica* (Richards 1990a; 1990b).

Using Richard's final criterion, Thomas (1997) suggested that in *Garcinia*, if a species shows a female biased sex ratio, it is probably a facultative agamosperm. If this assumption is a good indication of the presence of agamospermy, then nearly all species in sect. *Garcinia* may be agamospermous. This is based on the observation of the herbarium specimens in this study which shows female-biased ratios for example in *G. malaccensis* (4 male out of 30 collections), *G. diospyrifolia* (3 male out of 33 collections) and *G. nitida* (2 male out of 13 collections). However, Richards (1990a) stressed that more information needs to be gathered to discover whether low male ratios are truly clear indications of facultative agamospermy.

To conclude, more study, especially of embryology, will be needed to confirm the

presence or type of agamospermy in *Garcinia* and it would be premature to claim the whole genus is agamospermous. The situation may be similar to *Shorea* (Dipterocarpaceae; Kaur et al., 1978; Chan, 1981; Bawa, 1998; Ng et al., 2004) where only some species are agamospermous.

### 3.6 Chromosome numbers

Basic cytological information is fundamental to the understanding of genetic variation and breeding systems of plant species (Soepadmo, 1989) but *Garcinia* species have been poorly studied, the latest papers being by Richards (1990b). Table 3.1 summarises the diploid numbers of chromosomes that have been reported in sect. *Garcinia*. The exact ploidy level of *Garcinia* is uncertain but Richards (1990c) claimed that *G. mangostana* is probably allopolyploid with respect to its related taxa *G. hombroniana* (= *G. celebica*) and *G. malaccensis* (misidentification of *G. penangiana*). It is, however, sufficient to say that *Garcinia* in general contains complex polyploid series.

*Garcinia* species have very small chromosomes and most studies have been based on male meiosis (except *G. mangostana*). Because of the small size of the chromosomes, variable numbers have been reported possibly due to inaccurate counts (Richards, 1990c). Even in the widely studied *G. mangostana*, the diploid number ranges from 56 to 96. If the data presented are correct, chromosomal instability may occur and directly contribute to morphological variation within the same species (Ramage et al., 2004). However, more data will be needed to verify this claim especially in relation to morphological change.

Table 3.1. Chromosomes numbers of some *Garcinia* species.

Species	2n	Source
1. <i>G. celebica</i>		
As <i>G. benthami</i>	48	Tixier (1990b; 1990c; 1990a)
As <i>G. hombroniana</i>	48	Richards (1990b)
As <i>G. speciosa</i>	c. 55	Krishnaswamy & Raman (1949)
2. <i>G. penangiana</i>	42-43	Ha et al. (1988)
3. <i>G. mangostana</i>	c. 76	Krishnaswamy & Raman (1949)
	96	Tixier (1960)
	88-90	Ha et al. (1988)
	60	Kaur et. al (1978)
	56-76	Soepadmo (1989)
4. <i>G. diospyrifolia</i>	68	Soepadmo (1989)

### 3.7 Ethnobotanical Uses

In Asia, *Garcinia* species are widely known as fruit trees and in sect. *Garcinia*, the most widely cultivated species is *G. mangostana*, in which the fleshy portion of the aril that is eaten is estimated to be about 25-30% of the fruit weight (Cruz, 2001). It can be eaten fresh or boiled together with the seeds and sugar added, or the pulp can be made into juice, syrup or jam. Other than mangosteen, some people in Peninsular Malaysia also plant *G. celebica* for its fruits (Burkill, 1935; Whitmore, 1973).

Some other wild *Garcinia* might have potential as fruit trees (Whitmore, 1973) but this is quite difficult to assess given the lack of published studies. Notes on herbarium specimens provide some information on the taste of the fruits. For instance, *G. cataractalis*, *G. malaccensis*, *G. nitida* and *G. diospyrifolia* have been recorded as having

a sweet and sour taste similar to mangosteen. *Garcinia* also has horticultural potential, for example *G. celebica* is now planted in Malaysia as a roadside plant or in gardens for its beautiful crown shape and colour. *G. diospyrifolia* might have potential for the same purpose and has attractive reddish flowers and fruits.

As a medium hardwood, *Garcinia* wood is suitable for poles, building bridges, flooring and pallets. It is, however, not known which species are used for construction but Watt (1890) and Burkill (1935) claimed that species that reached large size for example, *G. malaccensis* and *G. celebica* were used to build houses and fences in India and Peninsular Malaysia. Whitmore (1973) stated that the wood of *Garcinia* is not generally suitable for timber although some species can be made into poles or smaller items such as instruments.

The most significant ethnobotanical use of *Garcinia*, other than as fruit, is as traditional medicine. In Peninsular Malaysia, Burkill (1935) documented the use of some *Garcinia* species such as *G. celebica* and *G. mangostana* where parts of the trees including bark, roots and leaves were made into a decoction to cure menstrual problems, as medicines after childbirth, for relieving dysentery, fever, itchiness and wounds. Other minor uses include gum-resin (gamboge), tannin and dye.

### 3.8 Natural Products and Pharmacological Potential

Over the years, research to find natural products in the species of sect. *Garcinia* has increased. Between the years 2000 and 2006, 13 publications of natural product research were published compared to seven before the year 2000. Natural products are chemical compounds, usually secondary metabolites, found in plants with a variety of functions, such as defence against fungi and bacteria, deterrence of herbivores, or attracting pollinators.

The number of species in sect. *Garcinia* that has been studied is still relatively small. The only three species examined are *G. celebica*, *G. mangostana* and *G. diospyrifolia*. The main compounds found are xanthone or triterpene-derived compounds, as summarised in Table 3.2.

Table 3.2. Some natural products extracted from species of sect. *Garcinia*.

Species	Chemical content
<i>G. celebica</i>	1. Friedolanostanes and Lanostanes Rukachaisirikul et al. (2000, 2005); Vieira et al. (2004). 2. Protostane triterpenes Rukachaisirikul et al. (2003).
<i>G. mangostana</i>	1. Xanthone Jung et al. (2006); Nilar & Harrison (2002); Suksamrarn et al. (2002); Huang et al. (2001); Gopalakrishnan & Balaganesan (2000); Gopalakrishnan et al. (1997); Ahluwalia & Tehim (1984); Parveen & Khan (1988); K-Sen et al. (1980); Asai et al. (1995); Nilar et al. (2005); Govindachari et al. (1971). 2. Benzophenone Nilar et al. (2005); Huang et al. (2001). 3. Triterpene Parveen et al., (1991).
<i>G. diospyrifolia</i>	1. Xanthone Goh et al. (1992).

Some of these compounds have been tested for their potential in pharmacology, for example as anti-HIV, anti-fungal or bacterial, anti-cancer agents, and as anti-histamines.

Table 3.3 lists species with pharmacological potential in sect. *Garcinia*.

Table 3.3. Species with pharmacological uses in sect. *Garcinia*.

Species	Pharmacological Potential
<i>G. celebica</i>	1. Anti-HIV Rukachaisirikul et al. (2003).
<i>G. mangostana</i>	1. Anti-bacteria/Anti-fungal Chomnawang et al. (2005); Voravuthikunchai & Kitpipit (2005); Sakagami et al. (2005); Suksamrarn et al. (2003); Iinuma et al. (1996); Sundaram et al. (1983). 2. Anti-cancer Nabandith et al. (2004); Moongkarndi et al. (2004a); Matsumoto et al. (2003); Ho et al. (2002). 3. Anti-histamine Nakatani et al. (2002); Chairungsrilerd et al. (1996). 4. Anti-HIV Chen et al. (1996). 5. Anti-oxidation Jung et al. (2006); Moongkarndi et al. (2004b); Williams et al., (1995). 6. Anti-Inflammatory Gopalakrishnan et al. (1980); Shankaranarayan et al. (1979).

## **CHAPTER 4**

### **MORPHOLOGICAL CHARACTERS**

#### **4.1 Introduction**

This chapter will provide a discussion of the morphological characters of the species within sect. *Garcinia*. There are two important groups of character. Firstly, the one that defines the sectional classification and the grouping of the species, where characters chosen are fairly constant for whole groups of species, but vary across the genus. These characters are mostly taken from the male flowers and fruits with some additional characters from the leaves. The significance of these characters is also demonstrated by the monophyly of the sections and groups from the molecular analysis (Chapter 5). Secondly, for species delimitation, characters are chosen which are constant or continuously variable within local populations of the same species, but show clear gaps in variation between putative species. These characters come from both vegetative and reproductive features.

#### **4.2 Materials and Methods**

The morphological characters are described from herbarium specimens and also from my own observations during field work in Malaysia and Indonesia. Herbarium materials were from herbaria of the Royal Botanic Garden Edinburgh (**E**) with additional specimens loaned from the Royal Botanic Gardens Kew (**K**), Leiden (**L**), Natural History

Museum Paris (**P**), Singapore Botanic Garden (**SING**), Arnold Arboretum (**A**), University of California (**UC**) and Forestry Department Sarawak (**SAR**). Apart from that, herbaria were personally visited during the field work, those of herbarium Kebun Raya Bogor (**BO**), Forest Research Centre Sabah (**SAN**), Forest Research Centre Sarawak (**SAR**) and Forest Research Institute Malaysia (**KEP**).

Information on the label of the herbarium sheets were stored in the PADME database system, developed at RBGE. Amongst the information included were scientific and local names, collector, date collected, locality or any notes written by the collectors. Geographical co-ordinates of the specimens were found on the GEOnet Names Server (<http://earth-info.nga.mil/gns/html/index.html>) and the co-ordinates were exported from PADME to DIVA-GIS (Hijmans et al., 2001) to map distributions.

Morphological characters were examined by careful observation of herbarium specimens with the help of a hand-lens or stereo microscope. To study the characters of flowers, specimens were first re-hydrated by boiling in water for 10-15 minutes before dissection. Measurements of the leaves are taken from the leaves presumed to be mature and the terminology for leaf morphology follows Hickey's (1979) classification. Detailed information on the field characters of the trees, for example the height or the colour of the bark, were taken either from my own observations, published floristic accounts or from the collectors' notes on the herbarium specimens.

### 4.3 Vegetative Characters

#### 4.3.1 Leaves

As in the entire genus, leaves in sect. *Garcinia* are simple and oppositely arranged while the margin is always entire and finely revolute towards the lower part of the leaves when dry. Whitmore's (1973) work demonstrated that some *Garcinia* species could be identified to species level by using leaf characters alone. Jones (1980), using radiography techniques, demonstrated that there is large amount of variation in leaf characters, such as the patterns of glandular lines and secondary venation that could be used for species recognition in *Garcinia*.

However, the use of leaf characters in isolation should be treated with caution throughout the whole genus. This is because convergence of vegetative characters means that they do not indicate higher level relationship accurately in all cases. For instance, *G. vidua* (previously included in sect. *Garcinia* by Jones (1980) and *G. nitida* have similar venation that is brochidodromous, with similar glandular line patterns. However, *G. vidua* is not related to *G. nitida* as indicated by its axillary and sessile inflorescences. Whitmore (1973) also found that there were many *Garcinia* species in Peninsular Malaysia (in different sections) with very similar leaf shape and secondary venation but that have different flower and fruit characters. Despite these problems of using vegetative characters at higher taxonomic level within sect. *Garcinia*, some species can be identified by using single 'spot' characters from the leaves or with combinations of characters (see below).

#### 4.3.1.1 Colour

Most of the leaves in *Garcinia* turn brownish when dry but for recent specimens less than five years old, the lamina usually is tinged greenish or yellowish in colour. However, the leaves of *G. penangiana* are a bright reddish colour when dry, making it easy to identify when compared with all other *Garcinia* species.

#### 4.3.1.2 The shape and size

The shape and sizes of the leaves varies greatly even within the same species (Whitmore, 1973) and this is clearly seen in widely distributed species such as *G. celebica*, found from eastern India to Papua, which has elliptic, widely elliptic, lanceolate, sub-orbiculate or round shaped leaves, with size ranging from 3.3-17.5 cm long and 1.7-11.0 cm wide. In general the leaves of sect. *Garcinia* are elliptic derivatives (very narrow elliptic, narrow elliptic, elliptic, wide elliptic, suborbiculate or orbiculate) or sometimes ovate or broadly ovate, and the sizes vary from the smallest of 0.8 cm (e.g. *G. exiguus*; Figure 4.1a) to the biggest of 25.5 cm long (e.g. *G. mangostana*; Figure 4.1b).

Nevertheless, some species have a distinct shape and size which can be used as a spot character. For instance, *G. harmandii* is the only species in the section with leaves of oblanceolate or spatulate shape (Figure 4.1c), *G. cataractalis* has small leaves (to 2.1 cm long) that are very narrow elliptic or lanceolate in shape (Figure 4.1d), and *G. exiguus* has small leaves (less than 2 cm long) that are broadly elliptic in shape (Figure 4.1a).

The drip tips formed by the elongation of the leaf apex are a common feature among *Garcinia* species and are ecologically associated with understorey trees in tropical rainforest. They retard the growth of epiphylls and reduce the loss of soluble nutrients from leaching by hastening the drainage of water films (Whitmore, 1988). For example, drip tips are found in *G. sanguisangud* or *G. diospyrifolia* var. *minor* (Figure 4.1e) which can sometimes reach 1.5 cm long.

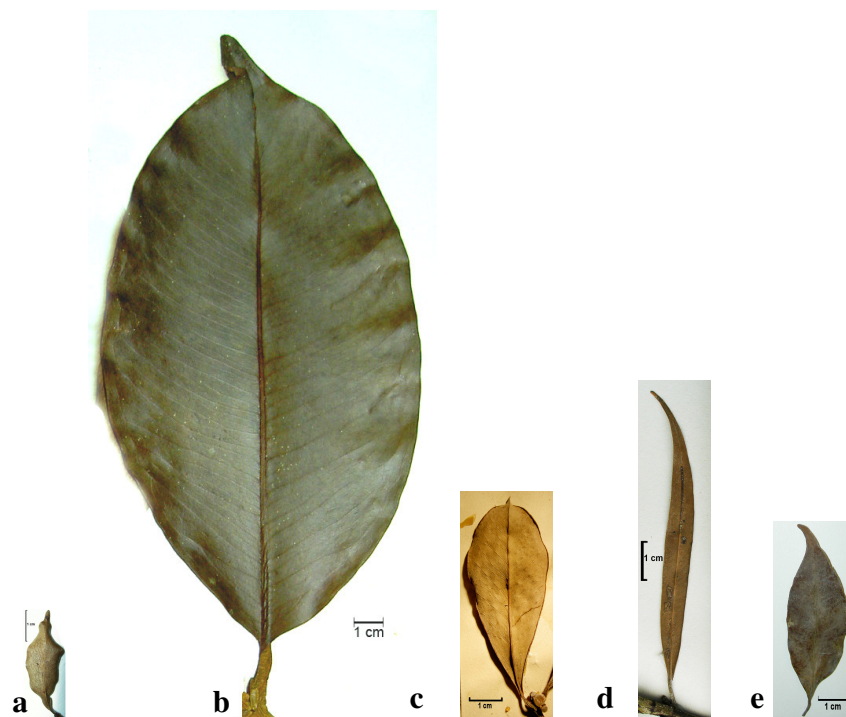


Figure 4.1. Leaf shape and size. — **a.** *Garcinia exiguus* Nazre (Coode *et al.* 7886, A), scale bar = 1 cm; **b.** *G. mangostana* L (Salma & Sohaime 787, MARDI); **c.** *G. harmandii* Pierre (Thorel 1018., K); **d.** *G. cataractalis* Whitmore (Whitmore FRI20350, K); **e.** *G. diospyrifolia* var. *minor* Ng ex Nazre (Wong 32216, A).

#### 4.3.1.3 Petiole

In the forest, *Garcinia* can be identified by their petioles that clasp the twigs in a way that is similar to the unrelated *Fagraea* (Loganiaceae), a genus that does not produce the yellow or white exudate of Guttiferae (Whitmore, 1973). The lower part at the base where the petiole is clasping the twigs is sometimes excavated and enlarged to produce a small ligule-like organ as in *G. venulosa* (Figure 4.2a). The petiole when dry has fine, very closely arranged, horizontal striae that are obvious in most species of *Garcinia* (Figure 4.2b). The upper surface of the petioles is usually sunken or channelled.

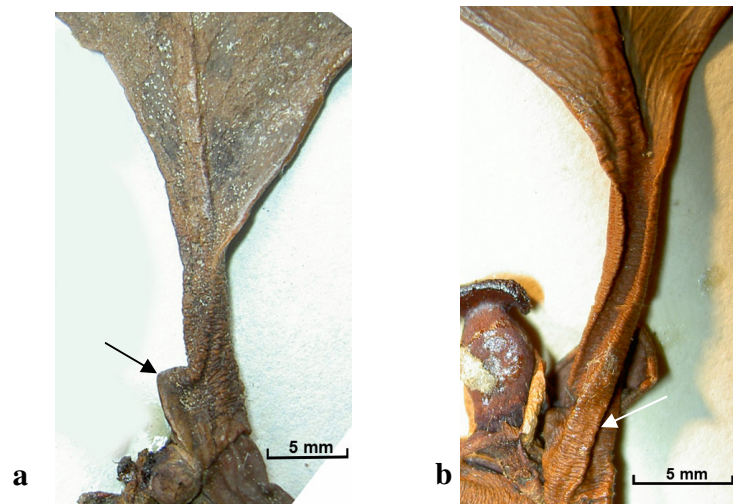


Figure 4.2 Petiole and ligule . — **a.** *G. venulosa* (Blanco) Choisy (*Cumming 2114*, K) showing ligule (arrow); **b.** *Garcinia celebica* L. (*Anon 115*, K) showing striae (arrow).

#### 4.3.1.4 Midrib

Descriptions of the midribs were taken from the dried leaves, with the characters derived from the upper side of the leaf, usually near the middle of the lamina. Three

different orientations of midribs are observed: 1) a raised midrib with a square-shaped cross section and flattened upper surface (Figure 4.3a); 2) a raised midrib with a convex-shaped cross section and weakly flattened, pointed or concave upper surface (Figure 4.3b); and 3) the midrib sunken or flattened to the same level as the lamina (Figure 4.3c). In sect. *Garcinia*, the first type of midrib is restricted to *G. diospyrifolia*, *G. ochraceus*, *G. penangiana* and *G. nitida*. While the second type of the midrib is commoner in *G. celebica*, *G. malaccensis*, *G. venulosa* and *G. rigida*, the last type of sunken or flattened midribs is distinct in *G. moselleyana* and *G. harmandii*.

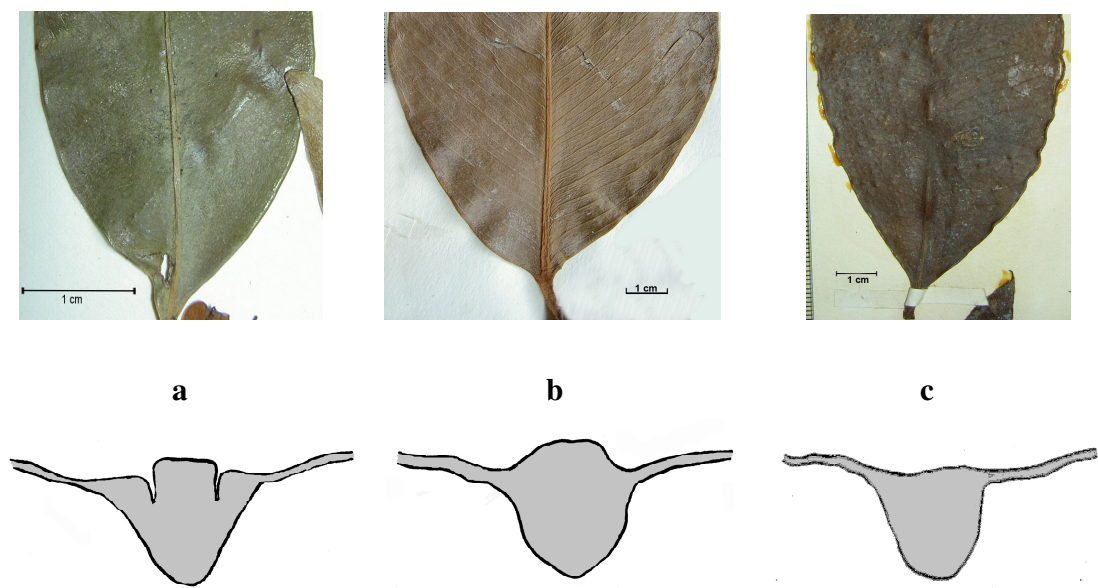


Figure 4.3. Abaxial surfaces of the midribs (above) and cross sections (below). — **a.** *Garcinia nitida* Pierre (Argent 9318, E); **b.** *G. celebica* L. (Harreveld s.n., L); **c.** *G. moselleyana* Pierre (Moseley s.n., K).

On the lower (adaxial) surface of the leaves, the midrib in sect. *Garcinia* is always raised and prominent, but sometimes shrinks and becomes more pointed or angular (in cross section), for example in *G. acuticosta* or *G. discoidea*.

#### 4.3.1.5 Secondary veins

Secondary venation in sect. *Garcinia* is mostly uniform, fine but clearly visible on the adaxial surface, and brochidodromous where the veins become united behind the margin to form intramarginal veins. For a few species that have big, thick leathery leaves such as *G. mangostana*, *G. malaccensis* and *G. venulosa*, a second intramarginal vein is present and located closer to the margin, which becomes a good spot character for these species (Figure 4.4a). However, the second intramarginal vein is only distinct in the female specimens and very faint or not present in the male. Inter-secondary veins are also a common feature, e.g. in *G. penangiana* (Figure 4.4b).

Cladodromous venation, where secondary and tertiary veins form tree like branches is exclusively found in sect. *Garcinia* in *G. moselleyana* and *G. harmandii*. The veins end without touching the margin or at the intramarginal vein (Figure 4.4c & 4.4d).

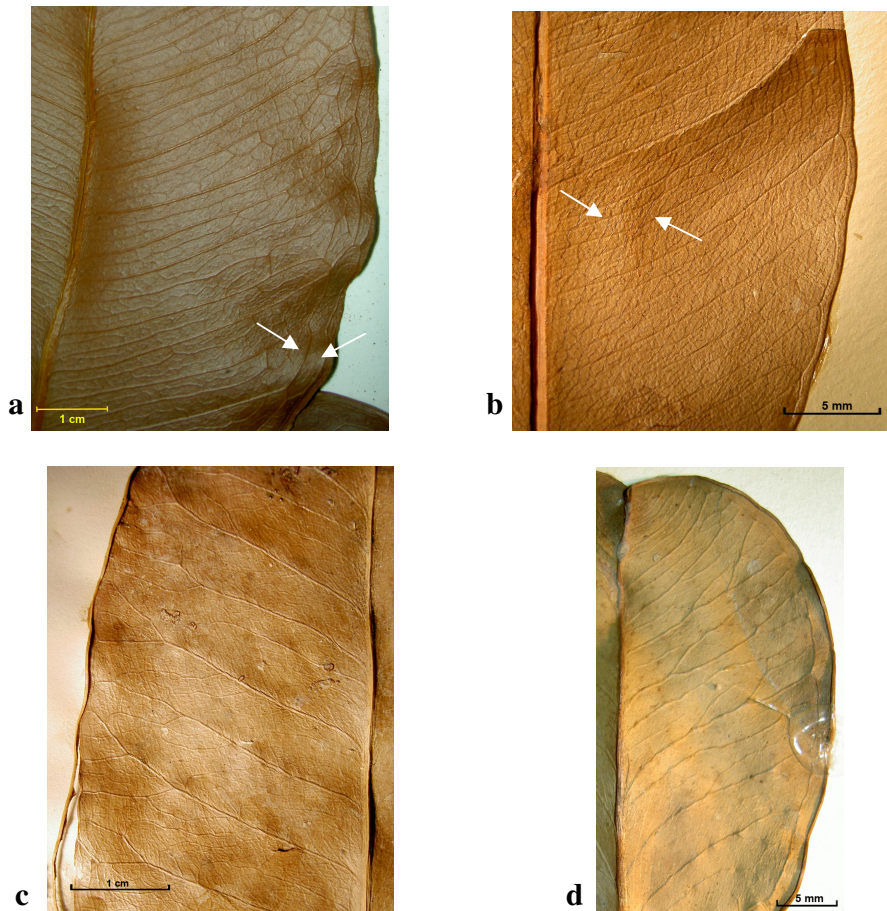


Figure 4.4. Secondary venation; a – b: brochidodromous, c-d: cladodromous. — **a.** *Garcinia mangostana* L. (Shamsudin s.n., MARDI), arrows highlight two intramarginal veins; **b.** *G. penangiana* Pierre (Henderson 21689, K), arrows highlight inter-secondary veins; **c.** *G. moselleyana* Pierre (Wenzel 3036, K); **d.** *G. harmandii* Pierre (Thorel 1018, K).

#### 4.3.1.6 Glandular Lines

Jones (1980) suggested that glandular lines might provide useful characters for species delimitation, but would not be not very useful for infra-generic treatments. There are two different systems of glandular lines between the upper and lower layers of the leaf. Sprecher (1919; in Jones 1980) was the first to realise this and found that the

glandular lines on the lower surface are situated in the lower parenchyma and those on the upper surface under the palisade layer. Anatomically, glandular lines are canaliform in the mesophyll and differ from the mesophyll cells by the surrounded rings of 5-7 bordered cells (Jones, 1980).

The pattern of glandular lines from the lower (abaxial) part of the leaves is quite useful in species delimitation and identification of species of sect. *Garcinia*, but only when used in conjunction with other leaf characters. For most species, the glandular lines are fine wavy lines of a thumb-print pattern, which only run in one direction, from the midrib to the margin, across the secondary veins, in blackish, greyish, brownish or dark green colours. However, in a few species such as *G. ochraceus*, *G. discoidea* and *G. moselleyana*, the glandular lines are not visible or very faint. Sometimes, in old herbarium specimens collected in the 19<sup>th</sup> century, the colour of the glandular lines is faded and obscure.

Three main variations of glandular line pattern are found in sect. *Garcinia*. The first is the most common where the lamina is dominated by long wavy lines of differing lengths, and occasionally shorter lines are present alongside the longer lines (Figure 4.5a & b). This type is found in *G. acuticosta*, *G. cataractalis*, *G. celebica*, *G. exiguus*, *G. malaccensis*, *G. mangostana*, *G. rigida*, *G. venulosa*, *G. harmandii* and *G. penangiana*. Sometimes the distance between the lines can differentiate two species. For example, the lines in *G. celebica* are much closer than to those of *G. rigida*. In few species, such as *G. penangiana* (Figure 4.5a) and *G. cataractalis*, the glandular lines are sometimes mistaken

as secondary venation due to their size and distinctness. In *G. exiguus*, the glandular lines are strongly raised compared to the secondary venation, which make the latter hardly visible on the lower surface of the lamina (Figure 4.5b).

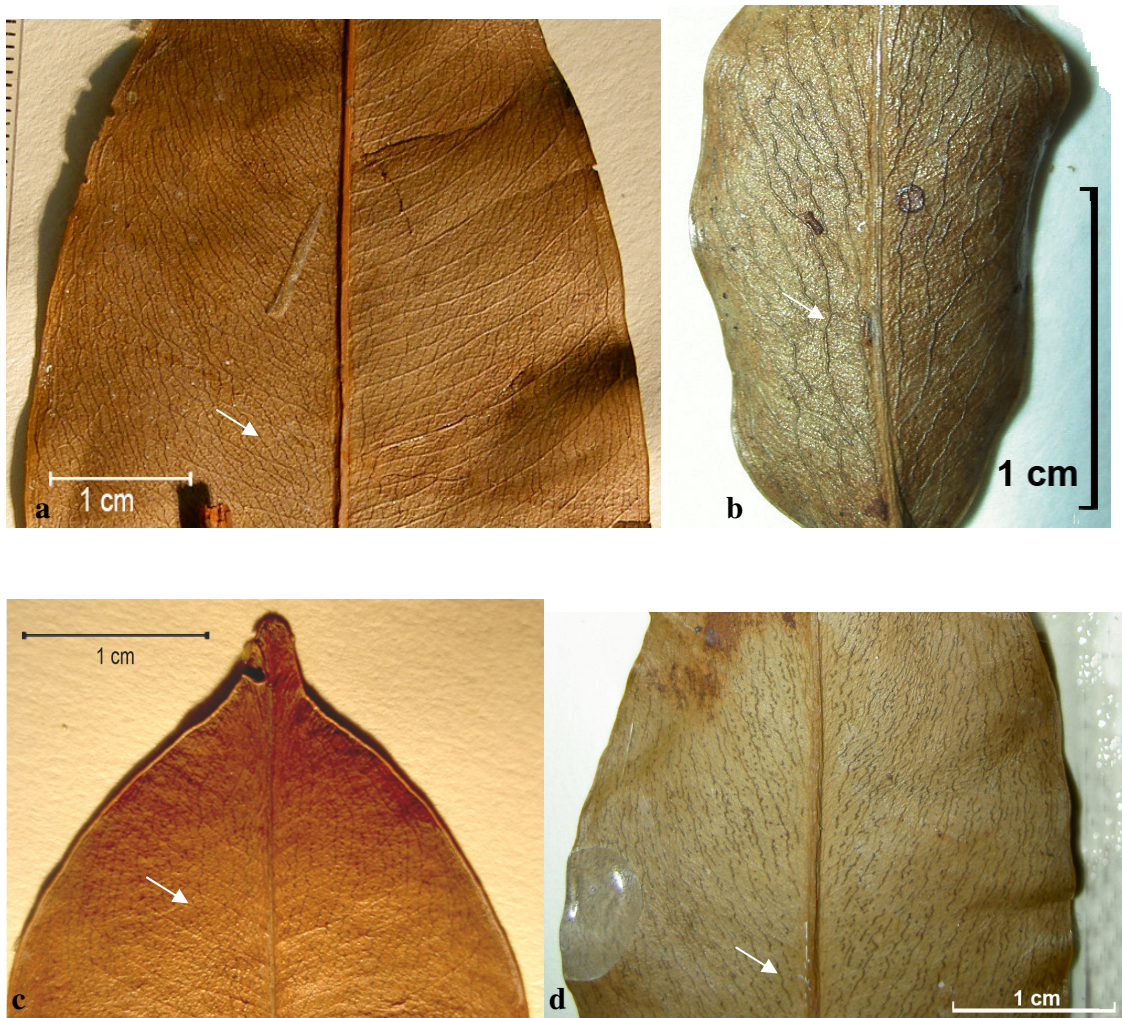


Figure 4.5 Glandular line patterns; arrows highlight glandular lines. — **a.** *Garcinia penangiana* Pierre (Henderson 21689, K); **b.** *G. exiguus* Nazre (Coode 7886, A); **c.** *G. nitida* Pierre (Haviland 2339, K); **d.** *G. sangudsangud* Nazre (Aban SAN32977, L).

In the second type, the lamina is dominated by long wavy lines but there are much shorter lines and dashes in between these long lines. Species with this characteristic pattern are *G. nitida* (Figure 4.5c) and *G. diospyrifolia*. The third pattern is found exclusively in *G. sangudsangud* which has long wavy lines that are nearly equal number to the short lines and dashes, and in addition there are many dots alongside the short lines and dashes (Figure 4.5d).

## 4.4 Floral Characters

### 4.4.1 *The Inflorescence*

The basic type of inflorescence in sect. *Garcinia* is a simple cyme (Figure 4.6a, b) in which few flowers (2 to 16) are clustered together and arranged mostly in terminal branches except in *G. moselleyana*, which has both axillary and terminal inflorescences. In this study, the term cluster as used by Whitmore (1973) is preferred for use in species descriptions because ‘cyme’ is defined as a flat-topped or round-topped determinate panicle inflorescence, in which the terminal flowers bloom first (Harris & Harris, 2004). Species of sect. *Garcinia* are dioecious and there are two obvious differences between male and female inflorescences. Firstly, the male flowers are usually greater in number, and secondly the male flowers are slightly smaller in size than the female flowers. The size of the flowers is often not uniform. For example in *G. celebica*, the male flowers range from 10 mm to 25 mm across. For species such as *G. malaccensis* or

*G. mangostana*, where occasionally the flowers are in triads, the flower in the middle is always largest.

The female flowers are usually solitary in most of the species in sect. *Garcinia* or occasionally in pairs such as in *G. nitida*. Since the female flowers are bigger than the male, the pedicels are stouter in the female compared with more slender, longer pedicels in the male. In general the shape of the pedicel is angular or round and bracts are usually present. Bracts can be persistent or caducous, are small and triangular reaching 1.7 cm long, but are insignificant in species delimitation.

Generally, inflorescence type is a good synapomorphic character for sectional delimitation in *Garcinia*. For instance, species removed from the section in this study, *G. anomala*, *G. costata*, *G. maingayi*, *G. pseudo-guttiferae*, *G. tonkinensis* and *G. trianaii* have thyrsoid inflorescences composed of 3-flowered cymes (Figure 4.6c) whereas species in sect. *Garcinia* have inflorescences arranged in clusters of few to many flowers.

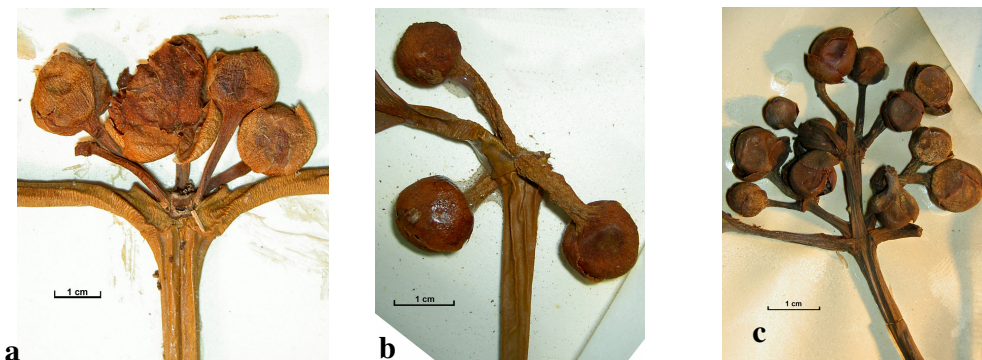


Figure 4.6. Inflorescences in section *Garcinia* (a – b) and omitted species from sect. *Garcinia* (c). — **a.** *Garcinia celebica* L. (Hamilton 1113, A); **b.** *G. mangostana* L. (Shamsudin s.n., MARDI); **c.** *G. maingayi* Hk. f. (Curtis 1610, K).

#### 4.4.2 Sepals and Petals

The number of sepals and petals in sect. *Garcinia* is always four except in *G. moselleyana* which has five. Pentamerous sepals and petals are not common in *Garcinia* except in sect. *Macrostigma*, though species such as *G. xanthochymus* (sect. *Xanthochymus*) have both pentamerous and tetramerous sepals and petals (Jones, 1980). However, *G. moselleyana* is included in the sect. *Garcinia* because the stamens are arranged into 4-lobed bundles and the anthers are 2-locous. When there are four sepals and petals, they are arranged in pairs (decussate).

The colour of the sepals is usually green or yellowish and the outer part is thicker than the inner, and when compared with the petals, the sepals are usually thicker but smaller. The petals, however, have a brighter colour, from yellow to white, cream, and pink or red. Between the male and female flowers of the same species, petal shape is usually not variable, but between different species, shape varies from ovate to elliptic, or oblanceolate to obovate.

Unlike the petals, the sepals (calyx) on female flowers are persistent and remain on the fruits. Morphologically, the calyx on the fruits is leathery for all species in sect. *Garcinia*; *G. malaccensis* and *G. mangostana* have a thicker calyx than other species. The shape is variable from ovate to obovate but always concave while the size varies from 1-3 cm long.

#### 4.4.3 Male Flowers

The male flower has been used most extensively in infrageneric classifications by Jones (1980) and species delimitation by Whitmore (1973) who worked on 49 species of *Garcinia* in Peninsular Malaysia. Both authors also used male flower characters to construct artificial groupings without formal taxonomic rank in order to help the identification process. In sect. *Garcinia*, however, using male flower characters alone is not sufficient to construct either higher groups or to use in species delimitation. The taxonomic account (Chapter 6) will mostly use male flower, fruit, and vegetative characters. However, for species where the male flower is absent, DNA sequence data are used to help the placement of the species into the groups.

##### 4.4.3.1 Stamens

The stamens in sect. *Garcinia* are always united in a bundle at the centre of the flower. Based on their shape, stamen bundles can be divided into three types;

1. Distinctly 4-lobed, stamens usually open when mature (e.g. *G. celebica* and *G. discoidea*).

The stamen bundle is attached at the base of the pistillode above the petals and the lobes are always arranged opposite the petals (Figure 4.7a; 4.8a & 4.9a).

2. Weakly 4-lobed, stamens hardly open when mature (e.g. *G. nitida*, *G. diospyrifolia*, *G. sangudsangud*, and *G. ochraceus*).

The attachment and the arrangement of the stamen bundle is similar to the above but the lobes will always enclose the pistillode, even in mature flowers (Figure 4.7b).

3. 4-angled (e.g. *G. penangiana*, *G. venulosa*, *G. mangostana* and *G. malaccensis*).

The stamens, if the pistillode is present, are located at the base of the pistillode, and are usually bigger than the stipe of the pistillode (Figure 4.7c – g; Figure 4.9b – f).

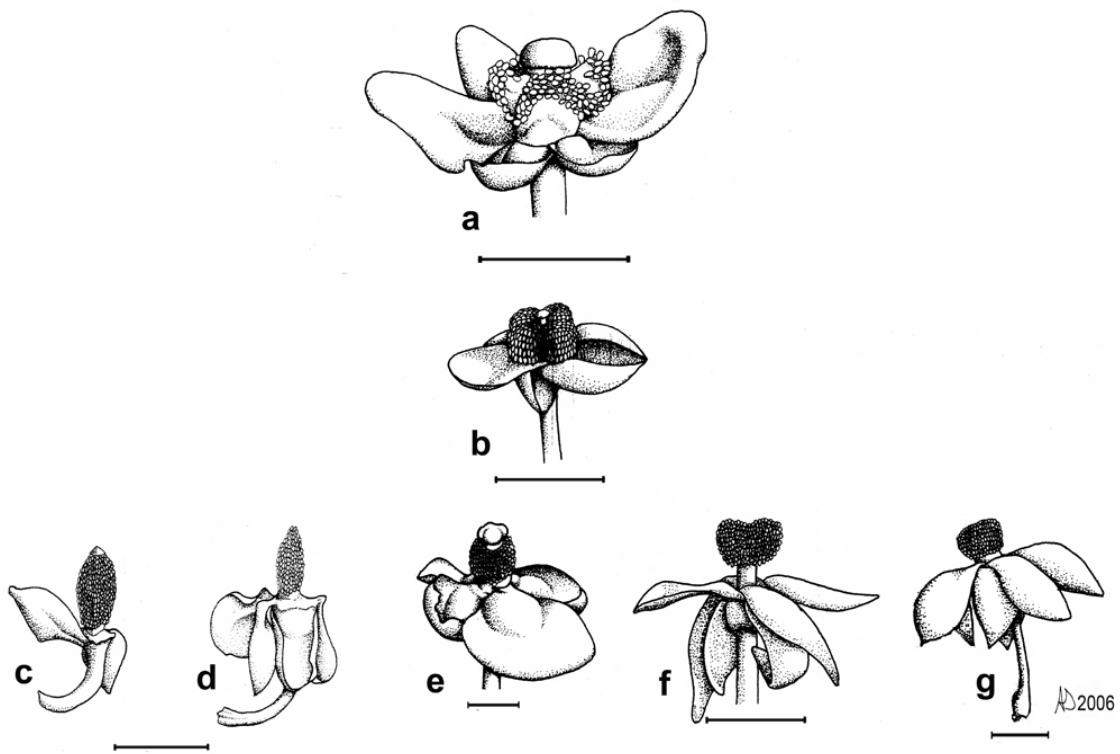


Figure 4.7. Variation of male flowers; Type 1= a, Type 2= b and Type 3= c to g. — **a.** *Garcinia celebica* L. (Nazre 062, E); **b.** *G. sangudsangud* Nazre (Aban SAN 32914, L); **c.** & **d.** *G. malaccensis* var. *malaccensis* (c. Ashton BRUN3392, SAR; d. Maingay 149, K); **e.** *G. mangostana* L. (Shamsudin s.n., MARDI); **f.** *G. penangiana* Pierre (Whitmore FRI20347, L); **g.** *Garcinia malaccensis* var. *pseudomangostana* Nazre (Madani 50574, L). Scale bar = 1 cm. Drawn by Anna Dorward.

The stamens number c. 100, and the anthers are two thecous, recurved and dehisce by longitudinal slits. These anthers are either with short filaments or completely sessile. Using stamen characters, *G. anomala*, *G. costata*, *G. maingayi* and *G. trianii* are omitted from sect. *Garcinia* because their stamens are arranged in an annular mass at the centre, without lobes and encircle the pistillode, compared to 4-angled or 4-lobed stamen bundles in sect. *Garcinia* (Figure 4.8). Another omitted species, *G. vidua* is, however, quite different from other species having very few anthers (c. 10) clustered in the middle of the flower and the pistillode is not present. From these characters it appears closely related to many species from sect. *Brindonia* sensu Jones (1980).

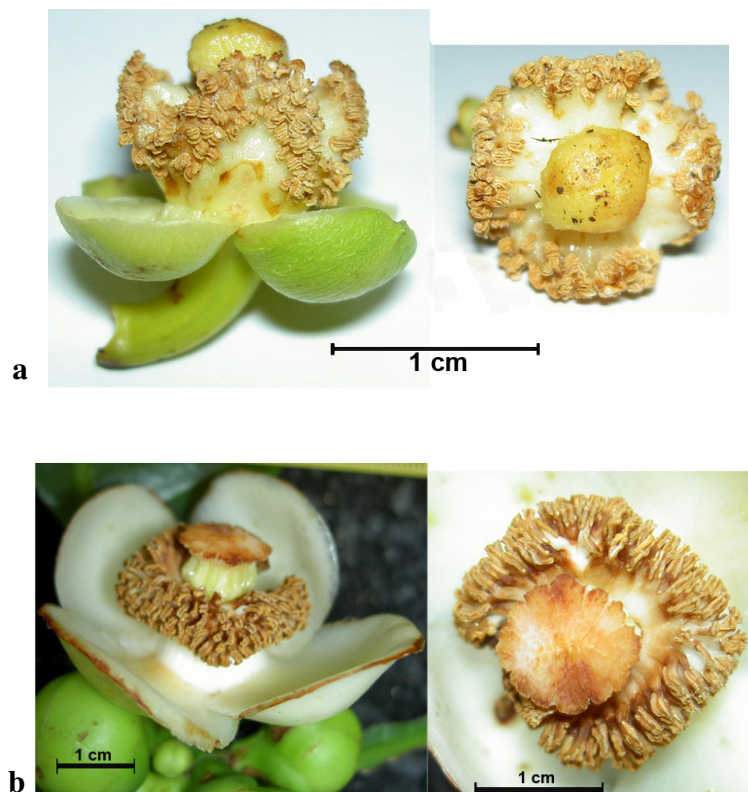


Figure 4.8. Comparison of male flowers in section *Garcinia* (a) and omitted species (b). — **a.** *G. celebica* L.; **b.** *G. costata* Hemsl. ex King

#### 4.4.3.2 Pistillode

The pistillode in the male flower of many *Garcinia* species is homologous with the gynoecium of the female flower (Jones, 1980). Morphologically, pistillodes in sect. *Garcinia* have a fungiform-like shape, consisting of a cap and the shaft (or stipe), which is homologous to the stigma and ovary, respectively (Figure 4.9a & b). The pistillodes are small, from 1 mm long, e.g. *G. malaccensis* var. *malaccensis* (Figure 4.9c & d), to 5 mm long e.g. *G. mangostana* (Figure 4.9b). The stipe can be slender or ovoid and the margin of the cap may be crenate or lobed. However, not all species in sect. *Garcinia* possess a pistillode (Figure 4.9e & f), which is lacking in, for example, *G. penangiana*, *G. venulosa* and in few individuals of *G. malaccensis*.

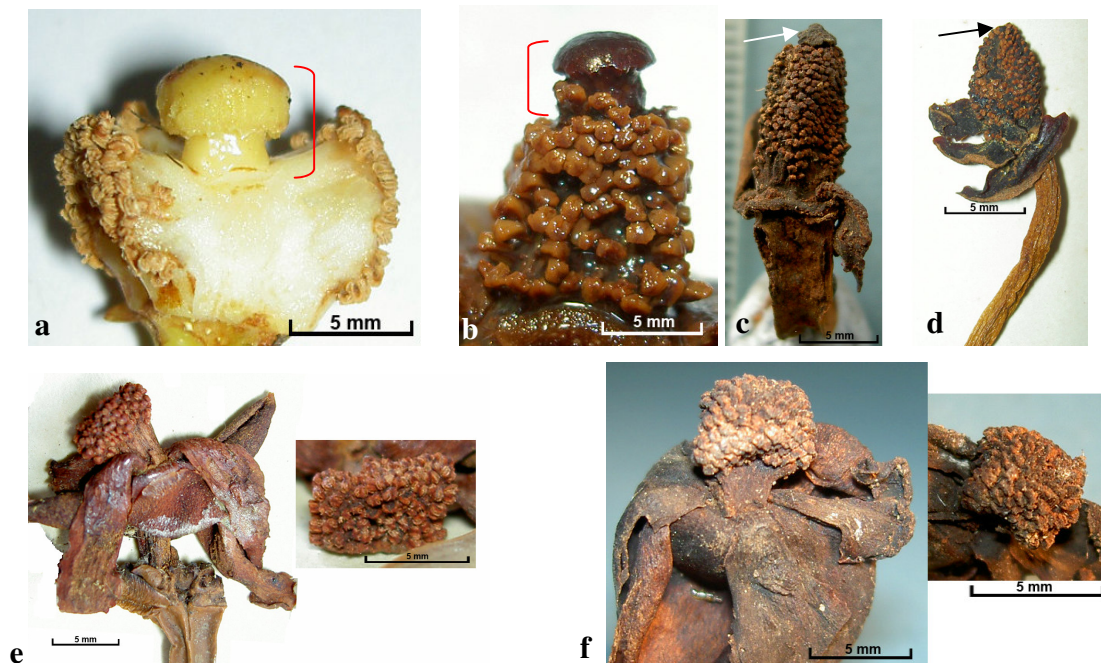


Figure 4.9. Variations of pistillode (a–d), bars and arrows highlight the pistillode; species without a pistillode (e–f.) — **a.** *Garcinia celebica* L. (Nazre 062, E); **b.** *G. mangostana* L. (Shamsudin s.n., MARDI); **c.** & **d.** *G. malaccensis* var. *malaccensis* (c. Ashton BRUN3392, SAR; d. Cockburn FRI18181, KEP); **e.** *G. venulosa* (Blanco) Choisy (Alvarez 18513, K); **f.** *G. penangiana* Pierre (Burbridge s.n., K).

#### 4.4.4 Female Flowers

##### 4.4.4.1 Ovary

For most of the species in sect. *Garcinia* (*G. acuticosta*, *G. cataractalis*, *G. discoidea*, *G. exiguus*, *G. nitida*, *G. ochraceus*, *G. rigida* and *G. venulosa*) the absence of female flowers on herbarium specimens hindered the use of the ovary for comparative studies. In general, the ovary in *Garcinia* is superior and very few species have constant locule numbers (Jones, 1980). In sect. *Garcinia*, most species have 4 or 5 locules as in *G. celebica* and *G. malaccensis*. Two shapes of the ovary are found in sect. *Garcinia*; ovoid (flask-shaped) as in *G. diospyrifolia* and *G. celebica* to broadly ovate (dwarf fungi-form) as in *G. nitida*, *G. mangostana*, *G. malaccensis* and *G. sangudsangud* (Figure 4.10a – c); and orbiculate or globose as in *G. harmandii*, *G. moselleyana* and *G. penangiana* (Figure 4.10d). Because little variation is found in the ovary, it has less taxonomic value and is not really an important character for species circumscription in sect. *Garcinia*.

##### 4.4.4.2 Stigma

The stigma in sect. *Garcinia* crowns the top of the ovary, is blackish in colour, and sessile or raised to 2 mm long. For most of the species, the stigma is usually fungiform cap-shaped or skull-cap shaped (hemispherical), smooth or corrugated, with or without lobes, and the margin is irregularly crenate (Figure 4.10a – d). Stigma characters

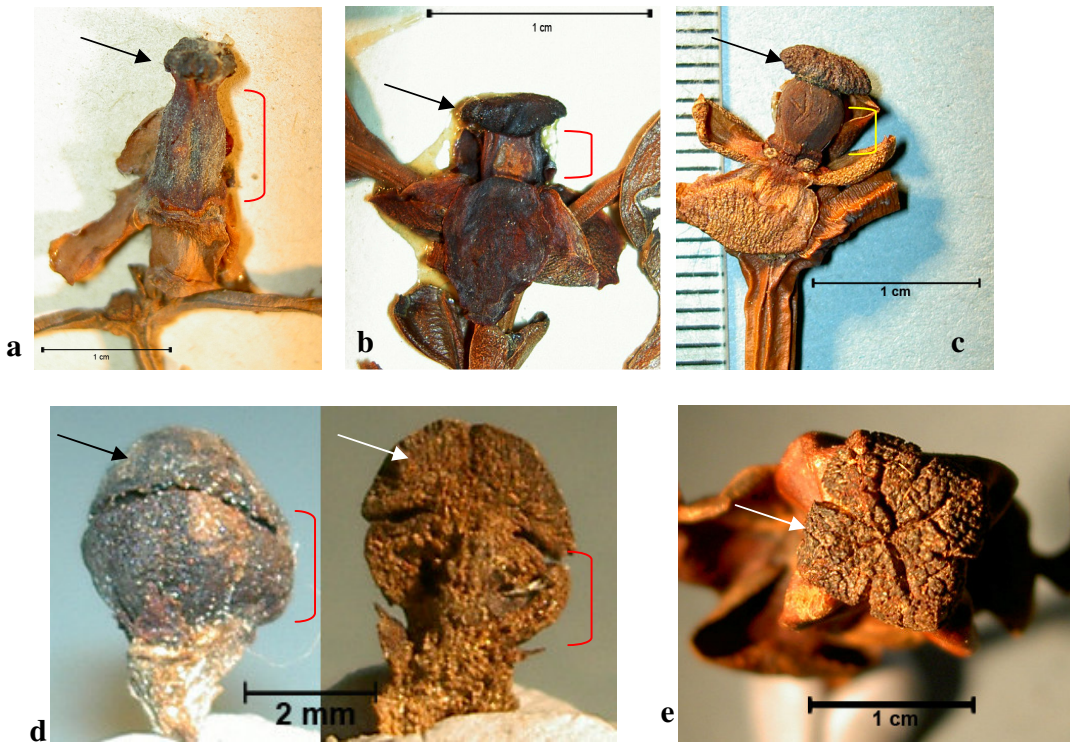


Figure 4.10. Variation in ovaries and stigmas, bars indicate ovary and arrows indicate stigma; a – c: ovoid or broadly ovate ovary with fungiform cap-shaped stigma; d : globose ovary with skull cap-shaped stigma; e: stigma in 4-bundles with corrugated surface. — a. *G. diospyrifolia* var. *diospyrifolia* (Zainal AA914, A); b. *G. nitida* Pierre (Haviland 2339, K); c. *G. celebica* L. (Anon 115,K) d. *Garcinia moselleyana* Pierre (Elmer 7187, K); e. *G. penangiana* Pierre (Chai S32334, L).

in the female flowers are not as useful as the stigma characters in the fruits (see below) for species circumscription because with the exception of a few species, less variation is found. For example, *G. moselleyana* has a skull-cap shaped stigma and is distinctly different from other species because the stigma is bigger or nearly the same as its globose ovary (Figure 4.10d). Another example is *G. mangostana* with its skull-cap shaped stigma of 4-5 lobes, and smooth surface, and *G. penangiana* with a skull-cap shaped stigma of 3-4 bundles with rather corrugated surface (Figure 4.10e). These characters are retained on the mature fruit (see stigma on fruit below) making the stigma characters even more

useful. However, the use of the stigma to identify other species such as *G. diospyrifolia*, *G. celebica*, *G. nitida*, *G. venulosa* and *G. malaccensis* is difficult because of the similarity of their fungiform cap shaped stigmas (Figure 4.10a – c).

#### 4.4.4.3 *Staminodes*

Staminodes of the female flower are homologous to the stamens of the male flower and differ from functional stamens by not producing any pollen. In sect. *Garcinia*, the appearance of the staminodes does not reflect the morphology of the stamens in the male flowers. For example, two species in sect. *Garcinia*, *G. malaccensis* and *G. mangostana* (Figure 4.11), have staminodes present as free stamens numbering few to 20, surrounding the ovary. However, the stamens for both species are many (c. 100) arranged in central, 4-angled bundles. The staminodes are usually caducous and are sometimes missing on the herbarium sheets.



Figure 4.11. The position of free staminodes in female flowers of *Garcinia mangostana* L. Arrow highlights staminodes.

## 4.5 Fruits

Fruits in sect. *Garcinia* are berries with walls that are thick or thin and woody, or occasionally very hard and woody. Whitmore (1973) described the fruit types in *Garcinia* as fleshy to woody berries because some fruits have an edible fleshy fruit wall. In sect. *Garcinia*, however, no species have an edible fruit wall.

In the absence of male flowers, for some species (e.g. *G. acuticosta* and *G. cataractalis*) fruits provide important characters for species recognition. These characters are mainly from the types of fruit wall, i.e. thick or thin, and variation in the shape of the stigma. Combined with other floral and vegetative characters, the fruit characters also provided significant characters for higher taxonomic groupings in the family (Jones, 1980).

### 4.5.1 Fruit wall

In general there are two types of fruit wall (Figure 4.12). *Garcinia mangostana*, *G. celebica*, *G. discoidea*, *G. venulosa*, *G. sangudsangud* and *G. discoidea* have thickly woody walls (Figure 4.12a). In contrast, *G. rigida*, *G. diospyrifolia*, *G. nitida*, *G. cataractalis*, *G. ochraceus* and *G. penangiana* have thin woody walls which rupture easily when dry (except for *G. rigida* and *G. penangiana*) (Figure 4.12b).

The fruit surface is normally smooth but occasionally some fruits in *G. celebica*, *G. discoidea* and *G. sangudsangud* have small lenticels or dents. Species with thin woody walls such as *G. cataractalis*, *G. diospyrifolia* and *G. nitida* sometimes have longitudinal glandular lines of dark reddish or maroon to dark brownish colour.

The smooth fruit wall without stripes is a good synapomorphy to define sect. *Garcinia* (Figure 4.13a). For example, species excluded from the section include *G. costata* that has a ribbed or segmented wall (Figure 4.13b). The other excluded species *G. maingayi* and *G. trianaii* have a deep green (or blackish when dry) fruit wall striped with pale green or yellow (or brownish colour when dry) (Figure 4.13c).

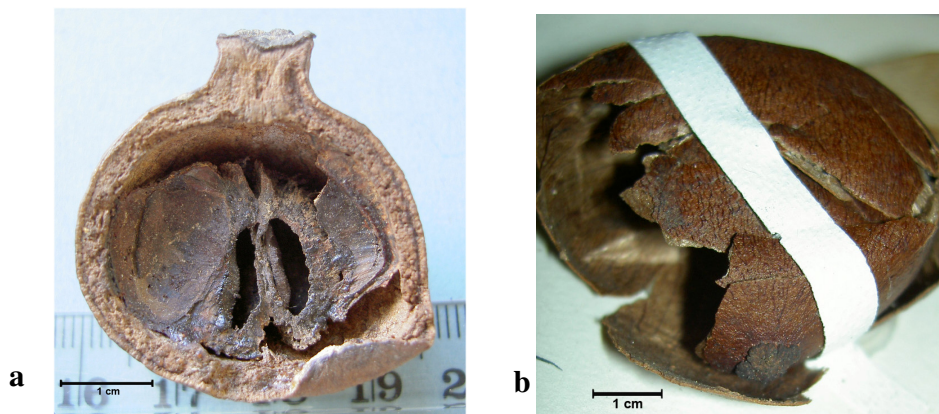


Figure 4.12. Fruit wall type; a. thick and b. thin and easily ruptured. — **a.** *G. celebica* L. (Blume 2026, L); **b.** *G. diospyrifolia* var. *minor* Ng ex Nazre (Wong 32216, A).

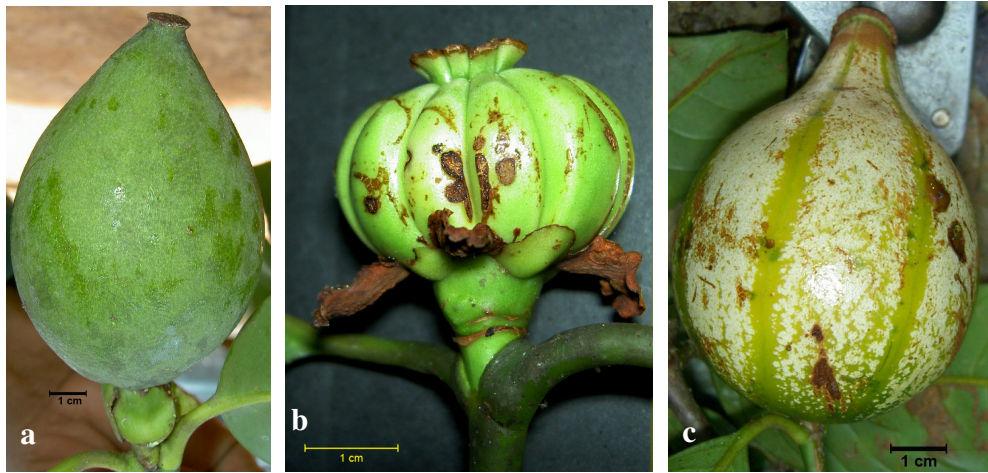


Figure 4.13. Comparison of fruit wall surfaces in section *Garcinia* (a) with omitted species from the section (b, c). — **a.** *Garcinia discoidea* Nazre with smooth wall without stripes; **b.** *G. costata* Hemsl. ex King with ribbed wall; **c.** *G. trianii* Pierre with striped fruit wall.

#### 4.5.2 Colour

The colour of the fruits is normally greenish when young and turning yellowish, reddish or dark purplish when mature. Dried fruit of species such as *G. diospyrifolia*, *G. nitida*, *G. ochraceus* and *G. penangiana* retain their reddish colour while the rest of the species turn brownish or blackish.

#### 4.5.3 Shape and size

The shape of the fruits varies from globose, ovoid (flask-shaped) to ellipsoid (Figure 4.14). The elongation of the stigma from the ovary to become a neck-like structure is quite common, for example in *G. celebica*, *G. diospyrifolia*, *G. nitida* and *G. malaccensis*, while the elongation of the base is rare and only observed in a few specimens of *G. celebica* and *G. malaccensis* var. *malaccensis*.

Fruit size ranges from very small (less than 1 cm) to large (more than 7 cm) (Figure 4.14). *Garcinia mosellejana*, *G. harmandii*, *G. exiguus*, are amongst the species with very small fruit, and *G. cataractalis*, *G. nitida*, *G. diospyrifolia* are species with small fruit (less than 7 cm). The largest fruits are found in *G. mangostana*, *G. malaccensis* and *G. discoidea*.

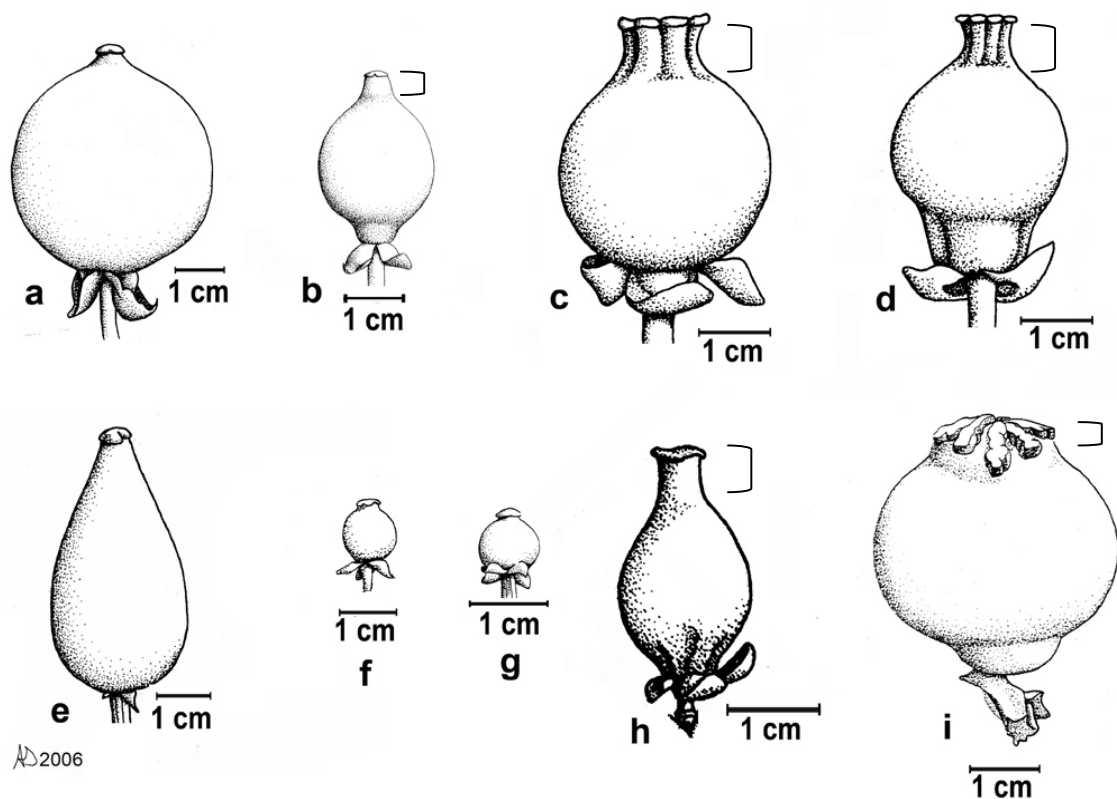


Figure 4.14. Variations of fruit, bars highlight elongation of stigma. — **a.** *Garcinia discoidea* Nazre (Shamsul SA02-10, E); **b.** *G. cataractalis* Whitmore (Whitmore FRI 20350, K); **c & d.** *G. celebica* L.(c: Burkill HMB259, SING; Yahya SFN21419, SING); **e.** *G. diospyrifolia* var. *diospyrifolia* (Soengeng 47, L); **f.** *G. harmandii* Pierre (Pierre 1371, K); **g.** *G. exiguus* Nazre (Chai S31750, SAR); **h.** *G. nitida* Pierre (Argent & Amiril 9318, K); **i.** *G. malaccensis* var. *malaccensis* (Whitmore FRI 8986, L). Drawn by Anna Dorward.

#### 4.5.4 Stigma

The persistent stigma of the fruit is an important character for species delimitation but can be used only in mature or nearly mature fruits. For instance, the young fruit of *G. celebica* strikingly resembles *G. sangudsangud* and *G. venulosa* with a fungiform cap shaped stigma. When mature the stigma of *G. celebica* is weakly or strongly raised with convex or concave surface and 4-8 weak lobes compared to the raised, unlobed stigma, with sunken or flattened surface in *G. sangudsangud*, and sessile stigma with 6-12 square shaped lobes in *G. venulosa*.

Whitmore (1973) considered the stigma on the fruit to be the next most important character after the male flowers for distinguishing species. There are three basic types of stigma in sect. *Garcinia*:

- 1) Distinctly lobed as in *G. mangostana*, *G. malaccensis*, *G. penangiana* and *G. venulosa* (Figure 4.15a – c).
- 2) Weakly lobed as in *G. acuticosta*, *G. cataractalis*, *G. celebica*, *G. diospyrifolia*, *G. discoidea*, *G. exiguus*, *G. moselleyana* and, *G. rigida* (Figure 4.15d – i).
- 3) Without lobes or with irregular crenate or wavy margin as in *G. harmandii*, *G. nitida*, *G. ochraceus* and *G. sangudsangud* (Figure 4.15j – m).

The number of lobes or bundles on the stigma always reflects the number of the locules and seeds. Occasionally the stigma is sessile as in *G. mangostana*, *G. malaccensis* var. *pseudomangostana* and *G. venulosa*, but it is commonly found raised (with neck) to 3 cm as in *G. celebica*, *G. cataractalis*, *G. diospyrifolia*, *G. malaccensis*, *G. nitida*. The overall

shape of the stigma is either a skull-cap or discoid with flattened, convex or sunken surfaces. The colour of the stigma is always darker on the fruit than in the flower, and turns black when dry.

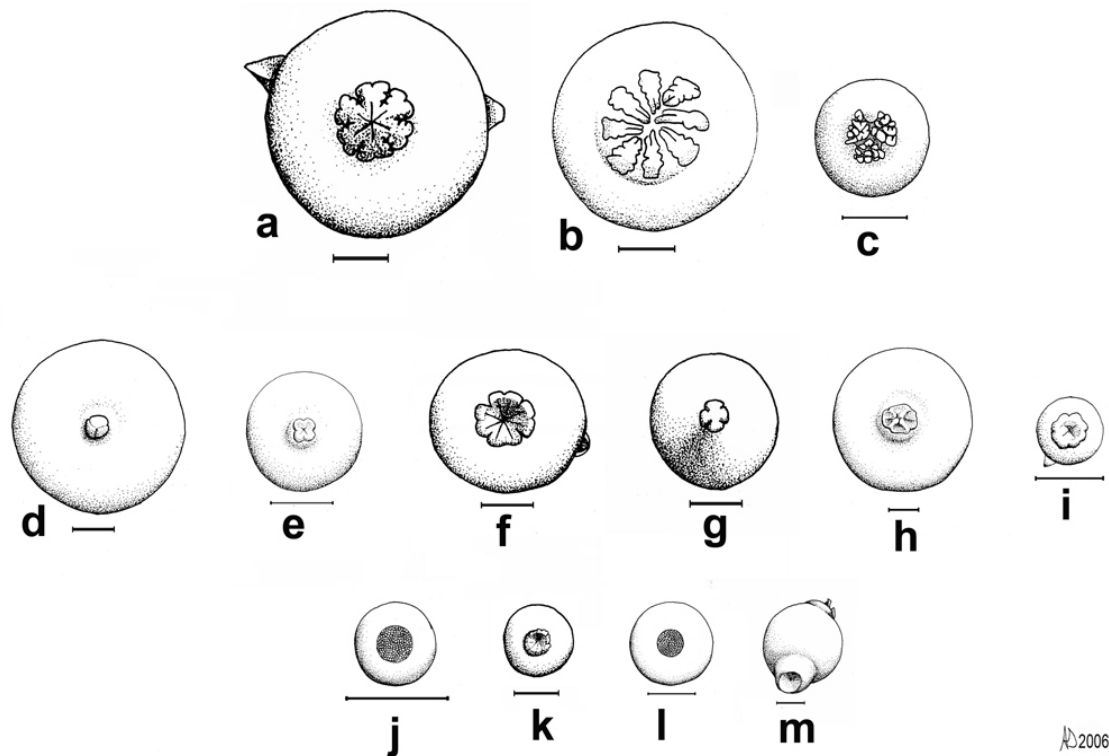


Figure 4.15. Variation of the fruit stigmas; distinctly lobed (a – c); weakly lobed (d – i); and without lobes (j – m). — **a.** *Garcinia mangostana* L. (Sibat S23041, L.); **b.** *G. malaccensis* var. *malaccensis* (Whitmore FRI 8986, L); **c.** *G. penangiana* Pierre (Sinclair SFN 40794, SING); **d.** *G. discoidea* Nazre (Henderson 19654, L); **e.** *G. cataractalis* Whitmore (Whitmore FRI 20350, K); **f.** *G. celebica* L. (Burkill HMB259, SING); **g.** *G. diospyrifolia* var. *diospyrifolia* (Kochummen 77731, L); **h.** *G. discoidea* Nazre (Shamsul SA01-10, E); **i.** *G. exiguus* Nazre (Chai S31750, SAR); **j.** *G. harmandii* Pierre (Thorel 1018, K); **k.** *G. nitida* Pierre (Argent & Amiril 9318, K); **l.** *G. ochraceus* Nazre (Streimann & Martin 52890, E); **m.** *G. sangudsangud* Nazre (Muin Chai SAN 33449, L). Scale bar = 1 cm. Drawn by Anna Dorward

#### 4.5.5 *Seed*

Observation of seeds from herbarium specimens are limited as not all fruits were cut open. Seeds are axile, 2-8 per fruit, and one seed per locule. Each seed is enveloped by fleshy aril or endocarpic pulp (Ng, 1992) and in sect. *Garcinia* the colour of the aril is usually white.

## CHAPTER 5

### MOLECULAR SYSTEMATICS OF SECTION *GARCINIA*

#### 5.1 Introduction

##### 5.1.1 *Classification of Section Garcinia sensu Jones*

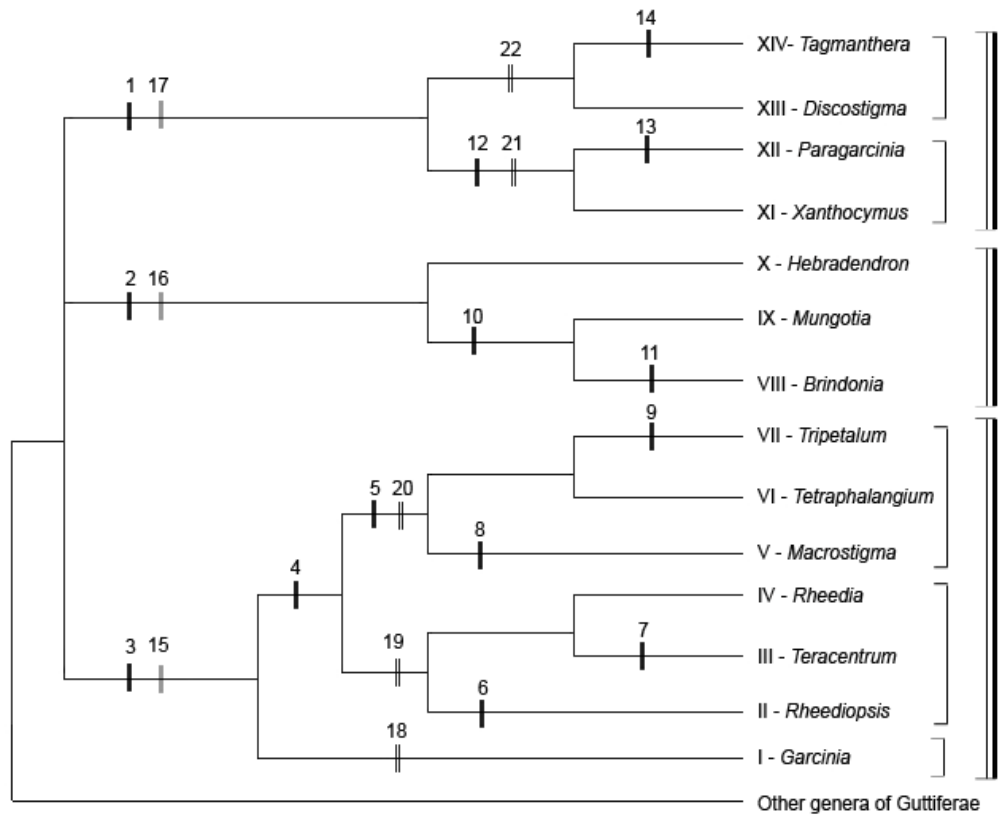
On evaluating the relationships of the sections in *Garcinia*, Jones (1980) proposed a hypothetical relationship based on morphology that is shown here in Figure 5.1, although she did not perform any formal cladistic analysis. Using anther characters, Jones first divided the representative species into three groups and further divided the groups into five subgroups based on stamen characters. These groups and subgroups were merely artificial and helped her to establish a sectional classification and they were not formally named.

The characters used to define the sectional classification are mainly taken from the male flowers. However, the over-reliance on these characters that this system adopted has led to some complications in the determination of both sectional and species limits within the genus. For example, for sect. *Garcinia* the use of stamen bundles, inserted in the middle of male flowers will lead to the inclusion of more species found on the islands of Madagascar and Fiji but the inflorescence and fruit characters of these species are not consistent with those of other members of the section.

Section *Garcinia* sensu Jones (1980) consists of 43 species that are distributed mainly in Southeast Asia although a few species are found on the Madagascar and Fijian islands, and in the eastern part of India. Morphological characters that united the section were based on the stamens which were described as a central mass either in the form of four connecting bundles or not, surrounding a fungiform pistillode; the anther was 2-thecous (Jones 1980).

The circumscription of sect. *Garcinia* by Jones (1980) contains a large number of species but there are no single characters that are unique to the section and shared by all of the members that could be considered synapomorphies. It is clear that use of either male or female flowers alone does not lead to a satisfactory infra-generic or species-level classification.

Given the problems associated with classification based solely on morphology, it is possible that molecular data may provide the information needed to identify significant morphological characters that could better determine species and sectional limits. Molecular data, in particular, DNA sequences have been used extensively in taxonomic studies to infer phylogenetic relationships or for classification purposes.



- Characters used for sectional classification
  - 1 - venation: eucamptodromous → reticulodromous
  - 2 - stipule : present → absent
  - 3 - inflorescence : terminal triad → solitary or thyrsoid
  - 4 - sepals : 5 → 4, 2, undivided calyx
  - 5 - petals : 5 → 6-8, 4, 0
  - 6 - stamen : partially united in bundles → completely united in bundles or annular mass, free
  - 7 - stamen : free from petals → adnate to petals
  - 8 - stamen : bundles antepetalous → bundles antesepalous
  - 9 - pollen : colpate → porate
  - 10 - pollen : ornamentation reticulate → ornamentation psilate, more complex
  - 11 - pistillode : present → absent
  - 12 - fasciodes : present → absent
  - 13 - staminodes : present → absent
  - 14 - ovary : number of locule equal to segment perianth → locules greater, locules fewer
  
- Characters used to divide groups
  - 15 - anther : 2-theous, ellipsoid, recurved, central connective
  - 16 - anther : 4-theous
  - 17 - anther : 2-theous, globose
  
- Characters used to divide sub-groups
  - 18 - stamen : united in 4-bundles or annular mass
  - 19 - stamen : partially united in 4-bundles or free stamens
  - 20 - stamen : united in bundles to the petals
  - 21 - stamen : united in 2,4 strap-shaped bundles
  - 22 - stamen : partially united in bundles

Figure 5.1. Hypothetical relationships of sections in *Garcinia* sensu Jones (1980). Tree originally drawn by Jones with some modification showing characters for her artificial groupings and subgroups. *Garcinia* was firstly divided into informal groups (characters 15-17) and subgroups (characters 18-22) then were divided to form sections (characters 1-14).

### 5.1.2 Molecular Studies of *Garcinia*

A key aim of this chapter is to use phylogenetic analysis of DNA sequence data to investigate the monophyly of sect. *Garcinia* and the relationships of its species. The application of molecular data for taxonomic purposes is perhaps best exemplified by the Angiosperm Phylogeny Group (APG, 2003). This group indicated that Guttiferae (Clusiaceae), the family to which *Garcinia* belongs, is placed in the order Malphigiales. Based on a phylogeny using *rbcL* sequences Guttiferae is paraphyletic if *Hypericum* is included (Gustafsson et al., 2002). *Hypericum* plus eight other genera was therefore split into its own family, Hypericaceae. The closest relatives of Guttiferae are the families Bonnetiaceae, Hypericaceae and Podostemaceae (APG, 2003). Plastid *rbcL* sequence data also indicate that the family Guttiferae consists of two subfamilies, Kielmeyeroideae and Clusioideae (Gustafsson et al., 2002). Gustafsson et al. (2002) placed *Garcinia* in the tribe Garcinieae that was placed in subfamily Clusioideae along with tribes Clusieae and Symphonieae.

*Garcinia* is placed in the tribe Garcinieae, but *Garcinia* is paraphyletic having *Rheedia* nested within it (Gustafsson et al., 2002). Gustafsson et al. (2002) also indicated that *Symphonia*, *Tripetalum*, *Pentaphalangium* and *Garcinia* formed a monophyletic group with moderate bootstrap support (67%). Greater sampling than that included in this study will be necessary to determine the monophyly of each of these genera.

Rismita-sari (2000) used internal transcribed spacer of ribosomal DNA (ITS) sequences to show that *Tripetalum*, *Pentaphalangium*, *Rheedia* were nested within

*Garcinia*. In her study, Jones's sectional classification was also evaluated indicating paraphyletic sects. *Brindonia* and *Xanthochymus*, while the rest of the sections including sect. *Garcinia* formed monophyletic groups. However, most of the species used in this study were of Indonesian and Australian origin and global sampling was therefore limited.

A regional approach for studying the phylogeny of *Garcinia* in tropical lowland forest of the Pasoh Forest Reserve and using 15 species sequenced for both the ITS and *trnL-trnF* regions indicated that five of Jones's (1980) sections (*Brindonia*, *Garcinia*, *Discostigma*, *Hebradendron* and *Xanthochymus*) formed a monophyletic group (Nazre, 2000). These sections represent the sections that are found in Peninsular Malaysia. Sect. *Garcinia* was shown to be closely related to sect. *Brindonia* while sect. *Xanthochymus* as in Rismita-Sari's (2000) studies is placed as sister to all other sections.

In contrast, ITS sequences of several species from Thailand, Malaysia and Indonesia (Yapwattanaphun et al., 2004) indicated that section *Garcinia* is a paraphyletic group because of the inclusion of *G. rostrata* and *G. sicyiifolia* (sect. *Discostigma* and sect. *Brindonia*, respectively, sensu Jones, 1980). The members of sect. *Garcinia* included by Yapwattanaphun et al. (2004) were *G. mangostana*, *G. malaccensis*, *G. speciosa* (a synonym of *G. celebica*) and *G. hombroniana* (also a synonym *G. celebica*). However, I believe that this outcome may be due to a misidentification. The text in this paper states that “the fruit of *G. rostrata* is quite similar to *G. hombroniana* except it is smaller and turns red when ripe” and no descriptions were given for *G. sicyiifolia*.

Morphologically, although the fruit of *G. rostrata* is smaller in size, (c. 1 cm across compared to *G. celebica* that is c. 4 cm across) the overall morphology for both species is distinctly different. For example the stigma of *G. rostrata* is small and discoid while with *G. celebica*, the stigma is raised or elongated and tipped with wavy lobes (Whitmore, 1973). The specimen identified as *G. rostrata* could therefore be *G. celebica* but no voucher specimens (for all species collected from BO in his study) were made by the authors and it is not possible to verify this. For *G. sizygiifolia*, there is also a possibility that this accession might be a misidentification of *G. rigida* due to the similarity of the sequence produced (98% similarity). Morphologically, *G. sizygiifolia* (sect. *Brindonia* sensu Jones) is distinctly different from *G. rigida* with the fruit nearly sessile, borne in axils, smaller in size at c. 1.5 cm long, and the stigma is sessile. The fruit of *G. rigida*, has a pedicel of 2.0 cm, is borne terminally, is c. 3.0 cm long and has a stigma raised to 3 mm long.

### 5.1.3 Hybridisation in *Garcinia*

Some species of *Garcinia* are believed to be of hybrid origin (Whitmore, 1973; Richards, 1990c). Evidence for this includes the capability of some *Garcinia* species to produce seed without fertilisation, a phenomenon defined as agamospermy (Gustafson, 1946-1947). Most agamosperms are highly heterozygous as hybrid species or hybrid derivatives (Grant, 1971), and at least ten wild species of *Garcinia* in Peninsular Malaysia are thought to be facultative agamosperms (Richards, 1990a). The most widely studied of

these species is mangosteen, perhaps an obligate agamosperm only found in cultivation (Richards, 1990a). Richards (1990c) suggested that mangosteen was a hybrid between *G. hombroniana* (now reduced to synonymy under *G. celebica*) and *G. malaccensis* based on morphological and cytological evidence. The combination of nuclear and plastid DNA sequences should provide evidence of hybridisation if tree topologies based on sequences from these separate genomes are different. They may also allow us to determine the parents of hybrid species as shown by studies in the family Caricaceae (Kyndt et al., 2005).

#### 5.1.4 Biogeography of Section *Garcinia*

Historically, the South East Asian (SEA) area was formed by processes beginning in the Palaeozoic and Mesozoic (540-65 million years ago) from assemblages of terranes that originated from the margins of Gondwana (Metcalf, 1998). These processes culminated in the mid-Miocene collision of the Sunda and Sahul (Australian) shelves. Some authors (e.g. Morley, 2000) have suggested that massive plant dispersal events across the border between these two shelves followed this geological event. Wallace (1863) described a distinct biogeographical discontinuity that more or less matched this line that has become known as Wallace's Line (Figure 5.2). This was subsequently modified by Huxley (1868). These biogeographical lines are indicated in Figure 5.2. Sect. *Garcinia* sensu Jones (1980) is distributed mainly in the SEA area on the west side of both Wallace's and Huxley's Lines. This is consistent with the section originating in

the west. This assumption will be tested using molecular data. If the eastern species are nested within those from the west then this is evidence for origin in the west with recent migration to the east. Dating of the molecular phylogeny will also indicate the timing of dispersal across Wallace's line.



Figure 5.2 The position of Wallace's Line and Huxley's Line.

## 5.2 Objectives

The first objective of this study is to test the monophyly of section *Garcinia* sensu Jones (1980). For that purpose, any available *Garcinia* species including sequences already submitted to GENBANK will be included in the phylogenetic analysis. Secondly, molecular data will be used to study the relationships of species within sect. *Garcinia*. Thirdly, this study will include multiple accessions of some of the individual species that will allow to test whether the species delimited in previous chapters are distinct taxonomic entities. Finally, the aim of this study is to determine whether any species in section *Garcinia* might be of hybrid origin and elucidate the biogeographic history of the section.

## 5.3 Materials and Methods

### 5.3.1. Region Chosen

The internal transcribed spacer of nuclear ribosomal DNA (ITS) is one of the most widely used regions to infer phylogeny at low taxonomic ranks. The two internal transcribed spacers evolve more rapidly than coding regions and were suggested to have great potential for studies among closely related species (Baldwin et al., 1995). This region has been used in previous studies in *Garcinia* (Nazre, 2000; Rismita-Sari, 2000; Yapwattanaphun et al, 2004), that indicated that it was sufficiently variable to resolve

relationships amongst species and ITS was therefore chosen for this study. The ITS region consists of two spacers, ITS1 and ITS2 flanked by the coding regions of 18S and 26S, with the 5.8S exon in the middle. The sequenced lengths of ITS regions for *Garcinia* are between 599-630 base pairs (bp) (Nazre, 2000; Rismita-Sari, 2000; Yapwattanaphun et al., 2004).

To corroborate any phylogenetic hypotheses based on a nuclear marker it is necessary to utilise other markers, preferably from another genome. Comparison of trees based on analysis of nuclear and plastid markers should also provide evidence for hybridisation as plastid markers are usually uniparentally inherited. Plastid DNA is maternally inherited, generally evolves at a slower rate compared to nuclear DNA (Palmer et al., 1998; Shaw et al., 2005). Two markers from non-coding regions of chloroplast DNA (cpDNA) were chosen, *trnS-trnG* and *trnD-trnT*. Both regions occur in the Large Single Copy (LSC) region of the chloroplast genome. Based on the *Nicotiana* chloroplast genome (Wakasugi et al., 1998), the *trnS-trnG* region is located within and between the *trnG* and *trnS* exons, and consists of the *trnG* intron and *trnS-trnG* spacer. The *trnG* intron has an average length of 763bp and ranges from 697-1000 bp, while the *trnS-trnG* spacer length averages 763 bp and ranges from 619-1035bp (Shaw et al., 2005). The *trnS-trnG* intergenic spacer was found by (Xu et al., 2000) to be the most informative cpDNA non-coding region within two closely related subgenera of *Glycine*. The *trnS-trnG* regions have been demonstrated to have phylogenetically informative variation even within species in *Corythophora* (Hamilton et al., 2003), *Moringa* (Olson, 2002) and *Lathyrus* (Kenicer et al., 2005). The *trnD-trnT* spacer is less widely used to infer

phylogenetic relationships. However, significant variation has been observed in studies of *Allium* (Friesen et al., 2000) and Arecaceae (Hahn, 2002). The average length of the *trnD-trnT* intergenic spacer is 1066 bp, ranging from 578-1403 bp (Shaw et al., 2005).

### 5.3.2. Taxonomic Sampling

Sources of plant material and voucher information used in this study are listed in Appendix II and a summary of the number of taxa sequenced for each gene is shown in Table 5.1 below. In this study, a total of 83 accessions representing ten species of section *Garcinia* (13 species out of 43 species sensu Jones) were sequenced for ITS, plus 53 accessions representing 32 species of 8 other sections of *Garcinia*. However, the accessions and species numbers sequenced for *trnS-G* and *trnD-T* are fewer compared to the ITS region because of time constraints. A total of 51 and 25 accessions of *trnS-G* and *trnD-T* regions were sequenced, respectively, with nine representatives of sect. *Garcinia* (Table 5.1). In addition, 41 ITS sequences were taken from GenBank (see Appendix III for the complete list of all species) with only five species belong to sect. *Garcinia* from a total of 29 species. These additional GenBank sequences were sampled predominantly from other sections in *Garcinia* in order to test the monophyly of sect. *Garcinia*.

Table 5.1. Summary of the accession and sequence numbers used in this study.

	<b>ITS</b>	<b><i>trnS-trnG</i></b>	<b><i>trnD-trnT</i></b>
No. of accessions sequenced	83	50	24
No. of sect. <i>Garcinia</i> accessions sequenced	30	23	17
No. of species sequenced	42	30	16
No. of sect. <i>Garcinia</i> species sequenced	10	9	9
(species no. sensu Jones, 1980)	(13)	(12)	(9)
No. of accessions from GENBANK	41	-	-
No. of sect. <i>Garcinia</i> accessions from GENBANK	26	-	-
No. of species from GENBANK	29	-	-
No. of sect. <i>Garcinia</i> species from GENBANK	5	-	-
(species no. sensu Jones, 1980)	(6)		
<b>Total no. of Accessions</b>	124	51	25
<b>Total no. of Species</b>	58	30	17
<b>Total no. of species in Sect. <i>Garcinia</i></b>	10	9	9

### 5.3.3. Outgroup Taxa

Another advantage of using ITS was the availability of sequences on GenBank that could be incorporated into the analysis and used as outgroups. Species of *Clusia* were used as the outgroup because previous studies indicated that this genus was related to *Garcinia*. The genus *Clusia* is neotropical with about 200 species, and grouped together with *Garcinia* and other genera within the subfamily Clusioideae (Stevens, 2001 onwards). Other close relatives of *Garcinia* such as *Rheedia*, *Symphonia* and *Tripetalum* were not deemed suitable because previous studies (Rismita-Sari, 2000) indicated that they were nested within *Garcinia*. The lack of availability of material of GenBank *trnS-G* or *trnD-T* sequences for *Clusia* or other close relatives of *Garcinia* meant that alternative outgroups had to be used for the analyses of the chloroplast data sets generated here. In

these cases our main focus was to determine the monophyly of section *Garcinia* and we therefore sequenced species of *Garcinia* that were indicated to be basally divergent in the ITS phylogeny and used those as outgroups.

#### 5.3.4. DNA Extraction

Approximately 1.0g of leaf tissue from silica gel dried or herbarium material was extracted using the CTAB (Cetyltrimmonium bromide) extraction buffer following the protocol of Doyle & Doyle (1990). However, for herbarium specimens, during the isopropanol step, the precipitation period was increased up to 40 days at -20°C because this has been shown to increase yield in other studies (e.g. Richardson et al 2000). In some cases, problematic samples with low yield or badly degraded DNA were also extracted using the Dneasy Plant Mini kit (QIAGEN, California USA) following the manufacturers protocol. Detailed steps on the protocol used for DNA extraction are found in Appendix IV. The quality and quantity of the DNA yielded was estimated by running it on a 1% agarose gel and comparing with a 1Kb+ DNA ladder (Invitrogen).

#### 5.3.5. Amplification

PCR was performed either using a Perkin Elmer or MJ Research thermal cycler, with 30-35 amplification cycles (1 min. denaturation at 94<sup>0</sup> C, 1 min. annealing at 48-66<sup>0</sup> C, 2 min. of extension at 72<sup>0</sup> C) in 25 µl volumes with the following reaction

components: 1-2 µl template DNA (10-100ng), 2.5 µl 10x PCR buffer (Bioline Ltd.), 1.25 µl MgCl<sub>2</sub> (50mM), 2.5 µl dNTP 2mM (200 µM each dATP, dTTP, dGtp and dCTP), 1.5 µl primers (10 µM each) and 0.125 µl Taq Polymerase (Biotaq Ltd.). Reaction volume was adjusted to 25 µl by adding deionised water. The primers used to amplify each region are listed in Table 5.2 below.

The PCR products were checked on a 1% agarose gel for quality, while the remainder of the products were purified with either Qiaquick PCR purification kit (Qiagen Ltd.) or GFX PCR purification kit (Amersham Bioscience), following the manufacturer's instructions. To check the quality and quantity of the purified DNA template, a 1% agarose gel electrophoresis was again performed prior to sequencing.

Table 5.2 Primers used for amplification and sequencing.

Primer name	Sequences	Annealing Temperature
1. <i>trnS-trnG</i> (Shaw et al., 2005)	<i>trnS</i> AGAT AGGG ATTC GAAC CCTC GGT <i>trnG</i> GCGG GAAT CGAA CCCG CATC	48 <sup>0</sup> - 66 <sup>0</sup> C
2. <i>trnT-trnD</i> (Demesure et al., 1995; Shaw et al., 2005)	<i>trnD</i> ACC ATT TGA ACT ACA ATC CC <i>trnT</i> CTA CCA CTG AGT TAA AAG GG <i>trnE</i> AGG ACA TCT CTC TTT CAA GGA G <i>trnY</i> CCG AGC TGG ATT TGA ACC A	48 <sup>0</sup> C-55 <sup>0</sup> C
3. ITS (White et al., 1990)	ITS2 GCAT CGAT GAAG AACG TAGC ITS3 GCTA CGTT CTTC ATCG ATGC ITS4 TCCT CCGC TTAT TGAT ATGC ITS5 GGAA GTAA AAGT CGTA ACAA GG	48 <sup>0</sup> C

### *5.3.6. Sequencing*

Cycle sequencing was performed with the CEQ™ Dye terminator Cycle Sequencing (DTCS) Quick Start Kit (Beckman Coulter Inc., USA). Each sequence reaction contains 10 µl volume of DTCS Quickstart mix (4 µl), 10pM primer (1 µl), purified DNA template (0.5-5 µl) and deionised water (0.5-5 µl). The thermal cycle parameters were 35 cycles of 96<sup>0</sup>C denaturation (20 sec.), 50<sup>0</sup>C annealing (20 sec.) and 60<sup>0</sup>C extension (4 min.).

The cycle sequencing products were then cleaned using an Ethanol clean up procedure (see Appendix IV) before electrophoresis and analysis on the CEQ™ 8000 Genetic Analysis System (Beckman Coulter Inc., USA).

### *5.3.7. Sequence Alignment*

Raw sequence data were analysed individually by CEQ Sequence Analysis (Beckman Coulter Inc., USA), assembled for both forward and reverse sequences to produce contig files in Sequencher™ (Genecode, USA) that were checked and corrected manually. Sequences were then exported to BIOEDIT Ver. 7.0.2 (Hall, 1999), and aligned using CLUSTAL X (Thompson et al., 1997). Sequence alignments were then refined and checked manually using BIOEDIT.

Gaps in the aligned sequences of cpDNA, were coded using GAPCODER (Young & Healy, 2003) using the simple indel coding method described by Simmons and Ochoterena (2000) in which gaps are coded as present or absent. Complete aligned sequences with indel coded site/regions are presented in Appendix VI and VII. Ambiguous and microsatellite regions which are too difficult to align were excluded from the analysis and are also indicated in Appendix VI and VII. Output of the NEXUS file from GAPCODER was incorporated into the sequence matrices and used for analysis in PAUP and MRBAYES (see below). Three different datasets were prepared for the analyses; 1) ITS, 2) *trnS-trnG*, 3) combined *trnS-G* and *trnD-T* (combining the two plastid sequence data sets can be justified because as chloroplast regions they are part of the same non-recombining molecule).

### 5.3.8. *Phylogenetic Analysis*

#### 5.3.8.1. *Choice of Methods for Phylogenetic Inference*

For any given set of aligned DNA sequences there may be a number of possible phylogenetic tree topologies. Optimality criterion approaches put a score on every possible tree, and then select the tree(s) with the best score; the parsimony criterion is based on tree-length (minimizing the total number of substitutions through the tree) and the likelihood criterion is based on a model of DNA sequence evolution. In a parsimony analysis, ideally all possible trees generated from the alignment will be assessed to

determine the shortest (most parsimonious) one. However, since the number of trees generated increases exponentially with the addition of each new sequence, a large dataset (c. >20 sequences) will make it computationally impossible to assess every tree. Heuristic search strategies are therefore employed to reduce the number of trees that need to be assessed. The search strategy employed here is described in section 5.2.9.2 below.

Parsimony is straightforward to understand and efficient in the sense of computational time. Maximum likelihood is an optimality criterion that determines the likelihood of a data set given a tree and a model of molecular evolution. If given some data,  $D$ , and a hypothesis  $H$ , the likelihood of that data is given by:

$$LD = \Pr(D|H)$$

which is the probability of obtaining  $D$ , given  $H$ . In the context of molecular phylogenetics  $D$  is the set of sequences being compared and  $H$  is a phylogenetic tree. The idea is not to estimate the phylogeny but to estimate the likelihood of observing the data  $D$  under certain specified conditions. Those conditions are a particular tree topology and a particular model of character evolution. The tree that makes our data the most probable evolutionary outcome is the maximum likelihood estimate of the phylogeny. Maximum likelihood searches are more complex than parsimony searches as they are model-based and consequently take a great deal more computer time. However alternative model-based approaches can be used to estimate phylogeny, as is done in the analyses presented in this chapter.

Bayes theory determines the probability of an evolutionary tree based on the sequence data. The analysis starts with a loose assumption or prior expectation about the tree topology, branch length and the evolutionary model used to assume the correct tree. By examining the alignment, it then improves the confidence of the tree topology (clades) by calculating the confidence or posterior probabilities using Bayes formula. However, to calculate the probability of every tree topology (clades) with every model is computationally impossible and a sampling method called Metropolis coupled Markov chain Monte Carlo (MCMC) is used where the aim is to efficiently search tree space for the most likely trees. The Markov chain starts the searches at a random tree and evolutionary parameters, and calculates the probability (likelihood). It then proposes a new state with modification of one of the parameters (either topology, branch length or evolutionary model), re-calculates the probability of the new state and either accepts or rejects it. Usually four chains are run at a time. Each chain event is called a generation and several thousands to millions of generations are run. Of the four chains, the chains that sample the trees are called the 'cold' chain and the rest are 'heated' chains. The heated chains more readily allow swapping of states. This strategy increases the chances of finding the globally optimal tree without getting stuck in one particular (locally optimal) tree space area.

An advantage of Bayesian Analysis over maximum likelihood is that it can partition data sets and determine different models of evolution for each one, i.e. data sets that contain two markers that might have different patterns of molecular evolution can be analysed under different models simultaneously. In terms of computing times, Bayesian

analyses are also more efficient than a maximum likelihood approach. Other advantages include the fact that the analyses are more explicit by using all available parameters, confidence values of each clade are given with posterior probabilities, and the results are probability values of the sampled trees and evolutionary models (rather than single probability of the evolutionary model as in Maximum Likelihood). For all of these reasons I chose to use Bayesian analyses in addition to the parsimony analyses for this study.

#### 5.3.8.2. *Maximum Parsimony*

Analyses to find the most parsimonious trees were run using PAUP\* Ver. 4.0b10 for Windows using 'Heuristic' methods under the Fitch (1971) parsimony criterion where character state changes were weighted equally and unordered. Character state change is optimised with accelerated transformation (ACCTRAN; (Swofford & Olsen, 1990) where the changes occur as early in relation to the root as possible.

Two rounds of heuristic searches were performed. The first search involves multiple replicates each of which has two stages: (i) the initial tree is obtained by connecting the taxa one at a time using stepwise addition and additional taxa are added in random order; (ii) to search for more parsimonious trees, branches were swapped using tree bisection reconnection (TBR). Only 10 shortest trees per replicate found during the swapping steps were saved to avoid searching for too long on suboptimal islands. All the

shortest trees from 10,000 replicates will be saved and used for a second round of searches. The second round of heuristic search starts with the shortest trees that were obtained from the first search. These trees were branch-swapped to completion or swapped on until 10,000 trees were produced, at which point the number of trees was limited and these were swapped to completion. To evaluate the confidence values of the clades, ten thousand replicates of the bootstrap (Felsenstein, 1985) were then carried out with simple addition of sequences, TBR branch swapping and holding five trees at each replicate. All trees produced from the parsimony search were visualised using TreeView 1.6.6 (Page, 1996).

#### 5.3.8.3. *Bayesian Analysis*

Bayesian analyses were performed using MRBAYES ver. 3.1.1 software for Windows (Ronquist & Huelsenbeck, 2003). A prior probability of the simplest model to fit the aligned sequence data was estimated by MrModeltest 2.2 (Nylander, 2004) using the Hierarchical Likelihood Ratio Test (hLRT). Each sample was run for a minimum of three million generations and four Markov chains running simultaneously. Running was stopped as data converged on the stationary distributions, assessed by viewing when the average standard deviation of split frequencies reaches less than 0.01 (Ronquist et al., MrBayes Manual ver. 3.1). The phylogenetic trees produced from Bayesian Inference were summarised by calculating the 50% majority-rule consensus tree after excluding the 'burn-in' trees. The burn-in trees were estimated by plotting (plot command) the

'Inlikelihood' against number of generations. The posterior probabilities produced are used as an estimation of the degree of robustness for the clades. In the combined Bayesian analysis each gene was treated as a separate partition.

#### 5.3.9. *Incongruence Length Difference Test*

Two plastid DNA datasets (*trnS-G*, combined *trnS-G* and *trnD-T*) were compared against the ITS dataset using the incongruence length difference (ILD) test (Farris et al., 1994) to determine whether these datasets were incongruent. The ILD test compares the number of steps required for minimum-length trees in separate, combined and random partitions of the data (Cunningham, 1997). The ILD test is performed in PAUP Ver. 4.0 (Swofford, 2003 ) using the Partition Homogeneity Test between 60 accessions of ITS against *trnS-G* sequences, and 32 accessions of ITS and cpDNA (*trnS-G* and *trnD-T*) sequences.

#### 5.3.10. *Age Estimation*

The ITS sequence data was used to determine age estimates for nodes within the ITS trees. Rate heterogeneity among lineages was first evaluated using the likelihood ratio test, which compares log likelihoods of both constrained and unconstrained hypotheses on one of the trees from the parsimony analysis. The molecular clock was rejected because the constrained and unconstrained analyses were significantly different,

so Sanderson's methods of nonparametric rate smoothing (NPRS; Sanderson, 1997) was applied to estimate divergence times using the software package r8s (Sanderson, 2003). The standard deviation for the NPRS derived dates was also estimated by keeping the tree fixed and bootstrapping the data set 100 times using PAUP. The resulting trees were then processed in r8s to calculate the standard deviation of divergence times for selected nodes. A 90 MY fossil, *Paleoclusia*, described by Crepet and Nixon (1998) from the Late Turonian was used to constrain the age of the node representing the split (most recent common ancestor) between *Clusia* and *Garcinia* as it is morphologically similar to both of these genera, and the estimated date for the crown of *Garcinia* and nodes within sect. *Garcinia* were calculated. Of particular interest would be any node representing the most recent common ancestor of species found to the east of Wallace's Line.

An additional, though somewhat crude approach to dating involves application of rates determined from other taxa, an approach used by Richardson et al. (2001a). Using the program DNAsp 4.0 (Rozas & Rozas, 1999), the mean number of substitutions in the ITS tree from tip to root within the *Garcinia* clade were calculated. The mean was divided with the published substitution rates (number of substitution per site per year) cited in Richardson et al. (2001a) to yield estimated dates for the crown node of *Garcinia*.

## 5.4 Results

### 5.4.1 ITS data

The aligned ITS sequence data contains 240 (39%) informative characters out of 615 characters used. Only a small number of indels are created to accommodate length differences between taxa. The largest was a nine base pair insertion found in *Symphonia globulifera* at position 470 to 479 (see Appendix V). Table 5.2 summarises sequence information, analysis parameters and tree characteristics from the Maximum Parsimony and Bayesian Inference analyses.

In the maximum parsimony (MP) analysis, the first round of heuristic searches produced 85 most parsimonious trees and the second round resulted in more than 10,000 most parsimonious trees. Figure 5.3 shows one of 10,000 trees with a tree length of 998 steps, CI=0.50, RI=0.88 and HI=0.51.

The Bayesian inference analysis (BI) of the ITS dataset produced a fifty percent majority rule consensus tree (Figure 5.4) that was almost the same as the strict consensus tree from the parsimony analysis. The main difference is in the placement of clades **D** and **E**, where in MP these form a clade that is the sister group of clade **A**, **B** and **C** but in BI, clade **D** is sister to a clade consisting of clade **A**, **B**, **C**, **F** and **G** while clade **E** is sister to the clade consisting of groups **A**, **B**, **C**, **D**, **F** and **G**. BI also exhibits higher probabilities than the bootstrap support values obtained with MP, a common phenomenon as described

by Huelsenbeck et al. (2002). However, these are two entirely different measures of support and are therefore not directly comparable.

Table 5.3. Summary of sequence information, analysis parameters and tree characteristics from Maximum Parsimony and Bayesian Inference analyses. CI = Consistency Index, RC = Rescaled Consistency Index, HI = Homoplasy Index, RI = Retention Index, hLRT = Hierarchical Likelihood Ratio Test, GTR+I+G = General Time Reversible with Invariant sites Estimated and a Gamma distribution of rates, HKY +G= Hasegawa, Kishino & Yano and a Gamma distribution of rates.

	ITS	<i>trnS-G</i>	<i>trnD-T</i>	<i>trnS-G + trnD-T</i>
No of Taxa	128	50	24	24
No. of Characters	615	1253	1325	2667
No. of Gap Coded Characters	-	53	30	58
G+C Contents (%)	51.34	23.5	32.75	-
<i>Maximum Parsimony</i>				
No. of Informative Characters	240	109	-	149
Tree Length	998	481	-	491
CI(RC)	0.50(0.44)	0.83(0.79)	-	0.88(0.70)
HI(RI)	0.51(0.88)	0.18 (0.79)	-	0.12(0.80)
<i>Bayesian Analysis</i>				
Prior Prob. of Best Suit Model (hLRTs)	GTR+I+G	HKY+G	HKY+G	-
Gamma Distribution (mean) [alpha]	0.19	0.92	0.43	-
Proportion of Invariable Sites [pinvar]	0.017	0.14	0.63	-
Base State frequency (mean):				-
pi (A)	0.24	0.38	0.34	-
pi (C)	0.23	0.11	0.16	-
pi (G)	0.25	0.14	0.16	-
pi (T)	0.29	0.37	0.35	-
Substitution Rate:				-
R [G-T]	0.07	0.14	0.14	-
R [C-T]	0.38	0.25	0.19	-
R [C-G]	0.06	0.14	0.12	-
R [A-T]	0.09	0.11	0.14	-
R [A-G]	0.28	0.16	0.27	-
R [A-C]	0.11	0.20	0.24	-

Sect. *Garcinia* sensu Jones (1980) is paraphyletic with most of the species in clade **A** and the rest in clade **G**, with 100 and 79 percent bootstraps values (BV), and with 100 and 96 percent posterior probabilities (PP), respectively. In this study, only species in clade **A** are considered to belong to sect. *Garcinia* (more discussion of this clade can be found in section 5.5.3). In both the MP and Bayesian analyses some moderately to strongly supported monophyletic clades were consistent with Jones's (1980) sectional classification.

Clade **A** is a highly supported group, with 100% bootstrap values (BV) and posterior probabilities (PP). In the parsimony analysis, *G. echinocarpa* is sister to the clade and in the Bayesian analysis it is unresolved in a polytomy with clades **A**, **B** and **C**, **D**, **F** and **G** and its relationship to other clades is therefore not clear. According to Jones (1980), *G. echinocarpa* belongs to sect. *Brindonia* (clade **F**), but she acknowledged the morphological differences from other species of sect. *Brindonia* by its scaly-walled fruits and *G. echinocarpa* is also the only species from sect. *Brindonia* found endemic in Sri Lanka while the rest of the species in sect. *Brindonia*, are found from India to New Guinea.

Clade **B** is moderately supported (75% BV, 95% PP) and consists of *G. urophylla* and related species confirming that Jones (1980) sect. *Hebradendron* is a natural group with distinct free or united stamens in a column. Both analyses of MP and PP show a close relationship (100% supported BV and PP) with clade **C**. Clade **C** resolved by BI is consistent with Jones, (1980) definition of sect. *Macrostigma* and contains two sub-

clades. The first sub-clade (76% BV and 99% PP) consists of *G. latissima* and related species from the east of Wallace's lines in Sulawesi, Maluku and Papua, and the second sub-clade (100% BV and PP) contains species (*G. sp. M6*) from the western side of Wallace's line in Peninsular Malaysia. These two sub-clades are not monophyletic in the MP strict consensus tree but BI analysis moderately supported (76%) their unification into sect. *Macrostigma* sensu Jones (1980). However, the inclusion of *G. sp. M6* in sect. *Macrostigma* merits further investigation.

Clade **D** is a monophyletic group corresponding to sect. *Discostigma* sensu Jones (1980), and is highly supported by MP analysis (90% BV) and BI (100% PP). Clade **E** however, consists of species from various sections (sensu Jones) and is only moderately supported (91% PP with less than 60% bootstrap support). Jones (1980) placed *G. cymosa* (*Tripetalum cymosum*) in sect. *Tripetalum*, *G. prainiana* in sect. *Xanthochymus* and *G. picrorhiza* in sect. *Discostigma*. There is a possibility that there was an error in the last two species identifications as both rely on secondary data and sterile herbarium specimens that are difficult to identify. If these two species are incorrectly identified and omitted from the analysis, this clade would correspond to sect. *Tripetalum* sensu Jones (1980).

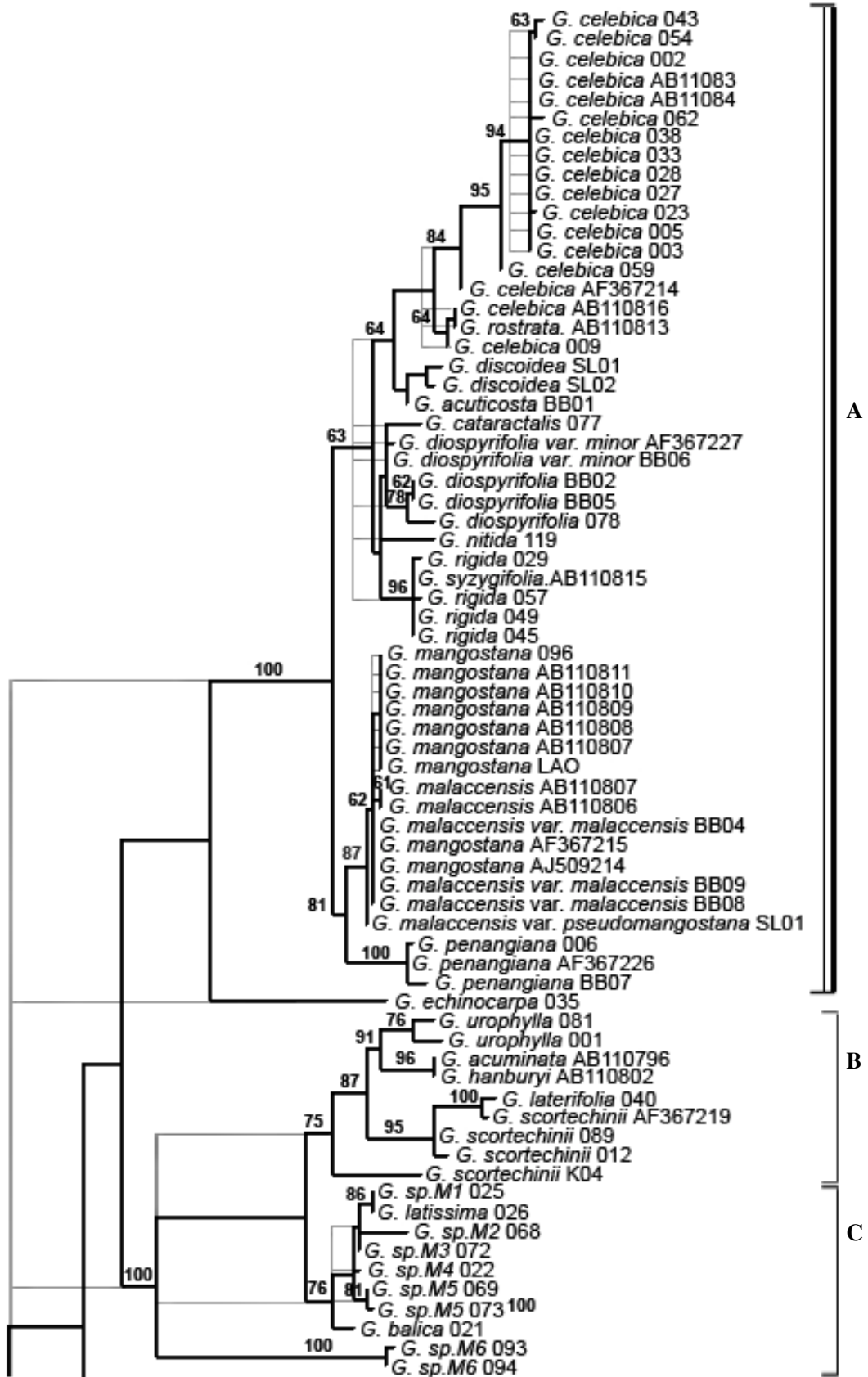
Clade **F** is consistent with sect. *Brindonia* sensu Jones (1980), and is significantly supported (93% BV and 100% PP) and is one of the biggest sections in *Garcinia* with 77 species distributed mainly in Southeast Asia. However, within the clade, the identification of *G. porrecta* (GenBank accession AB110812, Yapwattanaphun et al. 2004) is dubious

as the name has been used illegitimately for *Garcinia celebica* in Bogor Botanic Garden (BO), Java. This material was collected from the same locality as my own collection (tree number VI.A.4 from BO) but no herbarium voucher is available. The collections from BO have two different species labelled as *G. porrecta* (accession no 062) and *G. porrecta* var. *schizogyna* (accession no 049 and 057) and were identified in this study as *G. celebica* and *G. rigida*, respectively, and both species are placed in clade **A**, i.e. sect. *Garcinia* here.

Clade **G** receives 79% BV and 96% PP and consists of species such as *G. maingayi* which were regarded by Jones (1980) as members of sect. *Garcinia* but are indicated here to be a sister clade of sect. *Brindonia* (clade **F**) with moderate to strong support (66 % BV, 98% PP). Clade **G** and clade **F** (sect. *Brindonia*) share similar morphology with segmented fruits.

Clade **H** is a strongly supported group (99% BV, 100% PP) where most of the species belong to sect. *Xanthochymus* sensu Jones (1980). However, as in clade **F**, there are some issues regarding the identification of species whose sequences were obtained from GENBANK. For instance, *G. costata* and *Tripetalum cymosum* (*G. cymosa*) will need verification in order to confirm placement of these two species in this clade. This is because accessions with the same identification were positioned in clades **E** and **G**, respectively. In addition, the placement of accessions of *G. megaphylla* (one in clade **H** and the other in clade **I**) within two different clades also needs to be studied further by verifying the identity of the voucher specimens.

The most basally divergent clade **I**, a highly supported group (100% BV and PP) consists mainly of members of sect. *Rheedia* sensu Jones (1980), represented by mostly South American species. Within this, there is a further possibility of misidentification for *G. latissima* (placed in sect. *Tripetalum* by Jones). Another problematic clade is that of *Symphonia*, here represented by *Symphonia globulifera*. This species is placed as sister to clade **H** in MP (with no support) but in the BI analysis, the clade is indicated, with low PP, as sister to a clade consisting of clades **A** to **G**. The nesting of both *Symphonia globulifera* and *Tripetalum cymosum* within *Garcinia* is well supported in all analyses of ITS data.



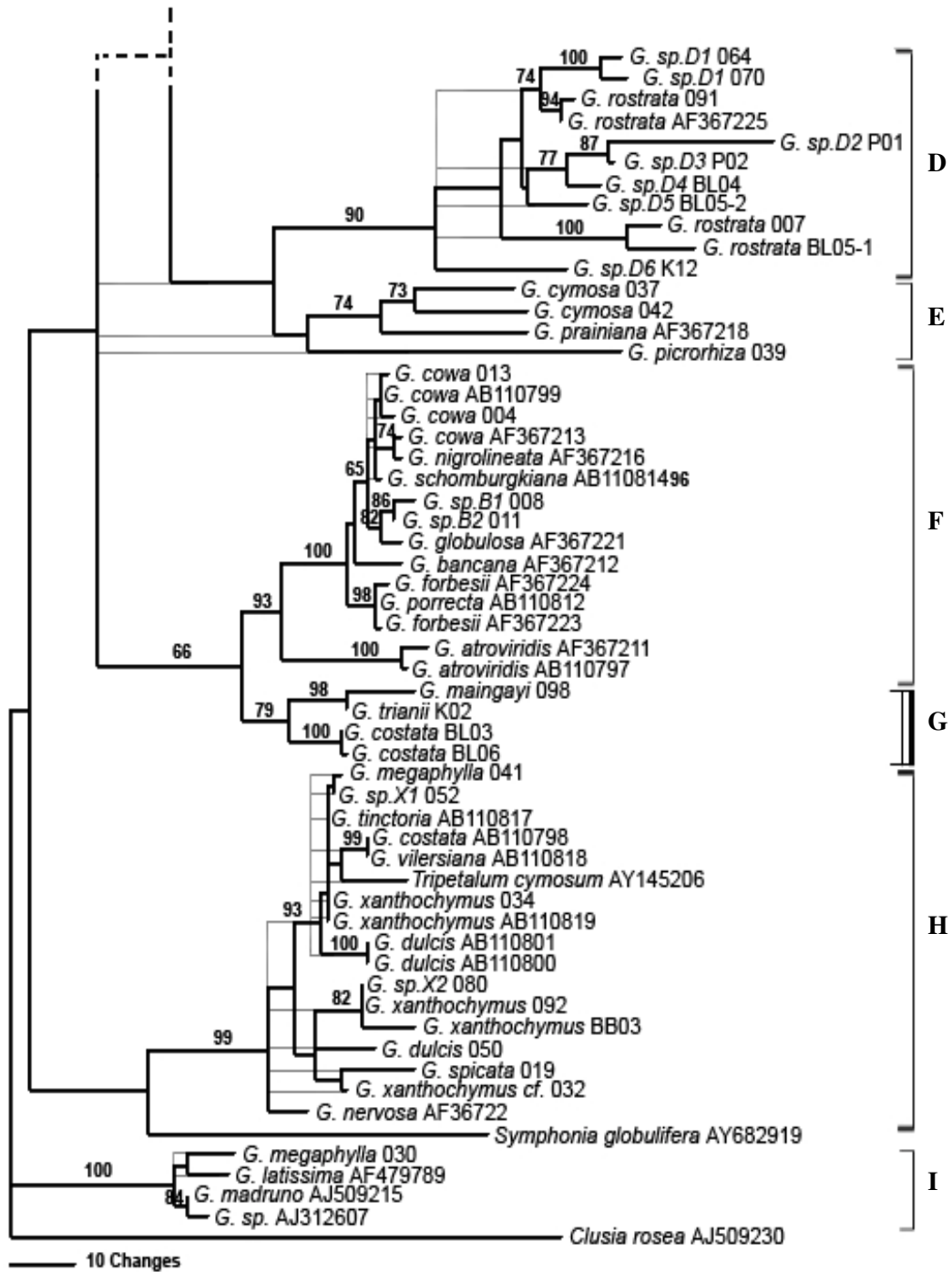
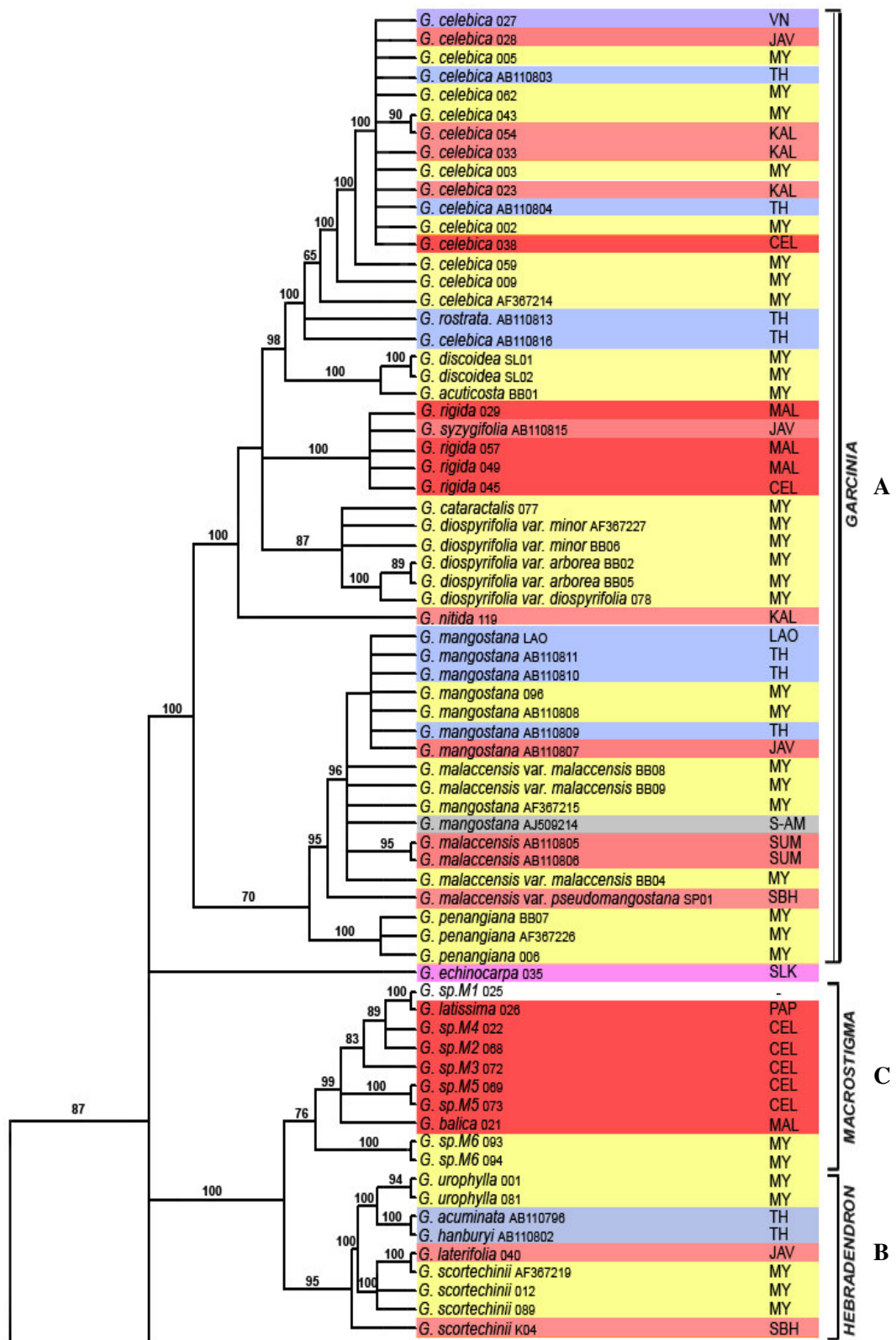


Figure 5.3. One of 10,000 most parsimonious trees from the analysis of the ITS data. Numbers above branches indicate bootstrap values (shown for values above 60% only). The grey branches indicate the topology of the strict consensus tree. Tree length = 998 steps, CI = 0.50, RI = 0.88 and HI = 0.51. The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980).



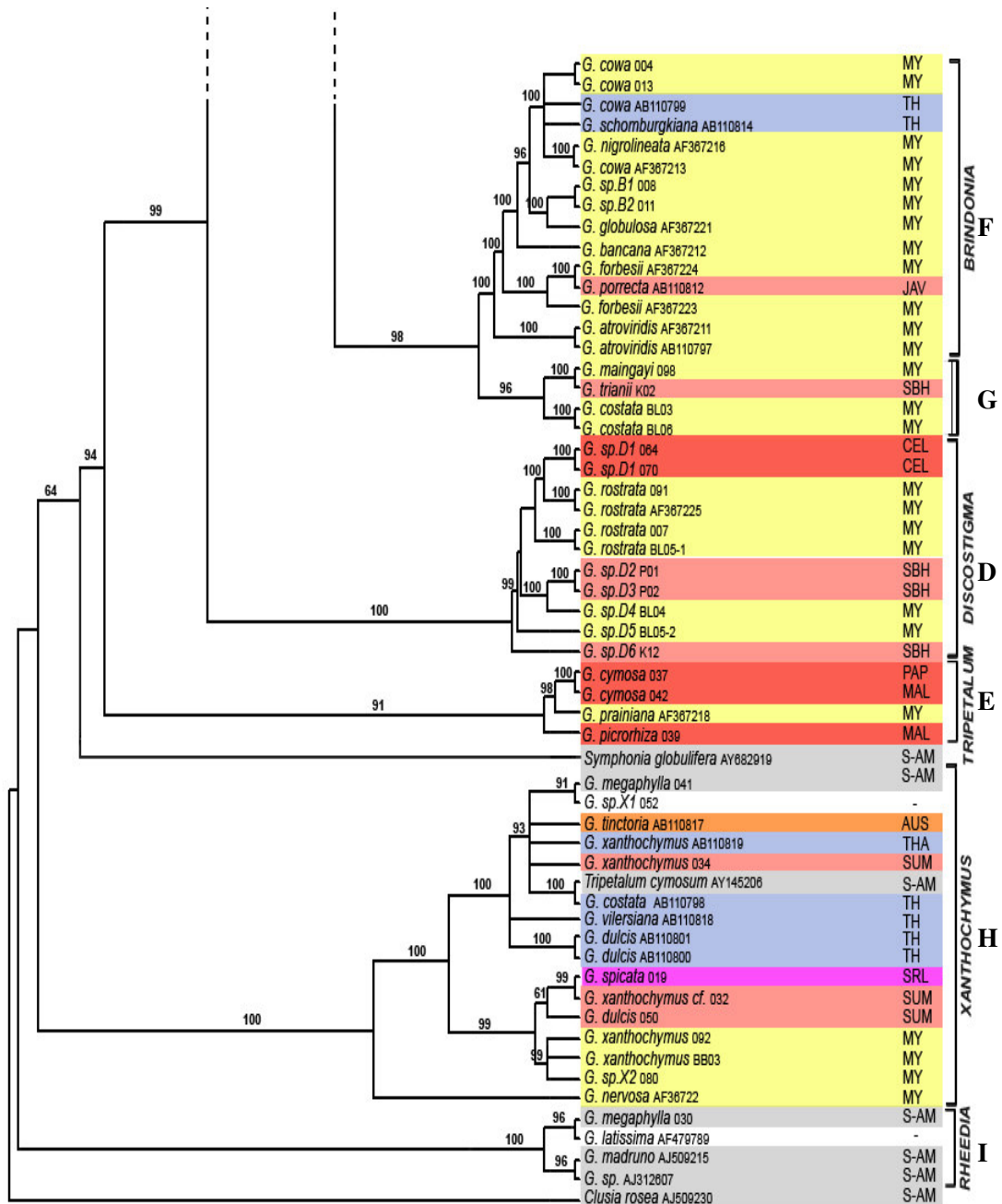


Figure 5.4. Fifty percent Majority Rule consensus tree resulting from the Bayesian analysis of ITS. Numbers above branches indicate posterior probabilities (shown for values above 60% only). The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980). Origin of samples collected; CEL=Sulawesi, JAV=Java, KAL=Kalimantan, LAO=Laos, MAL=Maluku, MY=Peninsular Malaysia, NWG=New Guinea, S-AM=South America, SBH=Sabah, SRL=Sri Lanka, SUM=Sumatra, TH=Thailand, Vn=Vietnam; Purple=Sri Lanka, Blue=Indochina & Thailand, Yellow=Peninsular Malaysia, Red=Malay Islands, Orange=Australia, Grey=South America and White=unknown origin.

#### 5.4.2 *trnS-trnG*

Aligned *trnS-G* sequences resulted in a total of 1253 characters of which 109 (8.70%) were potentially phylogenetically informative. However, there is great variation in the length of the sequences among taxa which resulted in many phylogenetically informative indels being coded from the sequence alignment, such as the large indel (403 bps) found from position 226 to 716 (Appendix VI). In total, 53 indels and 65 informative substitutions were used in the analysis. The first round of heuristic search resulted in a total of 1408 trees and the second round yielded more than 10,000 most parsimonious trees. The tree length was 481 steps and the CI (0.83) was higher than for the ITS region but the RI (0.79) and HI (0.18) were lower compared to ITS.

Because of the decreased taxon sample size and lack of variability, not all clades that were evident in the ITS analysis are evident in the plastid DNA analyses (Figure 5.4 and 5.5). MP and BI analyses show some dissimilarity in the position of infrageneric clades and most clades are not highly supported by BV (less than 90%) but moderately to highly supported by PP. However, the topologies of the MP and BI trees are broadly similar.

As with ITS, sect. *Garcinia* sensu Jones is paraphyletic, with taxa from the section found in clades **A-1** and **G** and also in clade **A-2** (*G. rigida*). Clade **A-1** is largely similar to clade **A** in the ITS trees, but is not strongly supported by the BV (<60%), though moderately supported by PP (93%). *Garcinia celebica* which is monophyletic in ITS trees

is paraphyletic in *trnS-G* trees and other clades within **A-1** are collapsed. Interestingly, one clade, **A-2** (consisting of accessions of *G. rigida* with strong support of BV [93%] and PP [99%]) previously nested within clade **A** in the ITS trees is outside this clade.

In the MP analysis, clades **D**, **E** and **F** are strongly supported. BI gives more resolution and higher support values as for clade **H**. The four clades (**D**, **E**, **F** and **H**) correspond to the sections *Discostigma*, *Tripetalum*, *Brindonia* and *Xanthochymus* sensu Jones (1980), respectively. However, sections *Macrostigma* and *Hebradendron* sensu Jones (1980) are not monophyletic in the *trnS-G* analysis with representatives of each being unresolved with respect to one another in the clade **B/C**. The monophyly of clade **B/C** is not strongly supported by BV (<60%) but strongly supported by PP (97%).

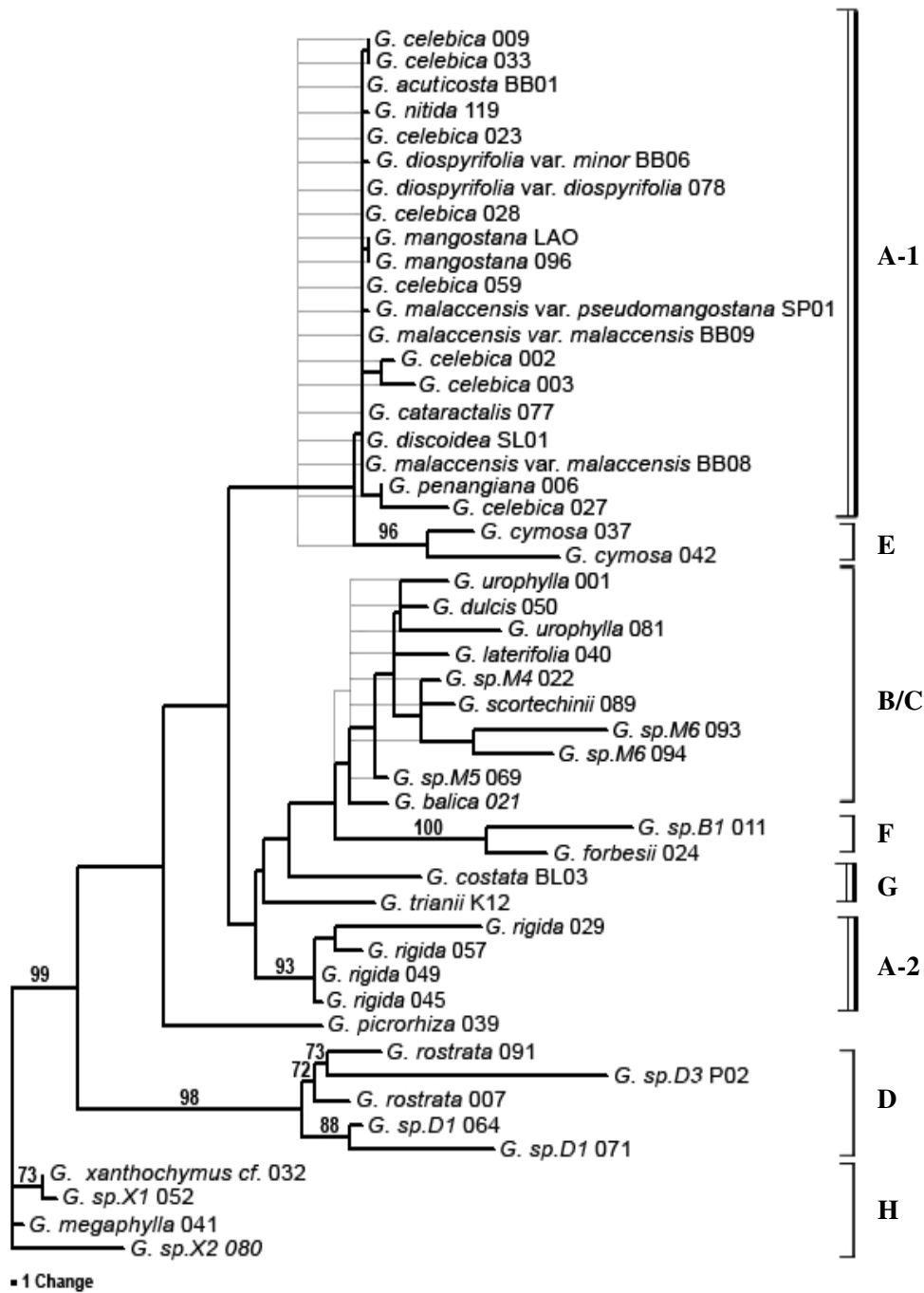


Figure 5.5. One of 10,000 most parsimonious trees from the analysis of the *trnS-G* region. Numbers above branches indicate bootstrap values (shown for values above 60% only). The grey branches indicate the topology of the strict consensus tree. Tree length=525 steps. CI=0.81, RI=0.76 and HI=0.19. The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980).

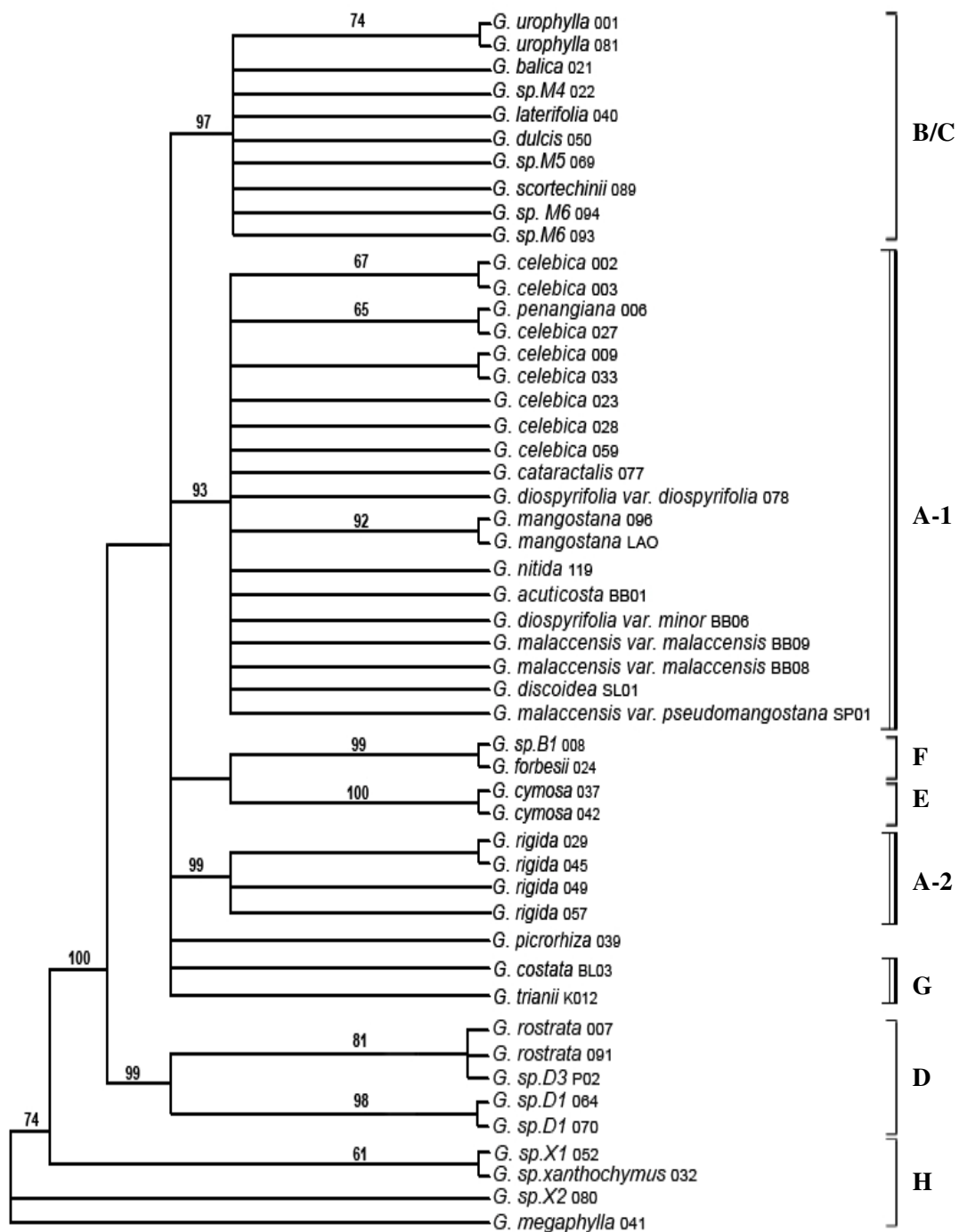


Figure 5.6. Fifty percent Majority Rule consensus tree resulting from Bayesian analysis of the *trnS-G* region. Numbers above branches indicate posterior probabilities (shown for values above 60% only). The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980).

#### 5.4.3 Combined *trnS-G* and *trnD-T*

The combination of the two cpDNA data matrices resulted in a total of 2499 characters of which a low number (149; 5.96%) were phylogenetically informative. From the 24 accessions of *trnS-G* and *trnD-T* regions, a total of 58 gap characters were used in the analysis. In the MP analysis, more than 10,000 parsimonious trees were produced in both the first and second round of heuristic searches with a tree length of 491 steps. The CI (0.88) and RI (0.80) were higher than for the *trnS-G* region but the HI (0.12) was lower compared to *trnS-G*.

The MP and BI analyses show a similar topology, and some of the clades are more resolved in this combined analysis compared to *trnS-G* trees. For example in Clade **A-1**, *G. celebica* is monophyletic with moderate BV (70%) and PP (74%) support. Another monophyletic clade in clade **A-1**, the group of *G. malaccensis* and *G. mangostana* differs from the ITS trees in the placement of *G. malaccensis* var. *pseudomangostana* which is nested outside the group.

Clade **A-2** is sister to clade **A-1** with low support (60%) in the parsimony analysis. However, in the BI analysis *G. rigida* (accession 045) is sister to clade **A1** but with PP less than 60%. This difference with the *trnS-G* analysis may be because of the limited sample of taxa used in this analysis. The paraphyly of sect. *Garcinia* sensu Jones (1980) indicated by ITS and *trnS-G* trees is still evident from the position of clade **E** and **G** in the parsimony and BI analysis. Sequencing of plastid regions from more taxa, particularly

from the clades **B**, **C**, and **F** will be necessary to provide further evidence for paraphyly of section *Garcinia*.

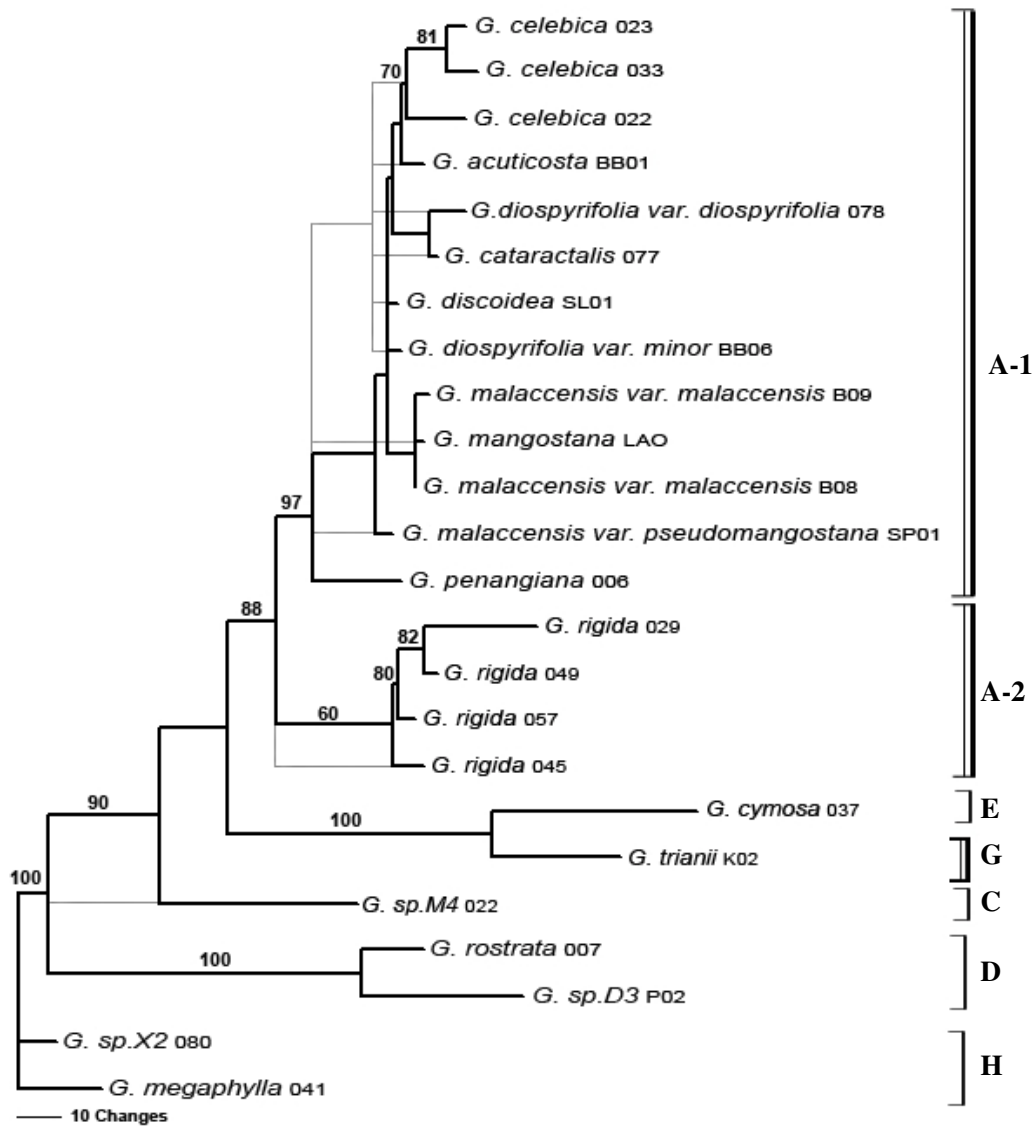


Figure 5.7. One of 10,000 most parsimonious trees from the analysis of the *trnS-G* and *trnD-T* regions combined. Numbers above branches indicate bootstrap values (shown for values above 60% only). The grey branches indicate the strict consensus tree. Tree length=514 steps. CI=0.87, RI=0.79 and HI=0.14. The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980).

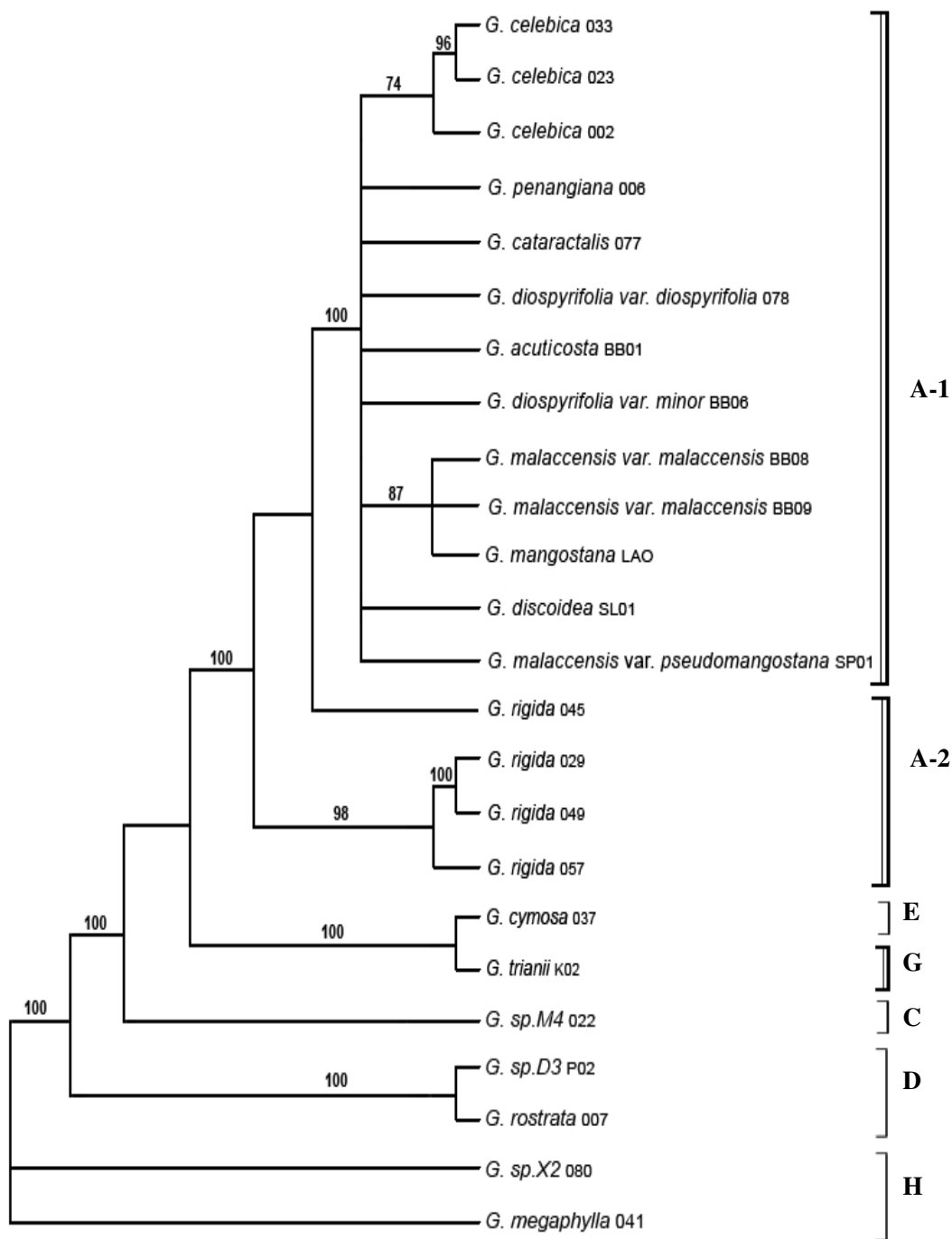


Figure 5.8. Fifty percent Majority Rule consensus tree resulting from Bayesian analysis of combined *trnS-G* and *trnD-T* sequences. Numbers above branches indicate posterior probabilities (shown for values above 60% only). The double lined bars indicate taxa that were placed in section *Garcinia* sensu Jones (1980).

#### 5.4.4 Congruence Test

Two tests were performed to test whether the ITS dataset should be combined with cpDNA dataset. Both ILD tests between ITS and *trnS-G* datasets, and ITS and combined *trnS-G* with *trnD-T* produced results indicating that these data partition had a significantly different signal (both with  $P= 0.001$ ).

To further investigate the congruence of the two datasets, the tree topologies for each dataset can be compared to assess the degree and sources of incongruence. This method has been practised for some authors, (e.g. Soltis & Kuzoff, 1995; Ronsted et al., 2002) who abandoned the incongruence length difference (ILD) test (Farris et al., 1994) because it indicated significant incongruence even when none was apparent. In sect. *Garcinia* plastid trees show incongruence with the ITS trees especially in the placement of clade **A-2** (*G. rigida*). In the ITS analysis it is placed within clade **A** and as sister to *G. diospyrifolia*, but in the *trnS-G* analysis it is placed outside clade **A** (**A-1**), and as its sister group in the combined plastid analysis.

There appears to be significant incongruence evident both in the ILD test and the comparison of tree topology, and therefore, combined analysis is rejected in this study. This is justified because if separate datasets are incongruent as a result of evolutionary independence, combined analysis may result in reduced resolution or an erroneous estimate of phylogeny (Hardig et al., 2000).

#### 5.4.5 Age Estimation

Three nodes within ITS trees have been calculated using the NPRS method (Sanderson, 1997) to estimate their ages and their standard deviation. Results show that the age for the crown of *Garcinia* is  $61.26 \pm 1.58$  MY, the most recent common ancestor of the group of *G. malaccensis* and *G. mangostana* is  $8.66 \pm 4.43$  MY and the split of accessions of *G. rigida* from their most recent common ancestors is  $21.58 \pm 2.90$  MY.

Another method of age estimation as used by Richardson et al.(2001a), estimated that ages for the crown node of *Garcinia* from published rates of woody taxa resulted in a huge range from 25.9 million years (MY) to 631 MY (Table 5.4). The Winteraceae rate gave ages of 575 to 631 MY for the crown of *Garcinia*, however, other rates gave more reasonable ages ranging from 25.9 to 99.7 MY. The Winteraceae age estimates are clearly incorrect (*Garcinia* is estimated to be considerably older than the oldest angiosperm fossils). This either indicates that the Winteraceae rate is incorrect, possibly resulting from errors in the calibration, or it suggests that the problem of rate heterogeneity among lineages is too great to permit use of a rate calculated in one lineage to accurately determine the age of another. Therefore, the ages calculated using the fossil calibration are the most accurate and are those that are discussed below.

Table 5.4. Estimated time of divergence of *Garcinia* based on the published ITS data.

Species (Source)	Habit/Minimum Generation Time (estimated)	Published Calibrated Rate (s/s/y)	Age of crown node of <i>Garcinia</i> (MY)
<i>Gossypium</i> (Mes et al., 1996)	Perennial shrubs/ ~1-3 yrs.	5.0 – 9.0 x 10 <sup>-9</sup> (ITS 1)	55.0 – 99.7
<i>Gossypium</i> (Mes et al., 1996)	Perennial shrubs/ ~1-3 yrs.	2.5 – 4.5 x 10 <sup>-9</sup> (ITS 2)	27.9 – 49.8
<i>Inga</i> (Richardson et al., 2001a)	Tree & Shrubs/~2-3 yrs.	2.34 x 10 <sup>-9</sup>	25.9
<i>Phyllica</i> (Richardson et al., 2001b)	Tree & Shrubs/~2-3 yrs.	2.44 x 10 <sup>-9</sup>	27.0
Winteraceae (Suh et al., 1993)	Tree & Shrubs/~6-10 yrs.	3.2 – 5.2 x 10 <sup>-10</sup> (ITS 1)	575.0
Winteraceae (Suh et al., 1993)	Tree & Shrubs/~2-3 yrs.	3.6 – 5.7 x 10 <sup>-10</sup> (ITS 2)	631.0

## 5.5 Discussion

### 5.5.1. Utility of ITS and cpDNA Sequences

A total of 10 species of *Garcinia* out of 17 species belonging to sect. *Garcinia* sensu Jones (1980) were collected and successfully sequenced for the ITS region. Nine species of sect. *Garcinia* were sequenced for the *trnS-G* region and six for *trnD-T*. Among these regions, ITS provides most phylogenetic resolution because of the higher level of variation compared to cpDNA. The phylogenetic analyses also produced ITS trees in which infrageneric and species relationships could be resolved. This result is in accordance with previous findings based upon studies using fewer samples (Nazre, 2000; Rismita-Sari, 2000; Yapwattanaphun et al., 2004) that showed that ITS provides greater phylogenetic information in *Garcinia* than other markers.

Nazre (2000) demonstrated that chloroplast markers of *trnL-trnF* are too conserved to resolve inter-specific relationships. To improve this resolution, two markers of cpDNA reported to be highly variable were sequenced in this study. However, the two chloroplast markers (*trnS-G* and *trnD-T*) combined still did not produce better resolution than ITS. This is because the chloroplast evolves at a slower rate compared to nuclear DNA (Palmer, 1987) and in *Garcinia* it appears that this rate difference may be accentuated. Although variation in chloroplast markers is insufficient to resolve inter-species relationships it does provide some good information on infrageneric relationships. It also indicates that some species might be of hybrid origin as shown by the contrasting topologies observed between ITS and cpDNA datasets (see discussion in section 5.5.4). Because of the better resolution and sampling of ITS, most of the discussion on the relationships within sect. *Garcinia* that follows will focus on the ITS trees.

#### 5.5.2. Section *Garcinia* and Morphology

The trees from the phylogenetic analyses indicated that sect. *Garcinia* sensu Jones is paraphyletic. In the ITS analysis, the members of the section are found in two clades, **A** and **G** (Figure 5.3 & 5.4) but in the *trnS-G* analysis, species from sect. *Garcinia* sensu Jones were found in three clades, clade **A-1**, **A-2**, and **G** (Figure 5.5 and 5.6). Sect. *Garcinia* therefore should constitute only clade **A** as shown in the ITS analysis or clades **A-1** and **A-2** in the *trnS-G* analysis, where the support values are significant and also supported with morphological evidence (Chapter 4). The incongruence in the position of

clade **A-2** in the *trnS-G* analysis may be due to a hybridisation event (see 5.5.4 below). However, in the combined plastid analysis, an increased taxon sampling is necessary to confirm whether sect. *Garcinia* is paraphyletic as indicated by the nuclear dataset.

Clade **A** contains most species that had been previously recognised as members of sect. *Garcinia* and the high support values for this clade are similar to those in previous studies (Nazre, 2000; Rismita, 2000; Yapwattanaphun et al., 2004). Another clade containing species from Jones's section *Garcinia*, clade **G**, consists of three species *G. maingayi*, *G. trianaii* and *G. costata*. This clade is supported by significant morphological evidence and it should therefore be treated as a different section. The most distinct character is the stout and ribbed leaves that are not found in other members of sect. *Garcinia* (see Chapter 6). Following Whitmore's (1973) artificial grouping, Jones (1980) placed these species into sect. *Garcinia* because of male floral characters; stamens in the centre, surrounding the fungiform pistillode and the 2-theous anthers. These morphological characters are homoplasious, i.e. both clades **A** and **G** share these characters. However, upon close investigation the stamens for species in clade **A** differ from those in clade **G** by the shape of the bundles with four lobes fused together whereas clade **G** does not have these four lobes (Figure 4.8).

Evidently, the morphological characters that support the monophyly of sect. *Garcinia* are a combination of both reproductive and vegetative features. The most important of these are the male flowers, especially the stamens and as Whitmore (1973) has demonstrated, leaf characters could also be used for classification. Jones (1980) also

used these characters for her sectional classification. In the absence of the male flower as shown in *G. discoidea* and *G. acuticosta*, the fruit characters, i.e. borne terminal, solitary and with distinct protruding stigma, together with leaf characters could be used.

### 5.5.3. *Phylogenetic Relationships within Sect. Garcinia*

Species in sect *Garcinia* can be grouped into four groups, as shown in figure 5.8 below, that also have significant morphological support. These groups are with one exception (Group 3), monophyletic.

**Group 1:** This group is moderately (64%) supported by BV but highly supported by PP (98%) and consists of three species, *G. celebica*, *G. acuticosta* and *G. discoidea*. No male flowers were found (from herbarium specimens and living trees) for *G. acuticosta*.

However, a combination of leaf characters, e.g. pointed cross section of midrib, narrowly elliptic of leaf shape, inconspicuous secondary nervation could be used to differentiate these three species from other species in sect. *Garcinia* which supports the suggestion of Whitmore (1973) that there is variation in leaf morphology that can differentiate groups of *Garcinia* species. The fruit characters that are shared by this group include: borne terminal, solitary with a thick fruit wall, ovoid or globose, with a distinct and prominent stigma that is either weakly to strongly raised and is either concave or discoid.

*G. celebica* is a heterogeneous species, both in terms of molecular and morphological features, and is moderately supported by bootstrap values (79%) and highly supported by

posterior probability (100%). Previously, *G. celebica* was divided into many species with many synonyms. The distribution of this section encompasses eastern India to Sulawesi partly because regional floras by different authors treated them as distinct from each other (see chapter 6 for explanation). The heterogeneous nature of the morphological characters is seen clearly in the leaves and fruits of *G. celebica*. The leaves vary in shape and size, from elliptic or wide elliptic to lanceolate, sub-orbiculate or round and the fruit shape also varies but characters that unite the species are the four lobed bundles of stamens surrounding the fungiform pistillode. This variation, together with its widespread distribution, might explain why within *G. celebica* there is considerable molecular variation. Richards (1990a) showed that two *G. celebica* populations in Peninsular Malaysia are facultatively agamospermic and this breeding system might explain morphological variation in this species (see Chapter 2).

The other species in this group are the newly described *G. discoidea* and *G. acuticosta*. The male flowers of *G. discoidea* are similar to *G. celebica* but are larger, with thicker sepals and petals and stout pedicels. Despite the absence of male flowers in *G. acuticosta*, it shares fruit characters that are very similar to *G. celebica* and *G. discoidea*, i.e. borne on a terminal branch, solitary, with a thick wall, but may be distinguished by the rather discoid stigma and the margin being not so clearly lobed as in *G. celebica*. Vegetative characters also differentiate *G. acuticosta* and *G. discoidea* from *G. celebica* as they have leaves that are elliptic, ovate, oblong or round with the main veins raised on both sides and with fine and distinct secondary nervation.

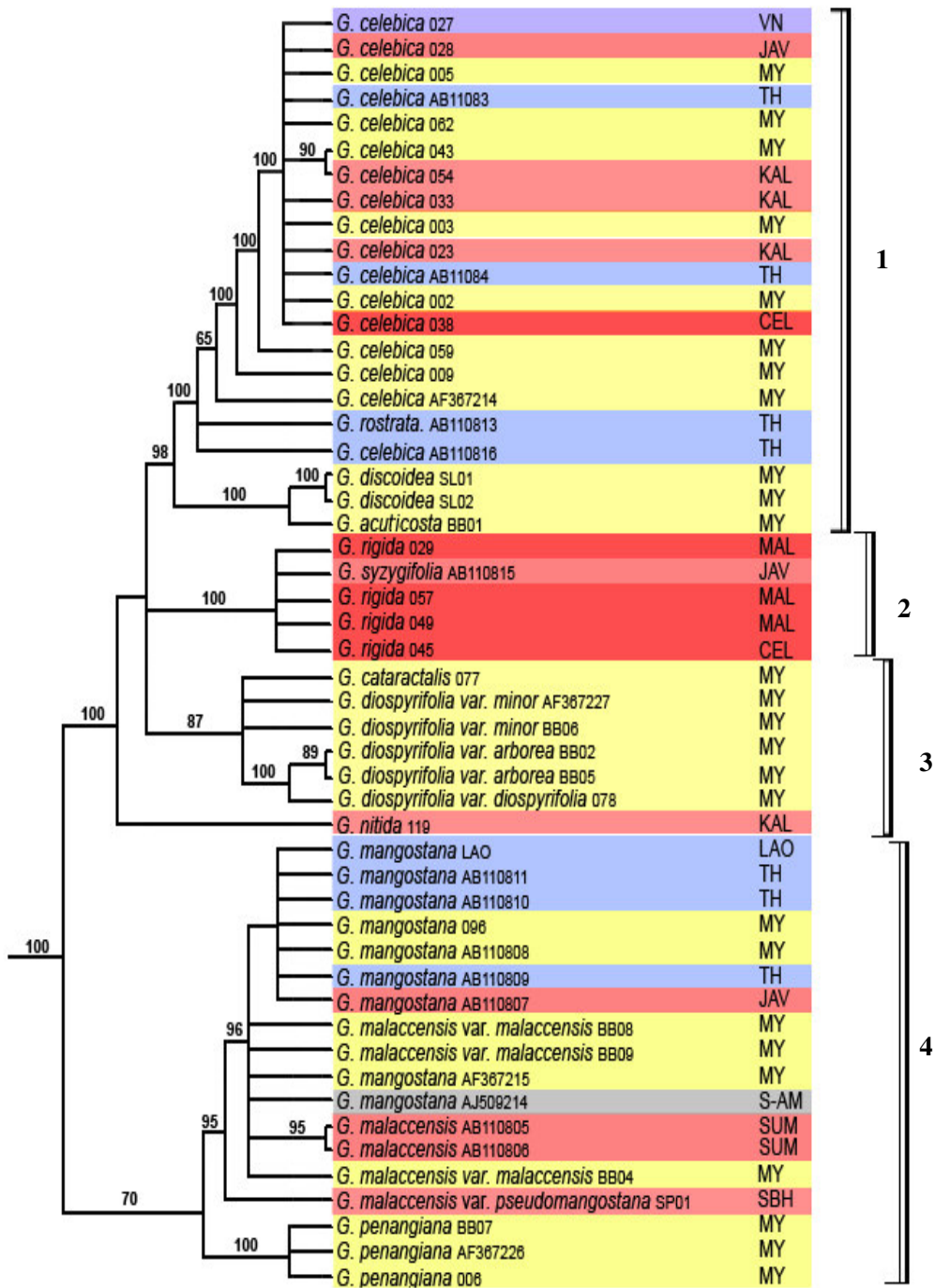


Figure 5.9. Groups within section *Garcinia* inferred from Bayesian analysis of ITS. Origin of samples collected; CEL=Sulawesi, JAV=Java, KAL=Kalimantan, LAO=Laos, MAL=Maluku, MY=Peninsular Malaysia, SBH=Sabah, SUM=Sumatra, TH=Thailand and VN=Vietnam.

**Group 2:** Group 2 is a strongly supported clade consisting of the poorly known species *G. rigida* (*G. sizygiifolia* as explained above is probably a misidentification). The ITS analysis placed *G. rigida* within clade **A** in sect. *Garcinia* but in the *trnS-G* analysis, it is placed outside of the clade, as the sister of clade **G** (*G. costata* and *G. trianii*). This contrasting topology could be evidence of hybridisation in the ancestor of *G. rigida*. Speciation from hybridisation processes in *Garcinia* is probable, for instance in *G. mangostana*, a species of suspected hybrid origin (Richards, 1990c). Determining the parents of *G. rigida* would need more detailed molecular studies utilising more variable plastid and nuclear markers and accessions of a wider sample of species. Since the samples for this study were collected from cultivated plants in the Bogor Botanic Garden, it is difficult to confirm the hybrid origin of this species.

Available information on *G. rigida* indicates that it is a rare species (four specimens) with a scattered distribution in Indo-china and Sumatra. In BO, the cultivated trees were indicated to have originated from Maluku and Sulawesi. Morphologically the leaves resemble *G. celebica* but differ largely with the glandular line pattern being more distinct and the lines more widely spaced. The fruit is similar to that of species in group 3, but is thin walled, rather rigid when dry and not easily ruptured. The fruit colour is paler and the stigma a flattened-discoïd shape compared to the bright red or maroon colour and concave or convex discoïd and lobed stigmas found in species in group 3.

**Group 3:** *G. diospyrifolia*, *G. cataractalis* and *G. nitida* share the same thin fruit wall; the fruit is easily ruptured when dry, and the mature fruit usually turns reddish and has a

smooth surface. These species are indicated as related, but not monophyletic by the ITS analyses. While *G. diospyrifolia* is distributed in Sumatra, Peninsular Malaysia and Borneo, *G. cataractalis* is restricted to Peninsular Malaysia and *G. nitida* to Borneo. The latter differs from the other two species by the size of the tree which can reach 30 m, while *G. diospyrifolia* and *G. cataractalis* are small trees to 15 m.

Within *G. diospyrifolia*, three varieties are observed that have morphological support; *G. diospyrifolia* var. *arborea* (62% BV and 89% PP) is sister to *G. diospyrifolia* var. *diospyrifolia* (78% BV and 100%PP). The relationship of *G. diospyrifolia* var. *minor* is unresolved with respect to other varieties of this species and *G. cataractalis*. Both *G. diospyrifolia* var. *diospyrifolia* and *G. diospyrifolia* var. *arborea* are widely distributed varieties found in Sumatra, Peninsular Malaysia and Borneo but *G. diospyrifolia* var. *minor* is only found in Peninsular Malaysia. Morphologically, these three varieties differ by the stature of the tree. *G. diospyrifolia* var. *diospyrifolia* is a slender and small treelet to 5 m, *G. diospyrifolia* var. *minor* is a bushy tree to 12m and *G. diospyrifolia* var. *arborea* is a medium-sized tree to 20 m.

Unlike other species in the group, *G. cataractalis* is a rheophyte and can be distinguished from *Garcinia diospyrifolia* and *G. nitida* by its small, very narrow elliptic leaves. *Garcinia nitida* on the other hand is a big tree, with leaves that nearly resemble *G. diospyrifolia*, but is characterised by the flask-shaped fruit where the stigma is distinctly elongated to 10 mm.

**Group 4:** This group contains the widely cultivated *G. mangostana* together with *G. malaccensis* and *G. penangiana*. The monophyly of this group is moderately supported by the MP bootstrap (81%) and low PP values of BI analysis (70%). The common characters shared between the species are the stamens being in a central bundle that is conical, four-angled or nearly spherical. *Garcinia mangostana* is only known in cultivation and many authors believed that it is the product of hybridisation of closely related species (Whitmore, 1973; Richards, 1990c). See section 5.5.4. for a discussion on the origin of mangosteen.

A clade with 87% BV and 95% PP consists of representatives of *G. malaccensis* and *G. mangostana* although neither of these species are themselves monophyletic (Figure 5.2, 5.3 and 5.9). Within this clade *G. malaccensis* var. *pseudomangostana* is sister to the clade containing the other representatives of these species. Paraphyly of *G. mangostana* might suggest multiple origins of cultivation.

In general, there are three groups of *G. malaccensis* from the molecular data; the Peninsular Malaysian group (*G. malaccensis* var. *malaccensis*), the Sumatran group which is moderately supported as a monophyletic clade, and the Sabah group (*G. malaccensis* var. *pseudomangostana*). It is difficult to ascertain the variety of *G. malaccensis* from Sumatra since no herbarium specimens of wild origin have been seen but contacts in Bogor Botanic Gardens (Rismita-Sari & Irawaty, pers. com) indicate the origin of trees in cultivation there is Sumatra.

The group that is sister to *G. malaccensis* var. *pseudomangostana* is moderately supported by BV (62%) and PP (96%). Morphologically, the two varieties of *G. malaccensis* are extremely similar vegetatively but the male flowers of *G. malaccensis* var. *malaccensis* have long 4-angled conical-shaped stamens, whereas *G. malaccensis* var. *pseudomangostana* has shorter four-angled square-shaped stamens (Figure 4.7). Both varieties have the pistillode present or absent but the size is small (less than 5 mm long). *Garcinia mangostana*, however, has a long four-angled stamen surrounding the prominent fungiform pistillode, while the leaves resemble *G. malaccensis* but with the venation more prominent and stout.

*Garcinia penangiana* is the sister to the rest of the group 4 clade and has stamen bundles which resemble *G. malaccensis* var. *pseudomangostana* but it differs chiefly by the absence of the pistillode, the sepals and petals being rather thinner and the veins on the leaves are much closer to each other.

#### 5.5.4. Hybridisation and Origin of Mangosteen

The phylogenetic trees indicated that nuclear and plastid data were incongruent (Figure 5.3 – 5.8). A possible explanation for this discordance is hybridisation and/or introgression amongst species found in different lineages, as has been shown in other studies. For example Kyndt et al. (2005) demonstrated that discrepancies of the

chloroplast and nuclear markers in the Caricaceae were evidence of hybrid species of *Vasconcellea*.

Chloroplast DNA is maternally inherited in angiosperms and is a non-recombining molecule, and as a result hybrid species should retain the same sequences as their maternal parents. Nuclear markers in contrast are biparentally inherited, but some, such as ITS, may be subjected to concerted evolution. Koch et al. (2003) outlined three possibilities for ITS in individuals that result from hybridisation between two different species with different ITS sequences: 1) unidirectional concerted evolution leads to the loss of one copy and fixation of the second; 2) concerted evolution results in new ITS sequences that represent a mixture of the two parental sequences; 3) both parental ITS sequences are retained in a 'non-concerted evolution' situation. Concerted evolution is a molecular process that homogenises different loci within multigene families (Arnheim et al., 1980). It is driven by two molecular processes, gene conversion and unequal crossing over (Koch et al., 2003) and is only effective with sexual reproduction.

Evidence for hybridisation in this study of *Garcinia* can be seen from the incongruence in the position of *G. rigida*. The ITS trees (Figure 5.3 and 5.4) show that *G. rigida* is nested within clade A (sect. *Garcinia*) and is closely related to *G. diospyrifolia*, but in the *trnS-G* chloroplast trees (Figure 5.5 and 5.6), it is nested outside clade A while in the combined parsimony analysis (Figure 5.7 and 5.8), *G. rigida* is sister to clade A. Pinpointing the exact maternal parent of this species based on the plastid phylogeny presented here is difficult as the number of species sampled is insufficient. However, the

maternal parent may be more closely related to a species outside of sect. *Garcinia*. The absence of any polymorphism in the electropherograms of ITS sequences of *G. rigida* (Appendix VIII) is consistent with a hybrid origin followed by concerted evolution and homogenisation to fixation to an ITS type of paternal parent related to *G. diospyrifolia*. This phenomenon is known as chloroplast capture, an introgression of the cytoplasmic genome of one species into the nuclear background of another (Soltis et al., 1991). *Garcinia rigida* is a rare species that is widely distributed from Indo-china to Sumatra and Maluku. The hybrid origin of this species is intriguing and merits further investigation. Because information on this species is limited, further investigation to determine its parentage should include more species and additional markers as the current study can not give a clear picture of its relationships.

Another hybrid species and the most famous member of sect. *Garcinia*, is the mangosteen (*G. mangostana*), and its relationship with wild relatives has been the subject of great interest. A number of attempts have been made to determine its relationships. Richards (1990c) was the first to suggest from cytological and morphological evidence that mangosteen arose from hybridisation of male *G. hombroniana* (= *G. celebica*) and female *G. malaccensis*. This claim follows on from the work of Whitmore (1973), who also suggested the close affinities of mangosteen with *G. malaccensis* and *G. hombroniana*. Richards (1990c) speculated that mangosteen might be an offspring of a single event, but he also suggested that there was a possibility that the cross might have occurred on more than one occasion.

Using ITS sequence data from a total of 17 species of *Garcinia*, Yapwattanaphun et al. (2004) found that mangosteen is closely related to *G. malaccensis* rather than to *G. hombroniana* (= *G. celebica*). They also concluded that the other ancestral parent may not be *G. hombroniana*, but could also be *G. rostrata*, *G. sicygiifolia* or *G. speciosa* (Yapwattanaphun et al., 2004). This, however, may be misleading as indicated earlier in this chapter due to the dubious identification of a few species in this work.

Morphologically, Richards (1990c) suggested that the affinities of mangosteen were with *G. hombroniana* (= *G. celebica*) and *G. malaccensis* based on female flower and fruit characters. However, as demonstrated above, the most significant characters used to infer relationships in sect. *Garcinia* are from the male flower. Male flower characters in this study (Chapter 6) show that mangosteen is more similar to the group of *G. malaccensis*, *G. venulosa* and *G. penangiana* than it is to *G. celebica*. The male flower for this group including mangosteen has mainly 4-angled stamen bundles with fungiform pistillode (or without pistillode) (Figure 4.9) whereas *G. celebica* has 4-bundles of stamens surrounding the fungiform pistillode (Figure 4.8a & 4.9a). The ITS trees (Figure 5.3 and 5.4) also demonstrated that *G. malaccensis*, *G. penangiana* and mangosteen form a group (*G. venulosa* was not sequenced) that is monophyletic although not highly supported.

The male mangosteen stamens are 4-angled surrounding the base of a prominent fungi-form pistillode (Figure 4.9b). In contrast male flowers of *G. malaccensis* have a smaller or absent pistillode (Figure 4.9c & d). *Garcinia venulosa* (Figure 4.9e) and *G.*

*penangiana* (Figure 4.9f) do not have a pistillode and the stamen bundles are somewhat shorter. However, there is variation in male flowers in different varieties of *G. malaccensis*. *Garcinia malaccensis* var. *malaccensis*, found in Peninsular Malaysia and Borneo has nearly conical, 4-angled or nearly round stamens, and the pistillode is very small or sometimes absent. *Garcinia malaccensis* var. *pseudomangostana* has shorter, 4-angled stamens bundles, with or without a pistillode and is only found in the eastern part of Borneo (Sabah and Kalimantan).

If mangosteen arose from hybridisation between male *G. celebica* and female *G. malaccensis* as claimed by Richards (1990c), the ITS sequence should reflect either both parental sequences, one of the putative parents sequences or perhaps a new sequence that is a combination of both parental types, i.e. one of the three possibilities suggested by Koch (2003). There is support for the role of *G. malaccensis* in the origin of mangosteen from the ITS trees that place these species in the same monophyletic group.

Richards (1990a) stated that mangosteen is an obligate agamospermous species mainly based on the absence of male individuals. If mangosteen is indeed an obligate agamospermous species this would indicate that no sexual reproduction has occurred since the F1 generation. Thus, concerted evolution in the nrDNA would not have occurred and both parental ITS sequences should be retained throughout the succeeding obligate apomictic generations as for example in *Passiflora* (Lorenz-Lemke et al., 2005). If *G. celebica* is one of the parent species, sequences of *G. celebica* should be retained in the hybrid and there should be many polymorphisms detected in mangosteen sequences

generated from a single individual, i.e. sites at which e.g. both an A and a T are present each state representing that of one of each of the parents. These polymorphisms should be frequent given the sequence divergence between accessions of *G. celebica* and the *G. mangostana*/*G. malaccensis* clade (Figure 5.9). However, the electropherograms of ITS sequences of mangosteen accessions indicated that there are no polymorphisms observed (see appendix VIII) which would rule out the possibility of *G. celebica* being one of the parents for mangosteen provided mangosteen is an obligate agamosperm. If obligate agamospermy in mangosteen were confirmed then this would indicate that *G. malaccensis* is one of the parents and only another closely related species with an identical sequence to *G. malaccensis* could be the other parent for mangosteen or that mangosteen is a 'cross' between two varieties of *G. malaccensis* with identical ITS sequences.

Two mangosteen accessions (AF 367215, AJ509214) have sequences identical or nearly identical to *G. malaccensis* var. *malaccensis* (BB04, BB08, BB09) from Peninsular Malaysia and other accessions of mangosteen are nested within the *G. malaccensis* clade (Figure 5.3). This suggests that the other parent of mangosteen may be another variety of *G. malaccensis*. However, at present only two varieties of *G. malaccensis* (*G. malaccensis* var. *malaccensis* and *G. malaccensis* var. *pseudomangostana*) are described (Chapter 6). More varieties may await description, for example, individuals from Sumatra (Figure 5.3) which have ITS sequences that are slightly different from other varieties. If mangosteen is confirmed as an obligate agamosperm then sequencing of more accessions

of *G. malaccensis* and related species will be necessary to determine more accurately its parents.

However, mangosteen could also be a facultatively agamospermous species in which sexual reproduction occurs from time to time. Male individuals have been reported in Peninsular Malaysia (Idris & Rukayah, 1987). The rare occurrence of males may be related to the cultural practice of people in Peninsular Malaysia (and Southeast Asia generally) who chop down any male fruit trees because they believe they are of no benefit to them. A male tree reported by Idris and Rukayah (1987) had the same fate when a research team from the National University of Malaysia (UKM) visited the village where it had been seen and found that the tree had been chopped down because the villager felt it was not worth keeping trees which did not produce any fruit. Burkill (1935) also recorded a report from Chavelier (1919; cited in Burkill 1935) that villagers in Indo-china had practised the same action towards mangosteen trees that bore male flowers. The occurrence of male trees of mangosteen might not be as rare as first thought and that increases the possibility that sexual reproduction might have occurred in the past.

If mangosteen is a facultative agamospermous species, i.e. there is some sexual reproduction, then the possibility that *G. celebica* or some other *Garcinia* species is a parent cannot be ruled out. Based on the molecular data in this study, concerted evolution in ITS sequences of mangosteen might have followed possibility one of Koch et al. (2002) resulting in only one of the parental sequences being retained. In other words, concerted evolution has homogenised the sequence in favour of one of the parents. Even

though there is no information on how much sexual reproduction is needed for ITS sequences to homogenise in mangosteen, other findings such as in *Armeria* (Fuertes Aguilar et al., 1999) and *Tragopogon* (Kovarik et al., 2005) indicate that concerted evolution and homogenisation to one parental sequence could occur at a very rapid rate (within two and thirty generations respectively). Results from age estimation (5.4.5) suggest that the most recent common ancestor of *G. malaccensis* and *G. mangostana* can be dated to 8.66 MYA ( $\pm$  4.43 MYA) suggesting there has been ample time for mangosteen to reproduce sexually that could result in concerted evolution and homogenisation of ITS copies in favour of one of the parents.

Yapwattanaphun et al. (2003) used Amplified Fragment Length Polymorphism (AFLP) to determine that from 20 individual mangosteens investigated, variation existed although the level of polymorphism was relatively low. However, they suggested that this variation resulted from somatic mutation and concluded that all mangosteen trees were a clone originating from a single hybrid ancestor. Ramage et al. (2004) who investigated 37 accessions of mangosteen using a Randomly Amplified DNA Fingerprinting (RAF) technique, disagreed with the single origin theory and suggested two alternatives. The first possibility was that mangosteen might have arisen from independent hybridisation events and multiple hybrids were developed in cultivation. The second view postulated that backcrossing events occurred between male and a closely related female species. Ramage et al. (2004) found that there is considerable genetic variation in mangosteen but some phenotypic differences (e.g. crown and leaf shape), might result from environmental differences or cultural practices such as pruning techniques and shading of the trees,

rather than from genetic factors. Variation in ITS sequences amongst accessions of mangosteen in this study suggests multiple origins (Figure 5.3 and 5.4). This supports the AFLP data of Yapwattanaphun et al. (2003), and the RAF data of Ramage et al. (2004).

Resolution and support values within plastid trees (Figure 5.5- 5.8) are poor. However, these trees, particularly those from the combined analysis (Figure 5.7 and 5.8), are consistent with Richards, (1990c) claim that *G. malaccensis* is the maternal parent for mangosteen. Interestingly, mangosteen individuals have unique *trnS-G* chloroplast sequences (Figure 5.5 and 5.6). Combined plastid trees (Figure 5.7 and 5.8) indicate that the most closely related taxon is *G. malaccensis* var. *malaccensis* from Peninsular Malaysia. Within *G. malaccensis*, there is strong evidence of genetic variation between varieties as shown by *G. malaccensis* var. *pseudomangostana* from Sabah that is distinct from individuals from Peninsular Malaysia as shown in the combined analysis of cpDNA (Figure 5.7 and 5.8). More accessions could be sequenced to determine whether any individuals of *G. malaccensis* had a sequence that matched that of mangosteen.

At present the parental species of mangosteen cannot be determined with any confidence, although it seems likely that *G. malaccensis* is one (probably maternal). If the reproductive strategy of mangosteen can be confirmed as obligate agamospermy, then the possibility that *G. celebica* is one of the parents can be ruled out. In order to determine this, more work, especially investigation into the breeding system, needs to be done. Given the current data, a number of hypotheses for the origin of the mangosteen still remain and these could be further tested by adding more accessions of mangosteen and

putative parents and also additional molecular data. Additional and more variable plastid regions could confirm the maternal parent, as the markers used here have low levels of variability. Highly variable low copy number nuclear genes could also be sequenced. These markers are less frequently subjected to concerted evolution (Small et al., 2004) and would therefore retain each paternal sequence. These markers have been used to determine the parents of hybrids in *Paeonia* (Sang & Zhang, 1999), *Glycine* (Doyle et al., 1999), *Clarkia* (Ford & Gottlieb, 1999), rice (Ge et al., 1999) and *Elymus* (Mason-Gamer, 2001). This approach could also be used to confirm the putative hybrid parents of *G. rigida*.

#### 5.5.5. *Distribution and Biogeography of Sect. Garcinia*

The distribution of sect. *Garcinia* is centred in Southeast Asia and spreads westwards to eastern India, and to the east across Wallace's line to New Guinea (Figure 3.1). Because the sampling of species in this study is limited with most of the species sampled being from the Southeast Asian region, it is not possible to determine the area of origin of the section. However, the fact that most of the species in sect. *Garcinia* are distributed mainly on the west side of Wallace's line suggests an origin here. The most widely distributed species *G. celebica* is found on both sides of Wallace's line. Other species of *Garcinia*, for example species from clade **C** (sect. *Macrostigma*) and clade **D** (sect. *Discostigma*) distributed to the east of Wallace's Line are also placed apically in the phylogenetic trees (Figure 5.2 and 5.3). The age estimation for the crown of *Garcinia* of

61.26 +/-1.58 MY and an age of 21.58 +/-2.90 for the split of accessions of *G. rigida* from their most recent common ancestor on the west side of Wallace's Line is consistent with a Miocene dispersal event from west to east after the Sahul and Sunda shelves converged (c. 20 MY), and this fits the theory of recent dispersal of species across this biogeographic division (Morley, 1998).

## 5.6 Conclusion

The ITS analyses in this study provide the best resolution of species relationships in sect. *Garcinia* and non-coding chloroplast sequence data are less informative. Most of the sections delimited by Jones (1980) prove to be monophyletic but sect. *Garcinia* is paraphyletic. Section *Garcinia* should only constitute clade **A** (or **A-1** & **A-2** from cpDNA) because of the molecular support values and also morphological characters that distinguish it from clade **G** (*G. maingayi*, *G. trianaii* and *G. costata*) that should be separated to become part of sect. *Brindonia* or should probably be treated as a new section. Stamen characters outlined by Whitmore (1973) and Jones (1980) are the most significant ones supporting monophyly of sect. *Garcinia*.

The occurrence of hybridisation events in sect. *Garcinia* in the wild is demonstrated by the incongruence between ITS and cpDNA trees but further studies should be made to determine the parental species, for example of *G. rigida*. Another

hybrid species, the cultivated mangosteen could be the product of hybridisation between varieties of *G. malaccensis* provided that it is confirmed as an obligate agamosperm. However, if mangosteen is a facultative agamosperm other *Garcinia* species besides *G. malaccensis* could be involved in its parentage.

## CHAPTER 6

### TAXONOMIC REVISION OF *GARCINIA* SECTION *GARCINIA*

#### 6.1 Introduction

The following taxonomic revision is based on the studies of morphological variation and phylogenetic analysis of DNA sequences that have been discussed in previous chapters. Species concepts and the nature of species delimitation used in this study will be briefly discussed.

#### 6.2 Agamospermy and Species Delimitation

The role of breeding systems in affecting phenotypic form cannot be ignored especially in *Garcinia* which is known to be agamospermous. Even though only three species of sect. *Garcinia* i.e., *G. celebica*, *G. mangostana* and *G. penangiana* have been confirmed as agamospermous, based on embryological studies (Richards, 1990a; Chapter 3), there may be other agamospermous species in the section as indicated by the low male ratio observed from herbarium specimens.

The problem with facultatively agamospermous species is that they produce morphologically similar lineages of asexual progeny, along with occasional sexual progeny. These sexual progeny result in more lineages that are slightly morphologically different from the original ones (Richards, 2003). This variation is not restricted to morphological characters but extends to cytological and molecular differences (Stace, 1998).

*Garcinia* is a problematic group of plants for the taxonomist (Ashton, 1988c). This is demonstrated by the large numbers of unidentified specimens in most herbaria visited during this study. Even for a well-documented geographical area such as Malaysia, some species included in floristic accounts have been left unnamed (Whitmore, 1973; Ashton, 1988a; Kochummen, 1997) because the authors believed they might occur outside Malaysia and therefore have an existing binomial name. The problem is a lack of a recent monographic revision, the last of which covering *Garcinia* on a world-wide basis was completed 80 years ago (Engler, 1925). Since that time many new species have been described.

There have been attempts to give special ranks to apomictic taxa to reflect their special nature in that they are 'different from' or 'not equivalent' to sexual species (Stace, 1998). The term 'agamospecies' was originally introduced by Turesson (1929) and defines populations whose constituents could be regarded on morphological, cytological or other grounds as having had a common origin (Babcock & Stebbins, 1938; Davis & Heywood, 1963). Love (1960) then proposed that the category agamospecies should be

employed at the subspecific level but the proposal was never really accepted and name changes were not formalised. Many other terms similar or equivalent to agamospecies have been used, for example formae apomictae, agamotype, apomictic-coenospecies, apomict, agamotype, amphimictic group (Davis & Heywood, 1963). Agamospermous taxa have not been widely accepted at the specific or subspecific rank, and the aggregate species concept (e.g. Weber, 1998) has been proposed as an alternative because a broad definition of agamospermous species that may contain many segregate agamospermous lineages has been maintained by many users (e.g. *Taraxacum officinale* agg.). The formal use of the term aggregate species is rejected by the ICBN but continues to be used informally (Stace, 1998). The microspecies concept has also been used to describe a uniform population or lineage that is morphologically distinguishable from other populations but the term has no real taxonomic definition rather than recognition of a certain type of variation (Asker & Jerling, 1992).

Although I believe it is necessary to consider the role of agamospermy in species delimitation in *Garcinia*, it is almost impossible in this under-studied group of plants. Not only is little known about the biology, cytology and mode of reproduction of *Garcinia* species but, in certain areas, some species are under represented in herbaria because of under-collection, e.g. *G. exiguus* (Borneo), *G. cataractalis* (Peninsular Malaysia) and *G. ochraceus* (New Guinea) or because they are very rare. Therefore, a pragmatic approach as proposed by Stace (1998) has to be used in sect. *Garcinia* without too much emphasis on the breeding system.

This is also the approach in taxonomic works on other genera known to have agamospermous species in the tropical forest such as *Shorea*, *Mangifera* and *Lansium* (Ashton, 1988a). These place little emphasis on breeding systems, but have proven to be stable classifications. These works use a relatively narrow species concept and recognise discontinuity of morphological variation at species or infraspecific rank (e.g. subspecies or variety). A similar approach to species delimitation is adopted in this study.

### **6.3 Species concept used in this study**

The justification for the use of a particular species concept in a taxonomic work is important because different taxonomists use different approaches to classifying the organisms they study. The biological species concept (Mayr, 1970) defined species as ‘groups of interbreeding natural populations which are reproductively isolated from all other groups’. This seems ideal for application to sexual organisms by stressing the ‘reproductive isolation’ mechanism where two different species will not interbreed. However, as outlined above, so little is known about breeding in *Garcinia* that this concept cannot be used. Furthermore, even if ample breeding data were available, the possibility of widespread agamospermy and the hybrid origin of species (Chapter 5) in sect. *Garcinia* could not easily accommodate this concept. In terms of practicality, particularly when using herbarium specimens, the biological species concept is impossible to implement.

Alternatively, the phenetic species concept (Cronquist 1978) defined as ‘phenetically distinct groups (species) that are separated from others by discontinuities’ uses overall similarities for comparison and is very difficult to apply in dioecious groups such as *Garcinia*, especially with a limited number of specimens. For example, the absence of male specimens in *G. acuticosta* and *G. cataractalis* means that only female characters can be used in species delimitation. Another difficulty is how to give weight to character states within an agamosperous species such as *G. celebica* when comparing them to unknown breeding type of other species. Therefore, the phenetic species analysis is rejected for this treatment.

Hennig (1966), who introduced the cladistic method, gave the name phylogenetic systematics to the use of empirical methods to form classifications. Cladistics uses derived characters (apomorphies) to define groups and requires taxa to be monophyletic. Two competing species concepts were born out of Hennig’s phylogenetic philosophy, the Monophyletic Species Concept (MSC), for example Baum (1992), and the Phylogenetic Species Concept (PSC), for example Nixon & Wheeler (1990). The MSC assumes that species are the smallest monophyletic unit that can be distinguished while the PSC states that monophyly is not demonstrable at species level and species are the smallest diagnosable units where constant characters are sought (Luckow, 1995). According to Wheeler & Platnick (2000), the phylogenetic species concept has its origin in the work of Rosen (1978, 1979) who defined a species as a geographically constrained group of individuals with some apomorphous characters.

In section *Garcinia*, the concept chosen is highly dependent on the availability of data and the practicality in use of the end product. This is because a concept which is firmly rooted in theory but which is difficult or impossible to apply to the study of real organisms is not much use (Cracraft, 2000). Using this logic, species delimitation within this work follows the essence of the Phylogenetic Species Concept based upon the definition by Wheeler and Platnick (2000) where a species is defined as the smallest aggregation of (sexual) populations or (asexual) lineages diagnosable by a unique combination of character states. Within this concept, important synapomorphic characters are identified to define the species limits and cladistic analysis of molecular data can be used to investigate further the nature of the species. In relation to agamospermy, because the PSC is applicable to both sexual and asexual organisms, this fits well.

Species with considerable intraspecific variation which might reflect agamospermy are treated as a single species with no intraspecific taxa if the intraspecific variation is continuous and with no obvious geographical patterns (e.g. *G. celebica*). However, if there are distinct variations and discontinuous patterns within the species, varietal rank is used because there is diagnosable variation within the species (e.g. *G. diospyrifolia* var. *diospyrifolia*). Although many proponents of the PSC, including Wheeler and Platnick (2001), do not recognise ranks below species level, they are, however, accepted, for instance, in the works of Nixon & Wheeler (1990) and Davis & Nixon (1992) where there are fixed differences detected among populations of the same species.

#### 6.4 Groups in Section *Garcinia*

Grouping the species facilitated their identification, especially when using fertile specimens. Even though some of the groups, e.g. Group I and II are monophyletic in the phylogenetic analysis, none will be given taxonomic rank as they are used here only for identification purposes. Moreover, further study is needed, especially phylogenetic analysis before declaring these groups natural and worthy of a taxonomic name.

**Group I** – *Stamen bundles conical or 4-angled; fruit wall thick or thin; calyx thickly leathery; stigmas separate or in bundles; secondary veins brochidodromous.*

The shape of the stamens is more or less conical and 4-angled; a pistillode is absent or, if present, fungiform. The fruit is like a mangosteen with a thick wall except for *G. penangiana* and *G. venulosa*, with a thin wall but which is not easily ruptured, thickly leathery, with a persistent calyx and tipped with segregated bundles of stigmas, fused in the middle.

Species: *G. mangostana*, *G. malaccensis*, *G. penangiana* and *G. venulosa*.

**Group II** – *Stamen bundles 4-lobed, open at maturity; fruit wall thick; calyx leathery; stigma lobed and sunken or convex; secondary veins brochidodromous.*

Stamens 4-lobed, united at the base and encircling the pistillode at the centre. Pistillode fungiform. Fruits with thick wall as in *Group I*, the stigma usually raised but sunken in the middle or convex with 4-5(7) lobes, the lobes sometimes indistinct.

Species: *G. acuticosta*, *G. celebica* and *G. discoidea*.

**Group III** – *Stamen bundles 4-lobed; fruit wall thick; calyx leathery; stigma skull-cap shaped; secondary veins cladodromous.*

Stamens as in *Group II*. Fruits small with thick wall, tipped with the sessile, rugose, skull-cap shaped stigma.

Species: *G. harmandii* and *G. moselleyana*.

**Group IV** – *Stamens bundles minutely 4-lobed, usually not open upon maturity; fruit wall thin and easily ruptured; calyx leathery; stigma sunken, skull-cap shaped or flattened; secondary veins brochidodromous.*

The lobing of the stamen bundles is not as distinct as in *Group II*, always surrounding the fungiform pistillode. Fruits with thin wall and the stigma raised or not, discoid or sunken, weakly lobed or not.

Species: *G. cataractalis*, *G. diospyrifolia*, *G. exiguus*, *G. nitida* and *G. ochraceus* .

**Group V** – *Stamens bundles minutely 4-lobed; fruit wall thick and very hard; calyx leathery; stigma sunken or flattened; secondary veins brochidodromous.*

The stamens as in *Group IV* but the fruits with thick wall and the stigma thinly flattened or usually sunken.

Species: *G. sangudsangud*.

**Group VI** – *Flowers unknown; fruit wall thin; calyx leathery; stigma discoid; secondary veins brochidodromous.*

No information on the flowers. The fruit wall closely allied to *Group IV* but not easily ruptured, and with a discoid stigma.

Species: *G. rigida*.

## 6.5 Generic Description

### *Garcinia* L.

*Garcinia* L. Sp. Pl. (1753) 443; Juss., Gen. Pl. (1789) 256; Blume, Bijdr. 5 (1825) 213; Miq., Fl. Ind. Bat. 1, 2 (1859) 506; Pierre, Fl. Forest. Cochinch. 1 (1882) 1; Backer & Bakh. f., Fl. Java 1 (1863) 90; Koord. & Valetton, Bijdr. Boomsoort. Java (1903) 356; King, J. As. Soc. Beng. 59 (1890) 148; Ridl., Fl. Mal. Penins. 1 (1922) 167; A.C. Sm. & S. Darwin, J. Arnold Arb. 55 (1974) 243; Seem., Fl. Vit. (1865) 10; A.C. Sm., Fl. Vit. Nova 2 (1981) 342. — *Mangostana* Rumph., Herb. Amboin. (Rumphius) 1 (1741) 132, Rumph. ex Gaertn., Fruct. Sem. Pl. 2(1), (1790) 105. — Type: *Garcinia mangostana* L.

*Rheedia* L., Sp. Pl. (1753) 1193; Planch. et Triana, Ann. Sci. Nat. (Bot.) ser. 4,14 (1860) 306; Hammel, Ann. Missouri Bot. Gard. 76 (1989) 927. — *Mammea* sensu Vahl, Eclog. Amer. 2 (1798) 40, pro parte quoad *M. humilis* Vahl. — *Lamprophyllum* Miers in Trans. Soc. London, 21 (1853) 249, pl. 26. — Type: *Rheedia laterifolia* L. [= *Garcinia humilis* (Vahl) C.D. Adams ].

*Cambogia* L., Gen. Pl., ed. 5 (1754) 225. — Type: *Garcinia gummi-gutta* (L.) N. Robson.

*Oxycarpus* Lour., Fl. Cochinch. (1790) 647. — Type: *Oxycarpus cochichinensis* Lour. [= *Garcinia cochichinensis* (Lour) Choisy ].

*Verticillaria* Ruiz et Pav., Fl. Peruv. Prodr. 81, (1794) pl.15 — Type: *Verticillaria acuminata* Ruiz et Pav. [= *Garcinia madruno* (Kunth.) Hammel].

*Xanthochymus* Roxb., Pl. Coromandel 2 (1798) 51, pl.196. — Type: *Xanthochymus pictorius* Roxb.[= *Garcinia xanthochymus* Hook. f.].

- Brindonia* Thouars in Dict. Sc. Nat. 5 (1804) 339. — Type: *Brindonia indica* Thouars. [= *Garcinia indica* (Thouars) Choisy ].
- Stalagmitis* Murray in Commentat. Soc. Regiae Sci. Gott. 9 (1789) 173. — Type: *Stalagmitis cambogioides* Murray [= *Garcinia ovalifolia* Oliv. ].
- Hebradendron* Graham in Hooker, Companion Bot. Mag. 2, (1836) 199, pl. 27. — Type: *Hebradendron cambogioides* Graham [= *Garcinia morella* Desr.].
- Discostigma* Hassk., Cat. Hort. Bot. Bogor. (Hassk.) (1844) 212. — Type: *Discostigma rostrata* Hassk. [= *Garcinia rostrata* (Hassk.) Hook. f.].
- Ternophyllum* Thwaites in Hooker's J. Bot. Kew. Gard. Misc., 6 (1854) 70. — Type: *Ternophyllum zeylanicum* Thwaites [= *Garcinia ternophylla* (Thwaites) Thwaites].
- Rhinostigma* Miq., Fl. Ned. Ind., Eerste. Bijv. (1860) 495. — Type: *Rhinostigma parvifolia* Miq. [= *Garcinia parvifolia* Miq.].
- Pentaphalangium* Warb. in Bot. Jahrb. 13 (1891) 382; Engl. in Engl. & Prantl, Naturl. Pflanzenfam. 3, 6 (1893) 240. — Type: *Pentaphalangium crassinerve* Warb. [= *Garcinia crassinervis* (Warb.) Kosterm.].

*Trees* to 30 m tall, occasionally shrubs. *Exudate* yellow, white or cream. *Twigs* angled or not, wrinkled lengthwise when dry. *Petiole* often finely striate horizontally, stout, broadly channelled on the adaxial surface and sometimes with ligule (an excavation at the base of the petioles clasping the twigs). *Lamina* leathery or papery, variable in shape but mainly elliptic; apex acute, acuminate or attenuate; base acute to cuneate or decurrent; midrib flattened to raised above, usually raised; margin entire and finely revolute; secondary veins brochidodromous or cladodromous. *Glands* fine, closely arranged in dark grey-black to greenish interrupted wavy lines or a mix of short and long lines, and sometimes also with dashes or small dots. *Inflorescences* terminal or axillary; flowers solitary or to 40 in thyrses or glomerules, flowers functionally dioecious; sepals and petals 2-5. *Male*: stamens in a central, annular mass or in 2-5 lobes or bundles, fused or separated; with or

without pistillode; anthers with 1, 2, 4 thecae or multi-thecous, sessile or with a short filament. *Female*: staminodes present or absent; ovary 2-13 locular, rarely unilocular, each locule with a solitary ovule; stigma raised or sessile sometimes elongated to 3 cm, discoid, flattened, convex or sunken, sometimes with a corrugated surface, margin with or without lobes, crenate or entire. *Fruit* a berry, rarely dehiscent, globose or ovoid, stigma with or without neck. Seed with fleshy edible pulp.

Distribution – Pantropical

## 6.6 Section Description

### *Garcinia* section *Garcinia*

*Garcinia* sect. *Mangostana* (Rumph. ex Gaertn.) Choisy, Mem. Soc. Hist. Nat. Paris 1 (1), (1823) 210 excl. *G. cambogia* (*G. gummi-gutta*) & *G. morella*; Planch. & Triana, Ann. Sci. Nat. Bot. ser. 4, 14 (1860) 325; Lanessan, Adansonia 10 (1871) 285; Lanessan, Mem. Gen. *Garcinia* (1872) 14; Pierre, Fl. Forest. Cochinch. 1 (1882) 11; Vesque in A. DC., Monogr. Phan. 8 (1893) 253, pro parte; Engler in Engler & Prantl, Nat. Pflanzenfam., ed. 2, 3(6), (1925) 221. — *Garcinia* subgenus *Mangostana* (Rumph. ex Gaertn.) Wight, Ill. Ind. Bot., 1 (1840) 120 excl. *G. merguensis*. — *Mangostana garcinia* Rumph. ex Gaertn., Fruct. Sem. Pl., 2(1) (1790) 105. — Type: *Garcinia mangostana* L.

*Garcinia* sect. *Conocentrum* Pierre, Fl. Forest. Cochinch. 1 (1882) 10; Engler, in Engler & Prantl, Nat. Pflanzenfam., ed. 2, 3(6), (1925) 224. — Lectotype (proposed here): *Garcinia malaccensis* Hook. f.

*Garcinia* sect. *Echinostigma* Pierre, Fl. Forest. Cochinch. 1 (1882) 10, pl. 60; Engler, in Engler & Prantl, Nat. Pflanzenfam., ed. 2, 3(6), (1925) 223. — Lectotype (proposed here): *Garcinia harmandii* Pierre.

*Garcinia* sect. *Gamodesmis* Pierre, Fl. Forest. Cochinch. 1 (1882) 10; Engler, in Engler & Prantl, Nat. Pflanzenfam., ed. 2, 3(6), (1925) 223. — Lectotype (proposed here): *Garcinia moselleyana* Pierre.

*Garcinia* sect. *Kiras* Pierre, Fl. Forest. Cochinch. 1 (1882) 12. — Lectotype (proposed here): *Garcinia celebica* L.

*Garcinia* sect. *Simosanthera* Pierre, Fl. Forest. Cochinch. 1 (1882) 18; Engler, in Engler & Prantl, Nat. Pflanzenfam., ed. 2, 3(6), (1925) 221. — Lectotype (proposed here): *Garcinia nitida* Pierre.

*Garcinia* sect. *Sphaerocentrum* Pierre, Fl. Forest. Cochinch. 1 (1882) 11. — Lectotype (proposed here): *G. cumingiana* Pierre [= *Garcinia venulosa* (Blanco) Choisy]

*Tree* or small tree to 25 m tall or bushy tree. *Exudate* yellow, white or cream. *Twigs* usually angled, stout, wrinkled lengthwise when dry. *Petiole* finely striate horizontally when dry, stout. *Lamina* leathery, pale brown or pale grey-brown, greenish, or reddish when dry, variable in shape but mainly elliptic, oblong or ovate; apex acute or acuminate; base acute to cuneate or decurrent; midrib flattened and inconspicuous, slightly or strongly raised above, strongly raised and sometimes angular (when dry) below; margin entire and finely revolute; secondary veins brochidodromous or sometimes cladodromous; tertiary veins fine and reticulate. *Glands* fine, closely arranged, dark grey, black or greenish, in interrupted short or long wavy lines, dashes and dots or a mixture, running from midrib to the margin across the secondary nervation, nearly parallel or not to the midrib. *Inflorescences*: terminal or rarely axillary, males with 1-18 flowers in a cluster; females usually solitary but also in groups of 2-3 flowers; bracts triangular, thickly leathery. *Male flower* with 4 sepals and petals or rarely 5 (*G. moselleyana*); leathery or papery; stamens in a central bundle, 4-angled or minutely to strongly 4-lobed; with or without pistillode, pistillode fungiform; anthers 2-theous, recurved, dehiscing by

longitudinal slits. *Female*: staminodes absent or present as fungiform or free stamen-like, surrounding the ovary; stigma raised or sessile, usually with corrugated surface, margin with lobes. *Fruit* globose or ovoid with thick or thin wall, stigma with or without neck; calyx persistent. Seed with fleshy edible pulp.

Distribution – 16 species and 5 varieties, eastern India to New Guinea.

Habitat & Ecology – Lowland to montane forest.

## 6.7 Identification Keys

### 6.7.1 Key to species from vegetative characters

1. a. Leaves with one intra-marginal vein or none.....3
- b. Leaves with two intra-marginal veins .....2
  
2. a. Secondary veins prominent, thick and raised on both sides; cultivated tree.....**9. *G. mangostana***
- b. Secondary veins visible and raised below, weakly raised above; wild tree.....**8. *G. malaccensis***
  
3. a. Leaf small, less than 1.0 cm wide .....4
- b. Leaf big, more than 1.0 cm wide.....5

4. a. Leaf very narrowly elliptic, length : breadth ratio > 3, to 8 cm  
long.....**2. *G. cataractalis***
- b. Leaf elliptic, length : breadth ration < 2, to 2.5 cm  
long.....**6. *G. exiguus***
5. a. Leaf mainly elliptic or ovate.....**6**
- b. Leaf spatulate or obovate.....**7. *G. harmandii***
6. a. Midrib raised above.....**7**
- b. Midrib flattened or slightly sunken above.....**10. *G. moselleyana***
7. a. Cross section of midrib (lower part of the lamina) square  
above.....**8**
- b. Cross section of midrib (lower part of the lamina) convex  
above.....**12**
8. a. Glandular lines invisible or visible but not raised below.....**9**
- b. Glandular lines prominent and raised below.....**13. *G. penangiana***
9. a. Glandular lines clearly visible below.....**10**
- b. Glandular lines invisible below.....**12. *G. ochraceus***
10. a. Glandular lines a mixture of long and short wavy lines, dashes and  
dots.....**11**
- b. Glandular lines dominated by long wavy lines, with occasional short  
lines.....**4. *G. diospyrifolia***

11. a. Glandular lines: many short wavy lines between long lines.....**11. *G. nitida***  
 b. Glandular lines: many dots and dashes between long lines.....**15. *G. sangudsangud***
12. a. Secondary nervation visible and distinct below.....**13**  
 b. Secondary nervation faint, inconspicuous or invisible below.....**5. *G. discoidea***
13. a. Leaves elliptic or ovate, twice as long as wide or less.....**14**  
 b. Leaves narrowly elliptic, 3 times as long as wide or more.....**1. *G. acuticosta***
14. a. Base of petiole without ligule-like appendage.....**15**  
 b. Base of petiole with ligule-like appendage.....**16. *G. venulosa***
15. a. Glandular lines closely arranged, c. 0.5 mm apart, usually not raised below.....**3. *G. celebica***  
 b. Glandular lines closely arranged, c. 1.0 mm apart, usually raised below.....**14. *G. rigida***

6.7.2 *Key to species for fertile specimens*

1. a. Fruit wall thick or thin but rigid, and not easily ruptured when dry; inflorescences solitary, or with up to 16 flowers; stamens in distinct 4-lobed bundles or in a bundle of 4-angled or conical-shape..... **2**

- b. Fruit wall thin and easily ruptured with fingernail when dry; inflorescences solitary, in pairs or triads and rarely fours; stamens in weakly 4-lobed bundles.....**Group IV**

***Key to species for Group IV***

- i. a. Leaves small, less than 1.0 cm wide.....**ii**  
 b. Leaves more than 1.0 cm wide.....**iii**
- ii. a. Fruit small, less than 11 mm long; stigma nearly sessile.....**6. *G. exiguus***  
 b. Fruit more than 11 mm long; stigma elongated, 6 mm long.....**2. *G. cataractalis***
- iii. a. Stigma sessile or raised to 2 mm long, flattened, convex-discoïd or skull-cap shaped.....**iv**  
 b. Stigma raised and elongated to 10 mm long, with flattened-discoïd or sunken surface.....**11. *G. nitida***
- iv. a. Stigma sessile, flattened-discoïd but slightly sunken in the middle, margin crenate.....**12. *G. ochraceus***  
 b. Stigma sessile or weakly raised to 2 mm long, skull-cap shaped, convex-discoïd or occasionally sunken, margin 4-6 lobed.....**4. *G. diospyrifolia***
2. a. Secondary veins brochidodromous or cladodromous with 1 intra-marginal vein or none; stigma on fruits with weakly lobed or irregularly dentate margin, raised to 8 mm or occasionally sessile; stamens 4-lobed, attached at the base of pistillode, pistillode in central .....**3**  
 b. Secondary veins brochidodromous with 2 intra-marginal veins, occasionally one intra-marginal vein; stigma on fruits distinctly lobed, square or wedge-shaped,

sessile or weakly sessile; stamens 4-angled or nearly cruciform, pistillode on top of stamen bundles or absent .....**Group I**

***Key to species of Group I***

- i.** a. Forest species found in the wild; mature fruit yellowish or reddish, turning brownish or reddish when dry; pistillode of male flower very small to 3 mm long, or absent.....**ii**

b. Cultivated species, not found in the wild; mature fruit dark purplish turning blackish when dry; pistillode of male flower distinct more than 5 mm long.....***G. mangostana***
  
- ii.** a. Twigs and pedicel stout; leaves with two intra-marginal veins, cross section of midrib convex; calyx of fruit thickly leathery, stigma bundles square, fused or separated; stamen bundles conical or square.....**iii**

b. Twigs and pedicel slender; leaves with intra-marginal veins, cross section of midrib square; calyx of fruit thinly leathery, stigma bundles wedge-shaped, fused; stamen bundles cruciform.....***G. penangiana***
  
- iii.** a. Found in Peninsular Malaysia, Sumatra and Borneo; stamens conical or square; pistillode very small on top of stamen bundle or absent; stigma on fruit weakly raised or sessile, surface corrugated, stigma bundles sometimes separate when mature.....**8. *G. malaccensis***

b. Found only in Philippines; stamens square; pistillode absent; stigma on fruit sessile, with smooth surface, stigma bundles not separate when mature.....**16. *G. venulosa***
  
- 3.** a. Secondary veins brochidodromous.....**4**

b. Secondary veins cladodromous .....**Group III**

***Key to Species of Group III***

- i. a. Inflorescences terminal; flowers with 4 sepals and petals; leaves mainly elliptic.....**7. *G. harmandii***
- b. Inflorescences terminal and axillary; flowers with 5 sepals and petals; leaves spatulate or obovate.....**10. *G. moselleyana***
  
- 4. a. Distinctly 4-lobed stamens; fruit wall thick or thin and stigma with lobes, sunken or convex surface; glandular lines long, wavy.....**5**
- b. Minutely 4-lobed stamens; fruit wall thick and stigma without lobes, sunken or flattened; glandular lines long, wavy with many dashes and dots.....**Group V [15. *G. sangudsangud*]**
  
- 5. a. Glandular lines distinct and raised below, c. 1 mm apart; fruit with thin wall, less than 0.5 mm thick..... **Group VI [14. *G. rigida*]**
- b. Glandular lines faint, not raised below, less than 1 mm apart; fruit wall thick, more than 1 mm thick.....**Group II**

***Key to species of Group II***

- i. a. Secondary nervation conspicuous and visible below; glandular lines fine but visible; sepals and petals thinly leathery; fruit usually drying brownish.....**ii**
- b. Secondary nervation inconspicuous or invisible below; glandular lines invisible or very faint; sepals and petals thickly leathery; fruit drying blackish.....**5. *G. discoidea***
  
- ii. a. Stigma raised to 2 mm long, discoid with nearly flattened surface, weakly 4-5 lobed, margin entire; male flower not known.....**1. *G. acuticosta***

- b. Stigma raised and elongated to 8 mm long, surface sunken or convex, 4-8 lobed, margin revolute; male flowers with 4-bundles of stamens.....**3. *G. celebica***

## 6.8 Species Descriptions

### 1. *Garcinia acuticosta* Nazre sp. nov. – Figure 6.1; Map 6.1.

— Type: Loh Hoy Shing FRI 13410 (holo KEP; iso L). Selangor, Peninsular Malaysia.

(Etymology: Lat. from sharp midrib when dry)

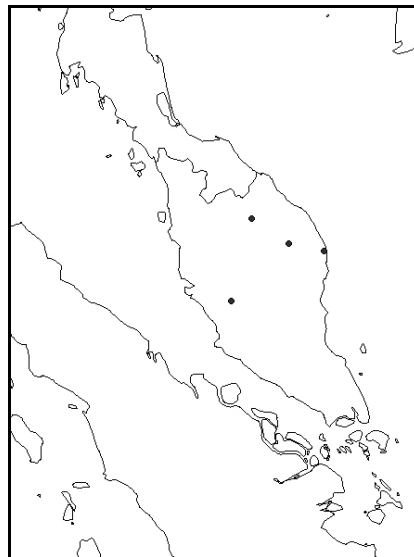
*Small tree* to 10 m tall. *Bole*: bark greyish-brown or reddish, flaking; inner bark reddish-brown; sapwood yellowish-red. *Exudate* white turning yellow. *Twigs* angled, stout, strongly longitudinally striate when dry, dark brown or chocolate to pale brownish (greenish-yellow in new specimens). *Petiole* finely striate horizontally, stout, darker than the leaves, 0.8-1.4 cm long. *Lamina* thickly to thinly leathery, pale brown or pale grey-brown, darker and shiny above, narrowly elliptic, 16.0-21.0 x 1.8-2.4 cm; apex acute or bluntly acuminate; base acute to cuneate; midrib nearly same colour as petiole, weakly raised above, raised and sharply angled in cross section below when dry; margin entire and very finely revolute; secondary veins fine, visible above and inconspicuous below, brochidodromous, closely arranged, weakly raised; tertiary veins nearly invisible, fine and very faint. *Glands* fine, dark grey-black to greenish interrupted wavy lines, nearly parallel to the midrib, running across the secondary veins to the margin. *Inflorescences* probably terminal. *Male flower* not seen. *Female flower* not seen. *Fruit* terminal, solitary,

ovoid, green turning yellow, surface smooth and wall thick, 5.0-5.3 x 2.3-3.7 cm; with 4 prominent thick leathery sepals, to 1.3 cm long; stigma weakly raised to 2 mm or sessile, black, discoid, flattened or slightly convex with 4-5 weak lobes, and hardened, to 5.5 mm across.

Distribution –Peninsular Malaysia.

Habitat & Ecology – Lowland and hill forest to 500 m, rare.

Notes – Previously, material of this species has been placed in *G. opaca* King (synonym of *G. diospyrifolia*) though it has very different leaves and fruit. The dry leaf usually has a raised and sharp midrib on the abaxial surface when dry, and the fruit walls are thicker than in *G. diospyrifolia*. Molecular data suggest that it is closely related to *G. discoidea* but it differs in leaf shape and the glandular lines are more widely spaced.



Map 6.1. Distribution of *Garcinia acuticosta* Nazre.

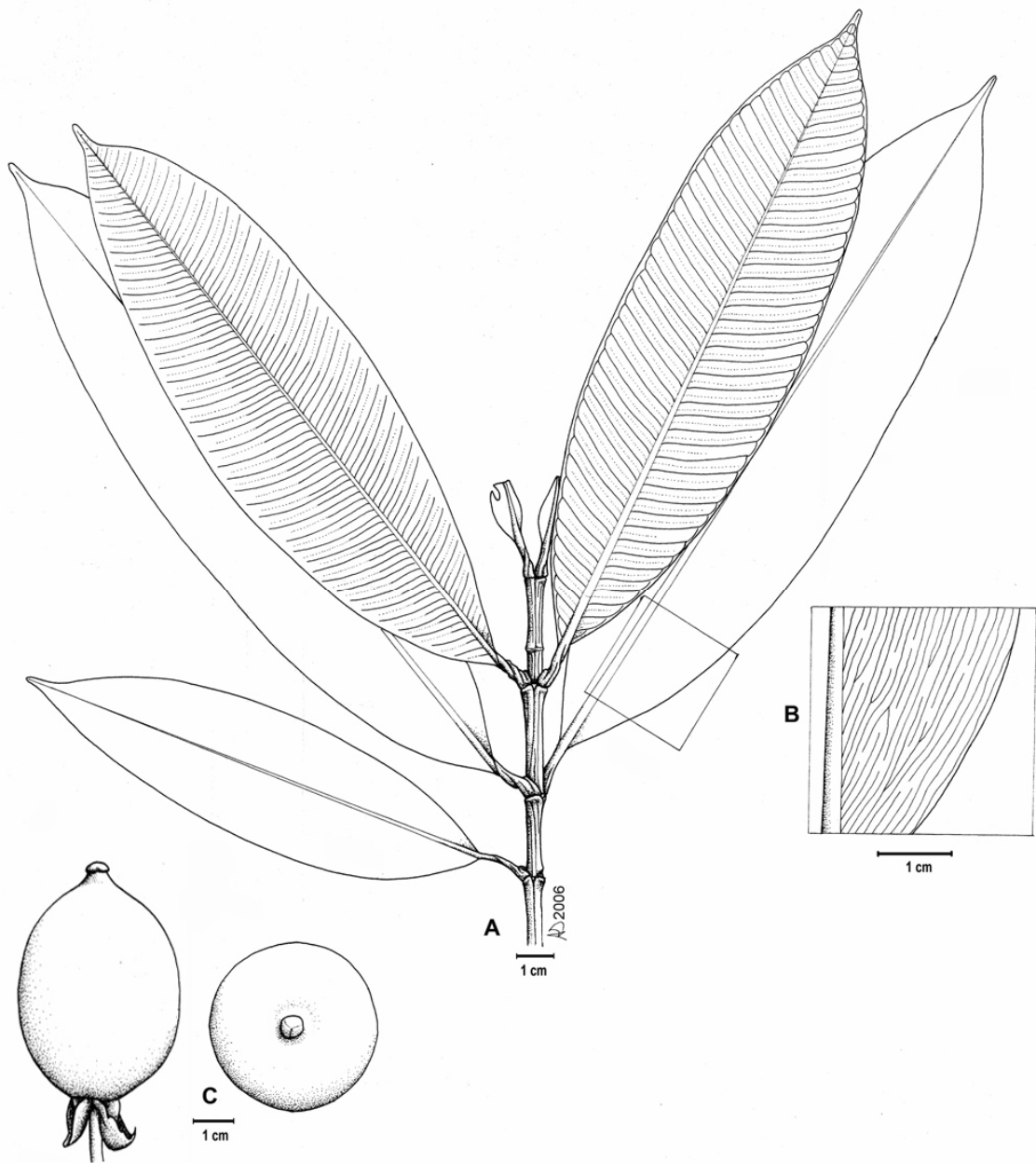


Figure 6.1. *Garcinia acuticosta* Nazre. **A.** habit (male); **B.** glandular lines; **C.** fruit and stigma [Nazre BB01, E]. Drawn by Anna Dorward.

**2. *Garcinia cataractalis* Whitmore** – Figure 6.2; Map 6.2.

*Garcinia cataractalis* Whitmore, Gard. Bull. Singapore 26(2) (1973) 274. — Type *Whitmore FRI* (holo KEP, iso K) Terengganu, Peninsular Malaysia.

*Bushy tree* to 2.5 m tall. *Twigs* angled, slender, longitudinally striate when dry, pale grey-brown to greenish. *Petiole* finely striate horizontally, pale grey-brown to greenish and whitish, 0.3-0.7 cm long. *Lamina* thinly leathery, pale brown or brown, darker and shiny above, very narrowly elliptic, 2.5-7.4 x 0.3-0.8 cm; apex acute or attenuate; base cuneate or decurrent onto petiole; midrib raised above, raised and sharp below; margin entire and finely revolute; secondary veins very faint above, invisible below; tertiary venation not visible. *Glands* fine, closely arranged and slightly raised below, dark grey-black to brown, continuous and interrupted wavy lines, nearly parallel to the midrib. *Inflorescences* perhaps terminal. *Male flower* not seen. *Female flower* not seen. *Fruit* globose to ovoid, weakly 2-3 lobed when dry, dark maroon or reddish-brown, smooth, 1.9-2.1 x 1.8-2.0 cm; stigma elongated to 6 mm long, skull-cap shaped with 4-5 weak lobes, blackish, to 4 mm across; four persistent, leathery, elliptic sepals, 0.5-0.6 x 0.3-0.5 cm; seeds 4 with a translucent aril, sour.

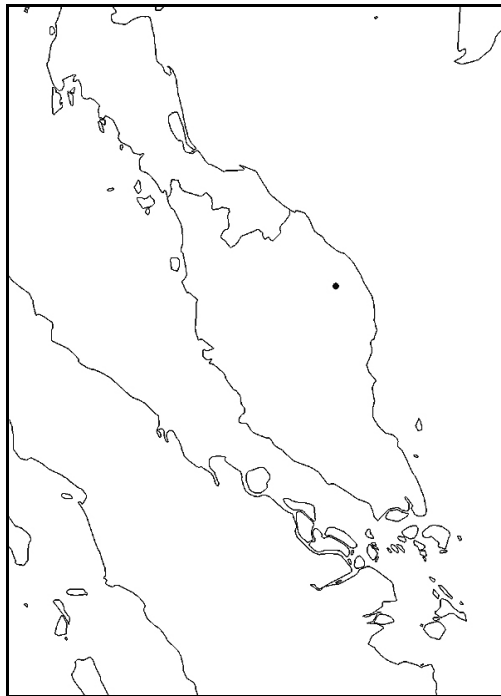
Distribution – Peninsular Malaysia.

Habitat & Ecology – The only rheophytic species in the section, usually found on flooded riverbanks.

Note – An endemic species only recorded in Peninsular Malaysia. *G. cataractalis* is a new inclusion to sect. *Garcinia* not previously listed by Jones (1980). Whitmore (1973) described the fruit as being terminal or axillary but I have yet to find any specimens

with the fruit borne in axils. This is an easily identified species which is characterised by being a dense and bushy small tree with very narrow, small, elliptic leaves.

The fruits resemble those of *G. diospyrifolia* and *G. nitida* but it is more closely related to *G. diospyrifolia* with which it shares similar vegetative characters of glandular lines and the nervation on the leaves. Molecular analysis and morphology supported the inclusion of *G. cataractalis* in sect. *Garcinia*.



Map. 6.2. Distribution of *Garcinia cataractalis* Whitmore.

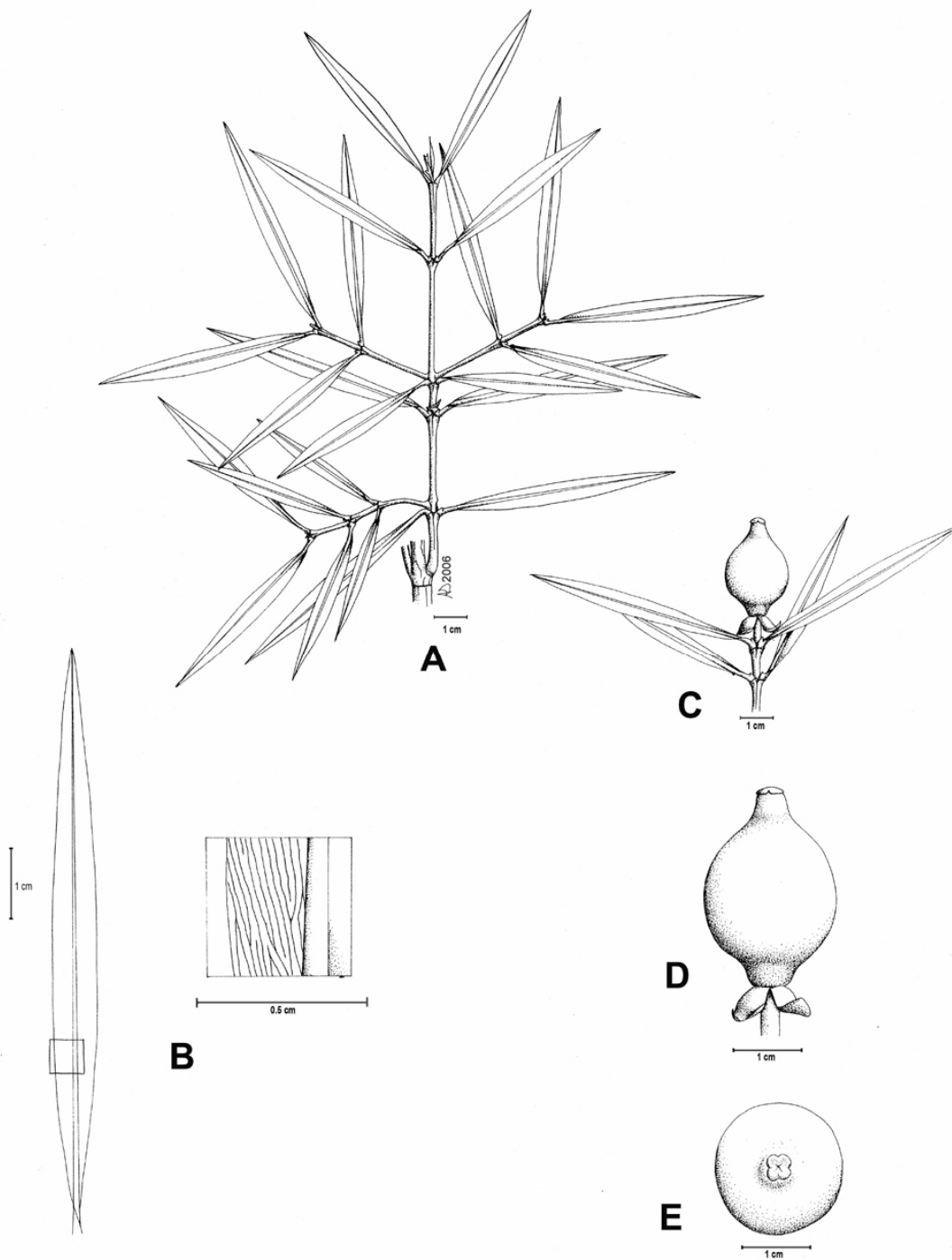


Figure 6.2. *Garcinia cataractalis* Whitmore. **A.** habit (female); **B.** lamina and glandular lines; **C,** **D, E.** fruits and stigma. [Whitmore FRI 20350, K]. Drawn by Anna Dorward.

### 3. *Garcinia celebica* L. – Figure 6.3; Map 6.3.

*Garcinia celebica* L. in Stickman, Herb. Amboin. (Linn.) (1754) 119; Linnaeus, Syst. Nat., ed. 12 (1767) 326 & in Murray, Syst. Veg. ed. 13 (1774) 368; Miquel, Fl. Ned. Ind. 1 (1859) 507; Planch. & Triana, Ann. Sci. Nat. ser. 4, 14 (1860) 173; Pierre, Fl. Forest. Cochinch. 1 (1882) 13; Vesque, Monogr. Phan. 8 (1893) 404; Boerlage, Cat. Pl. Phan. 2 (1901) 69; Koorders & Valetton, Bijdr. Boomsoort. Java 9 (1902) 367; Merrill in Interpr. Herb. Amb. (1917) 373; Backer & Bakhuizen, Fl. Java (Spermatoph.), (1963) 387. — *Mangostana celebica* Rumph. ex Gaertn., Fruct. Sem. Pl. 2(1), (1790) 105. — Lectotype (proposed here): "*Mangostana celebica*" in Rumphius, Herb. Amboin. (Rumphius) 1 (1741) 134, pl. 44.

*Garcinia cornea* L. in Murray, Syst. Veg. ed. 13 (1774) 368; Blume, Bijdr. Fl. Ned. Ind. (1825) 214; Roxb., Fl. Ind. ed. 2, 2 (1832) 629; Miquel, Fl. Ned. Ind. 1 (1859) 506; Lanessan, Mem. Gen. Garcinia (1872) 21; Pierre, Fl. Forest. Cochinch. 1 (1882) 12 & Fl. Forest Cochinch. 1 (1883) pl. 78B; Merrill in Interpr. Herb. Amb. (1917) 374. — *Garcinia porrecta* Anders. in Lanessan, Mem. Gen. Garcinia (1872) 21, nom. nud. — *Garcinia porrecta* Laness. in Vesque, Monogr. Phan. 8 (1893) 397, nom. illegit. — Lectotype (proposed here): "*Lignum corneum*" in Rumphius, Herb. Amboin. (Rumphius) 3 (1743) 55, pl. 30.

*Garcinia speciosa* Wall., Pl. Asiat. Rar. 3(11) (1832) 37, pl. 258 ; Planch. & Triana, Ann. Sci. Nat. ser. 4, 14 (1860) 171; Hook f., Fl. Brit. India 1 (1872) 260; Kurz, Forest. Fl. Burma (1877) 88; Pierre, Fl. Forest. Cochinch. 1 (1882) 14 & Fl. Forest Cochinch. 1 (1883) pl. 79I & 79H; King, J. Asiat. Soc. Bengal, Pt. 2 Nat. Hist. 59 (1890) 154; Vesque, Monogr. Phan. 8 (1893) 402; Parkinson, Forest. Fl. Andaman Isl. (1923) 90; Craib, Fl. Siam 1 (1931) 117; Maheshwari, Bull. Bot. Surv. India 6 (1964) 123. — Lectotype (proposed here): *Wallich 4855* (K), Mertaban & Amherst, Burma.

*Garcinia fabrilis* Miq., Fl. Ned. Ind. Eerste. Bijv. 496 (1861). — Lectotype (proposed here): *Diepenhorst HB 2152* (L), Priaman, Sumatra, Indonesia.

*Garcinia basacensis* Pierre, Fl. Forest. Cochinch. 1 (1882) 15, pl.58; Vesque, Monogr. Phan. 8 (1893) 398. — Lectotype (proposed here): *Harmand 1074* (lecto P; iso K) Laos.

*G. benthami* Pierre, Fl. Forest. Cochinch. 1 (1882) 12, pl.55 & 56; Vesque, Monogr. Phan. 8 (1893) 392; Boerlage, Cat. Pl. Phan. 2 (1901) 70. — Lectotype (proposed here): *Pierre 700* (lecto P [barcode P329872]; iso A, K), Cambodia.

*Garcinia ferrea* Pierre, Fl. Forest. Cochinch. 1 (1882) 12, pl.57. — Lectotype (proposed here): *Pierre 3634* (lecto P), Cambodia.

- Garcinia hombroniana* Pierre, Fl. Forest. Cochinch. 1 (1882) 12 & Fl. Forest Cochinch. 1 (1883) pl.79D, 79E & 79F; King, J. Asiat. Soc. Bengal, Pt. 2 Nat. Hist. 59 (1890) 155; Vesque, Monogr. Phan. 8 (1893) 395; Ridley, Fl. Malay Penin. 1 (1922) 171; Engler, Nat. Pflanzenfam. ed. 2, 21 (1925) 223; Craib, Fl. Siam. 1 (1931) 115; Whitmore, Tr. Fl. Malaya 2 (1973) 212. — Lectotype (proposed here): *Hombron s.n.* (P), Malacca, Peninsular Malaysia.
- Garcinia riedeliana* Pierre, Fl. Forest. Cochinch. 1 (1882) 12, Fl. Forest. Cochinch.1 (1883) pl.79A. — Lectotype (proposed here): *Riedel s.n.* (K) Gorontalo, Sulawesi.
- Garcinia rumphii* Pierre, Fl. Forest. Cochinch. 1 (1882) 13 & Fl. Forest Cochinch. 1 (1883) pl.77A; Vesque, Monogr. Phan. 8 (1893) 400. — Lectotype (proposed here): *Treub 4169* (P), Bangka Island, Indonesia.
- Garcinia kurzii* Pierre, Fl. Forest. Cochinch. 1 (1882) 14 & Fl. Forest Cochinch. 1 (1883) pl.78C. — *Garcinia speciosa* Wall., King, J. Asiat. Soc. Bengal, Pt. 2 Nat. Hist. 59 (1890) 155; Vesque, Monogr. Phan. 8 (1893) 403. — Lectotype (proposed here): *Kurz 24* (P), South Andaman
- Garcinia affinis* Pierre, Fl. Forest. Cochinch. 1 (1882) 15 & Fl. Forest Cochinch. 1 (1883) pl.78C & 79G, nom. illegit.
- Garcinia jawoera* Pierre, Fl. Forest. Cochinch. 1 (1882) 37. — Lectotype (proposed here): *Pierre 4607* [= Blume 1273] (lecto P; iso L), Java towards Tandjoor, Indonesia.
- Garcinia krawang* Pierre Fl. Forest. Cochinch. 1 (1882) 37. — Lectotype (proposed here): *Korthals 1313a* (P), Lampei Island, S. Kalimantan, Indonesia.
- Garcinia kingii* Pierre ex Vesque, Monogr. Phan. 8 (1893) 407. — Lectotype (proposed here): *King's Collector s.n.* (K), Andaman Island.
- Garcinia porrecta* Wall. ex Vesque, Monogr. Phan. 8 (1893) 390, nom. inval. — *Garcinia porrecta* Wall., Numer. List (1831) 4852, nom. nud.

*Tree* to 30 m tall. *Bole* fluted, sometimes with steep buttresses dark brown to pale grey-brown, fissured and flaking to expose cream bark; inner bark pink. *Exudate* white, cream or yellow. *Twigs* slightly angled or distinctly 4-angled, drying brownish or reddish brown, striate longitudinally. *Petiole* stout, finely horizontally striate, dark brown or yellowish to greenish, especially in newly collected specimens, 7-23 mm long. *Lamina*

variable in shape and size, from elliptic or broadly elliptic to lanceolate, sub-orbulate or round, 3.3-17.5 x 1.7-11.0 cm, leathery, pale brown or reddish brown when dry but rather greenish in new specimens, usually paler below; apex acute, acute-acuminate, shortly acuminate or blunt; base cuneate to slightly decurrent, obtuse or acute; margin entire, weakly and finely revolute; midrib weakly or strongly raised above, raised and slightly pointed below; secondary veins visible or faint on both sides, slightly raised, brochidodromous; tertiary veins conspicuous, reticulate. *Glands* visible below as fine, black, pale green or brown, closely arranged, c. 0.5 - 0.8 mm apart, wavy, lines running about 45 degrees from the midrib towards the margin. *Inflorescences* terminal; males in clusters of 2-14(18) flowers of different sizes, pedicel slender or stout, angular, to 2.0 cm long, female flowers usually solitary or sometimes in pairs or triads; sepals and petals four; bracts triangular, to 1.7 cm long. *Male flowers* sometimes fragrant, pedicel slender to stout, four angled, variable in size to 25 mm across when open; sepals green to yellowish green, concave, ovate, leathery, usually thicker than petals, 5-8 x 6-7 mm; petals cream to yellowish, oblong, ovate or obovate-elliptic, thinly papery or leathery, 4.3-5 x 9-10 mm; stamens in four lobed bundles, opposite petals; anthers 2-theous, nearly sessile; pistillode dark blackish, fungiform, stipe short to 3 mm long or sometimes nearly sessile. *Female flower* solitary, 10-12 x 12-15 mm across; sepals ovate, concave, thickly to thinly leathery, 5-9 x 4-6 mm, brown; petals green yellowish, obovate or broadly elliptic, thinly coriaceous; staminodes absent; ovary sub-globose or flask-shaped, locules 4-6; stigma discoid or weakly concave, to 5 mm wide. *Fruits* ovoid to ellipsoid or globose, sometimes shiny, smooth or occasionally wrinkled, green but reddish when mature, pale cream brown or brown to reddish brown when dry, variable in size, to 5.0

cm across; stigma usually raised to 8.0 mm, surface concave or weakly convex, smooth or wavy and corrugated, margin minutely 4-8 lobed; calyx persistent, concave and leathery; seeds 4-6 with a white aril.

Distribution – Eastern India to Malesia

Habitat & Ecology – Mainly found on the coast and in lowland forest near the sea but also distributed in inland forest to lower montane forest (1200 m alt.). Occasionally, also found on limestone.

Notes – The taxonomic confusion and resulting long synonymy of *G. celebica* has two sources;

1) Nomenclatural problems

The epithet *celebica* has priority in the long list of synonyms. The species was called *Mangostana celebica* by Rumphius (1741) in his pre-Linnaean Herbarium

Amboinensis. Stickman (Linnaeus, 1754), who revised Rumphius's work, validated the name and placed this species under *Garcinia* together with *G. mangostana* L.

Some time later Murray (Linnaeus, 1774) added another Rumphius species, *Lignum corneum*, as *G. cornea* L. According to Murray (Linnaeus, 1774), *G. cornea* and *G.*

*celebica* differ from each other by their inflorescences (trifloris vs. unifloris). This distinction does not work, especially in the male flowers. There is no surviving type specimen to accompany these two species except in the descriptions and figures by

Rumphius (1741). Based on this, other works, such as Miquel (1859 & 1860) and

Pierre (1882-1885), acknowledged *G. celebica* and *G. cornea* as two different species and further described new closely related species (e.g. *G. fabrilis* Miq., *G.*

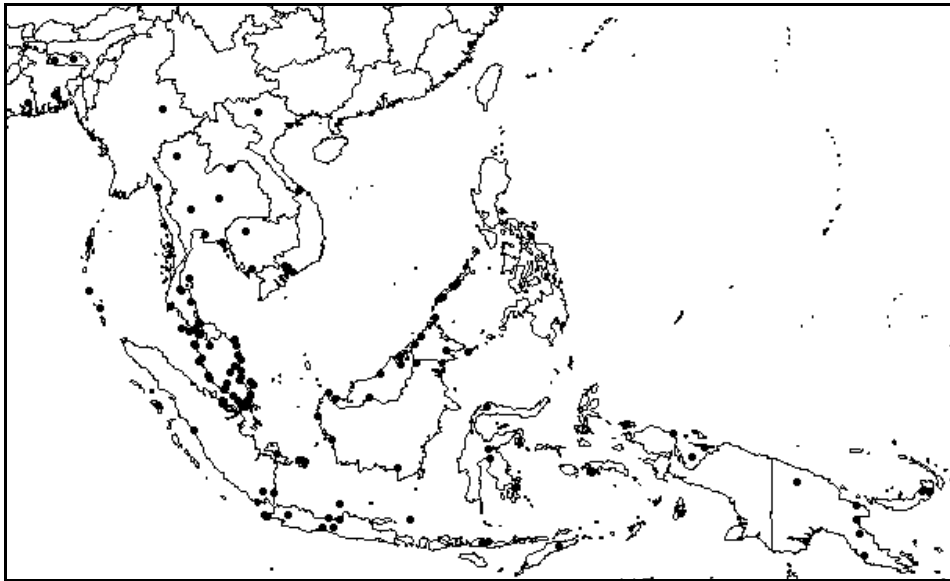
*hombroniana* Pierre) defined by their geographical distributions. Confusion over the

true identity of the Rumphius species may be one of the reasons why numerous other species have been described from throughout its distribution. Most new names (*G. affinis*, *G. basacensis*, *G. benthami*, *G. calleryi*, *G. ferrea*, *G. hombroniana*, *G. jawoera*, *G. krawang*, *G. kurzii*, *G. rumphii*) were created by Pierre (1882-1885) largely based on geographical distribution, the latex colour and the position of the stamen lobes (opposite petals or sepals). The characters that unite all the previous names are in the male flowers and the fruit: the stamens being in four distinct lobes surrounding the pistillodes, thinly leathery sepals and petals; and mature fruits usually with a neck and a sunken stigma with lobes.

## 2) The usage of characters and discontinuity of variation

The usage of the stamen lobes as an important character by Pierre (1882-1885) when describing new species from different geographic regions is misleading because some specimens he observed were still in bud and the mature bundle positions cannot be ascertained at this stage. The colour of the latex is also not a stable character: for example, within *G. celebica* (*G. hombroniana*) in Peninsular Malaysia, Whitmore (1973) found that the colour could be white, opaque or yellow, though the cause of this variability is not known. There is also more variability in the other characters used by Pierre (1882-1885) and Whitmore (1973): for example the length of the pedicel, the shape of the fruit and the degree of elongation (neck) of the stigmas. I found that there are no clear cut discontinuities in these morphological characters or any characters exclusive to certain geographical areas. One explanation for this variation is facultative agamospermy in *G. celebica* which

has been confirmed by Richards (1990b) in a population in Peninsular Malaysia. However, proving the effect of agamospermy on variation by using herbarium materials is impossible. It will be interesting if further study could look at this problem, especially at the population level.



Map 6.3. Distribution of *Garcinia celebica* L.

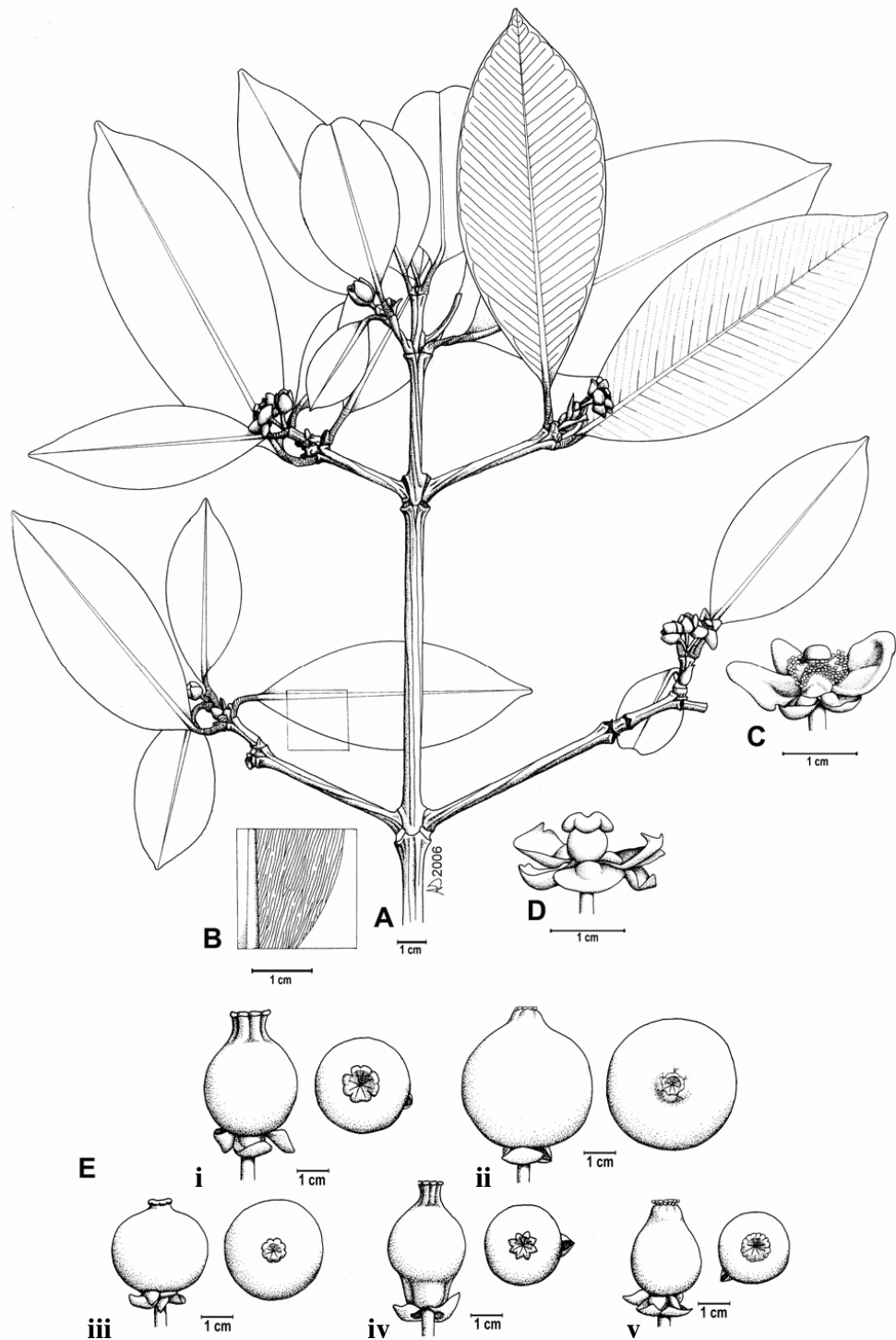


Figure 6.3. *Garcinia celebica* L. **A**, Habit (male); **B**, glandular lines; **C**, male flower; **D**, female flower; **E**, fruit and stigma. [(A & B: Curtis 690, SING); (C: Nazre 062, E); (D: Nazre s.n., E); (Ei: Burkill HMB259, SING); (Eii: Ahmad Shukor AS29, SING); (Eiv: Yahya SFN21419, SING); (Ev: Pierre 700, K)]. Drawn by Anna Dorward.

#### 4. *Garcinia diospyrifolia* Pierre – Map 6.4.

*Garcinia diospyrifolia* Pierre, Fl. Forest. Cochinch. 1 (1882) 37. — Lectotype (proposed here):

*Korthals s.n.* (lecto L; iso K, UC), Gunung Sekubang, Kalimantan.

Slender or bushy, medium sized tree to 15 m tall. *Exudate* white or light yellow. *Twigs* slender, angled, longitudinally wrinkled when dry, brownish red to brown or yellowish. *Petiole* finely horizontally striate, close and nearly parallel, dark brown to black or reddish brown, 0.7-1.6 cm long. *Lamina*: leathery, dull dark green above, green below, and when dry turning reddish brown or grey brown to brownish, darker and slightly shiny above, pale below, elliptic to narrowly elliptic, elliptic-oblong or elliptic-obovate, 8.0-13.0 x 2.5-5.5 cm; usually with caudate apex, sometimes attenuate or acuminate, to 20 mm long; base cuneate or decurrent onto petiole; midrib raised but with flattened surface or nearly square in cross section above, especially at the base of the lamina, raised and sharp below; margin entire and finely revolute. *Nervation*: secondary veins visible, fine and closely arranged, sometimes conspicuous above, brochidodromous; tertiary veins invisible. *Glands* fine but clearly visible below (not visible on the older specimens), dark brown to blackish short and long wavy lines, running across the secondary veins from midrib to the margin. *Inflorescences* terminal, male flowers 1-3, female solitary; sepals and petals 4. *Male flower*: rarely found, pedicel slender, slightly angular, to 10 mm long, sepals yellowish turning brown when dry, concave, ovate, leathery, usually thicker than petals, 5-6 x 6-7 mm; petals reddish turning dark-maroon to brownish when dry, oblanceolate or obovate, 4.0-7.5 x 5.0-6.5 mm; stamens in four weak lobes not open or weakly open at maturity, enclosing the pistillode; pistillode dark blackish, fungiform in

shape; anther 2-theous. *Female flower* with a flask-shaped ovary to 10 mm long, tipped with a convex or skull-cap shaped stigma, margin wavy; staminodes absent. *Fruit*: ellipsoid, ovoid or flask shaped, reddish-brown or bright maroon, wall smooth, thin, easily ruptured when dry, to 3.7 x 5.0 cm; calyx leathery, tipped by a small skull-cap shaped, convex-discoid or occasionally sunken stigma, sessile or weakly raised to 2 mm long, to 2 mm across, margin with 4-6 lobes; sepals thinly leathery, reddish brown, elliptic, concave; seeds 4, with white aril.

Distribution – Peninsular Malaysia and Borneo.

Habitat & Ecology – A common treelet or small tree from lowland to lower montane forest.

Notes – *G. diospyrifolia*, described by Pierre (1882-1885) from Borneo (Kalimantan), and *G. opaca*, described by King (1891) from Peninsular Malaysia, are the same species. Whitmore (1973) created a new variety of *G. opaca* (var. *dumosa*) differing mainly in the overall appearance of the tree in Peninsular Malaysia (bushy vs. tree) and listed specimens that he studied. Curiously, he included KEP 97842 collected from Gunung Benom which is a tree.

*Key to varieties*

1. a. Slender or bushy tree to 12 m tall.....**2**
  - b. Medium sized tree to 20 m tall.....**c. var. arborea**
  
2. a. Slender tree to 7 m tall; fruit turning reddish on drying, ovoid or flask-shaped, stigma raised.....**a. var. diospyrifolia**

b. Bushy tree to 12m tall; fruit turning brownish on drying, globose, stigma sessile.....**b. var. *minor***

**a. var. *diospyrifolia*** – Figure 6.4.

*Garcinia opaca* King, J. Asiat. Soc. Bengal, Pt. 2 Nat. Hist. (1890) 160. — Lectotype (proposed here): *King's Collector 7232* (lecto K; iso P), Perak, Peninsular Malaysia.

*Garcinia opaca* var. *dumosa* Whitmore, Gard. Bull. Singapore 26 (1973) 277. — Type: *Mohamad Shah bin Mohamad Nur 1560* (holo SING) Bukit Terom, Pahang, Peninsular Malaysia.

Slender tree to 7 m tall. *Lamina* 9.0-15.0 x 3.5-5.0 cm; midrib raised, square in cross section above. *Fruit* green turning reddish or dark maroon when dry; ovoid to flask-shaped, stigma discoid or skull-cap shaped, flattened or sunken, margin lobed.

Distribution: Peninsular Malaysia and Borneo.

**b. var. *arborea*** Nazre var. nov. – Figure 6.5.

— Type: *Rahim Ismail KEP 97842* (holo KEP; iso K, L) Gunung Benom, Pahang, Peninsular Malaysia.

(Etymology: Lat. from its stature, a tree)

Small to medium sized tree to 20 m tall. *Lamina* 7.0-12.5 x 2.0-5.0 cm; midrib raised, square in cross section above. *Fruit* green turning reddish brown when dry; ovoid, stigma discoid, lobes not distinct.

Distribution: Peninsular Malaysia and Borneo.

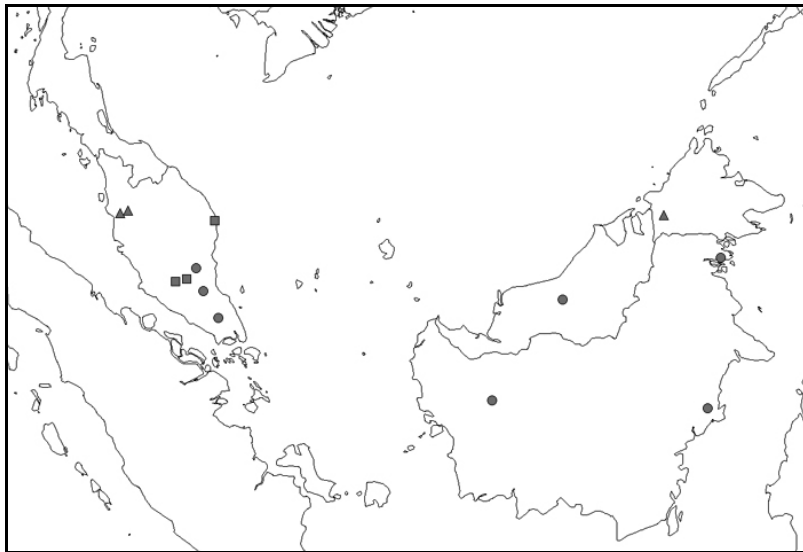
c. var. **minor** Ng ex Nazre var. nov. – Figure 6.6.

— Type: *K.M. Wong 32216* (holo KEP) Pasoh, Negeri Sembilan, Peninsular Malaysia.

(Etymology: Lat. from its small leaf)

Treelet and bushy tree to 12 m tall; bole greenish, to pale brown, smooth to scaly. *Lamina* small, 3.5-8.5 x 1.7-4.5 cm; midrib raised above. *Fruit* green turning red, smooth, brownish when dry with distinct dark reddish-brown or blackish longitudinal wavy lines, very closely arranged; globose or ellipsoid, to 3.5 cm across.

Distribution: Peninsular Malaysia



Map 6.4. Distribution of *Garcinia diospyrifolia* Pierre. *G. diospyrifolia* var. *diospyrifolia* (circle); *G. diospyrifolia* var. *arborea* (triangle); *G. diospyrifolia* var. *minor* (square).

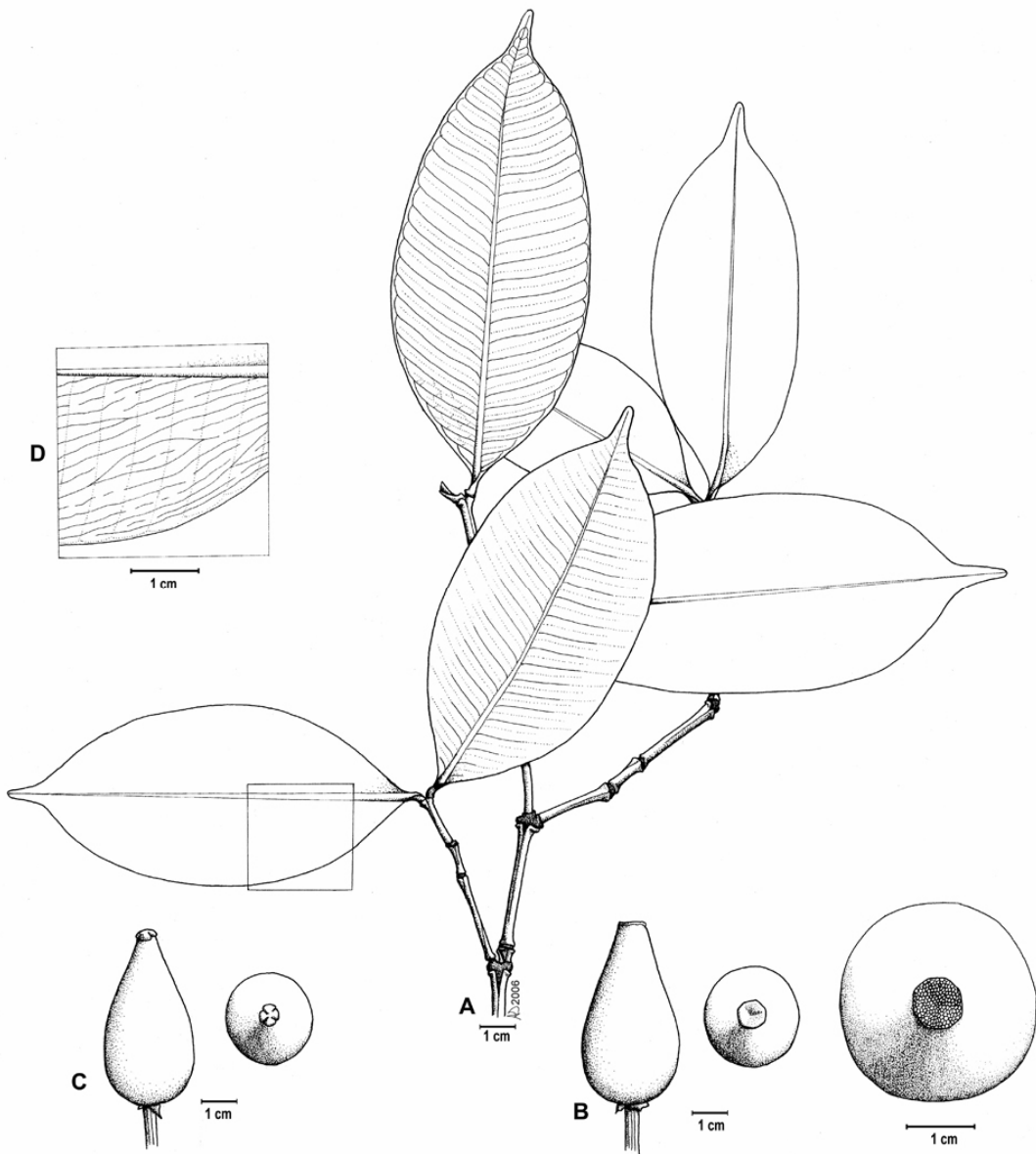


Figure 6.4. *Garcinia diospyrifolia* var. *diospyrifolia*. A. habit (female); B, C. fruit and stigma; D. glandular lines. [(A, B, C, D: Soegeng 47, L); (B: Kochummen 7773, L)]. Drawn by Anna Dorward.

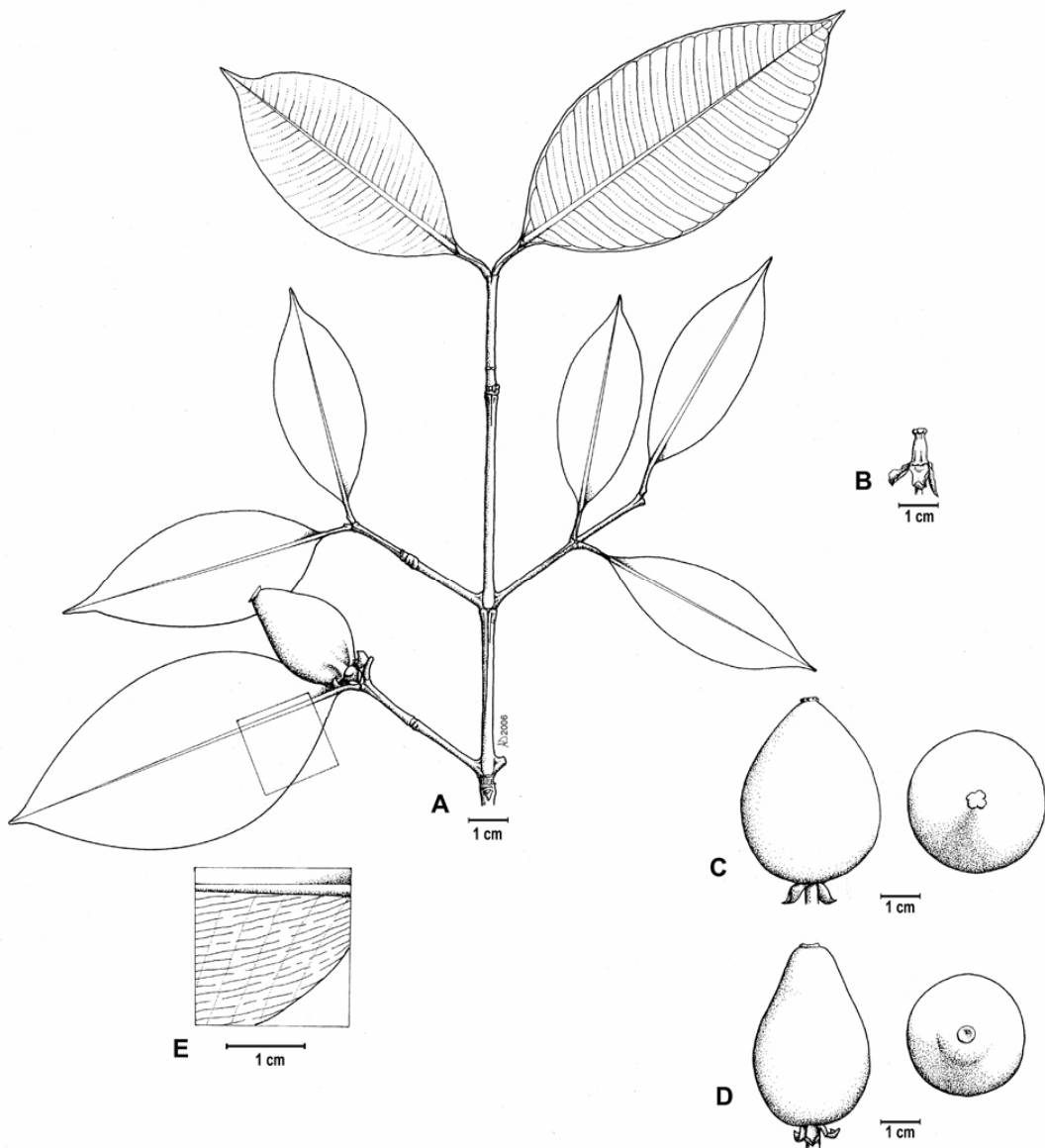


Figure 6.5. *Garcinia diospyrifolia* var. *arborea* Nazre. **A.** habit (female); **B.** female flower; **C, D.** fruit and stigma; **E:** glandular lines [(A, B, C, E: King's Collector 7232, P); (D: Pereira JTP65, E)]. Drawn by Anna Dorward.

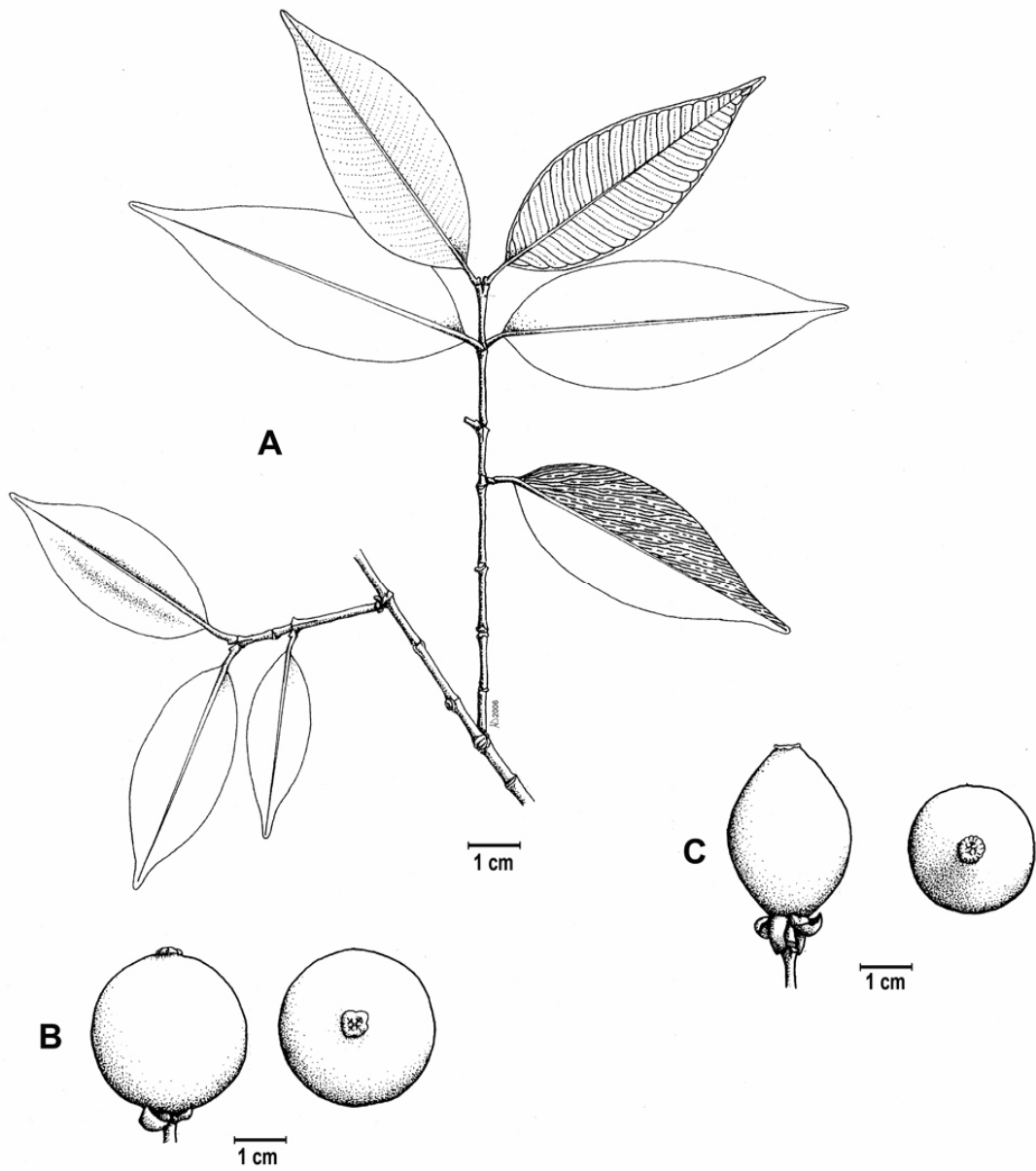


Figure 6.6. *Garcinia diospyrifolia* var. *minor* Ng ex Nazre. **A.** habit (female); **B, C.** fruit and stigma [(A, B: Wong 32216, A); (C; Rosemary 541, L)]. Drawn by Anna Dorward.

**5. *Garcinia discoidea* Nazre sp. nov. – Figure 6.7; Map 6.5.**

*Garcinia hombroniana* variety A Whitmore, Gard. Bull. Singapore 26 (1973) 277. — Type:

*Corner 34940* (holo SING), Johor, Peninsular Malaysia

(Etymology: Lat. from discoid shape of stigma)

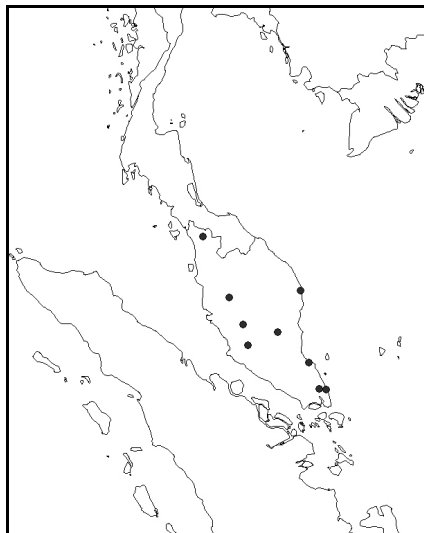
*Tree* to 20 m tall. *Bole*: bark greyish brown, dippled and flaking, inner bark reddish brown, sapwood yellowish red. *Exudate* white turning pale yellow or cream. *Twigs* angled, stout, longitudinally wrinkled when dry, dark brown to yellowish or pale green. *Petiole* finely striate horizontally, stout, dark brown to reddish brown (or brownish-yellow to yellowish in new specimens), 1.3-2.5 cm long. *Lamina* thickly leathery, dark brown to brown-maroon (pale brown or pale grey-brown to greenish in new specimens), darker and shiny above, elliptic to broadly elliptic, oblong or ovate, 7.6-20.6 x 2.7-9.3 cm; apex blunt, acute or acuminate; base obtuse to cuneate; midrib same colour as petiole, slightly raised above, raised and sharp below; margin entire and finely revolute; secondary veins fine, visible above but inconspicuous and nearly invisible below, closely arranged, brochidodromous; tertiary veins invisible. *Glands* very fine, inconspicuous, visible on new specimens especially at the base of the leaf, invisible on much older specimens, closely arranged dark grey-green to blackish, interrupted wavy lines, nearly parallel to the midrib, running from midrib across secondary veins to margin. *Inflorescences* possibly terminal. *Male flowers*: rarely found, sepals and petals 4, very thickly leathery, outer usually thicker; pedicel to 15 mm long, stout, angled; sepals concave, broadly ovate, 9.5-11.5 x 11.0-13 mm; petals broadly ovate or oblanceolate, 12.0-13.0 x 11.0-14 mm; stamens arranged in 4-lobed central bundle surrounding the

pistillode; pistillode fungiform; anthers 2-theous, sessile. *Female* flower not seen. *Fruit* solitary and terminal, ellipsoid or ovoid, green when unripe, pale to dark brown or blackish maroon when dry, smooth and finely horizontally striate or sometimes with fine lenticels, 2.0-5.6 x 0.6-4.0 cm; tipped by discoid black stigma, slightly raised to 3 mm, surface flattened or sometimes sunken in the middle, weakly 4-lobed, to 10 mm across; with thick leathery prominent sepals, always curved downwards; seeds 1-4 with a white aril.

Distribution – Peninsular Malaysia widespread but uncommon.

Habitat & Ecology – Hill forest from 600 m to montane forest. Occasionally found in swamp forest.

Notes – The male flowers closely resemble *Garcinia celebica* L. with stamens in 4 lobes. The flowers of *Garcinia discoidea* are, however, much thicker and leathery. The leaf differs from *G. celebica* by being much thicker and the secondary veins are almost invisible on the lower surface.



Map 6.5. Distribution of *Garcinia discoidea* Nazre

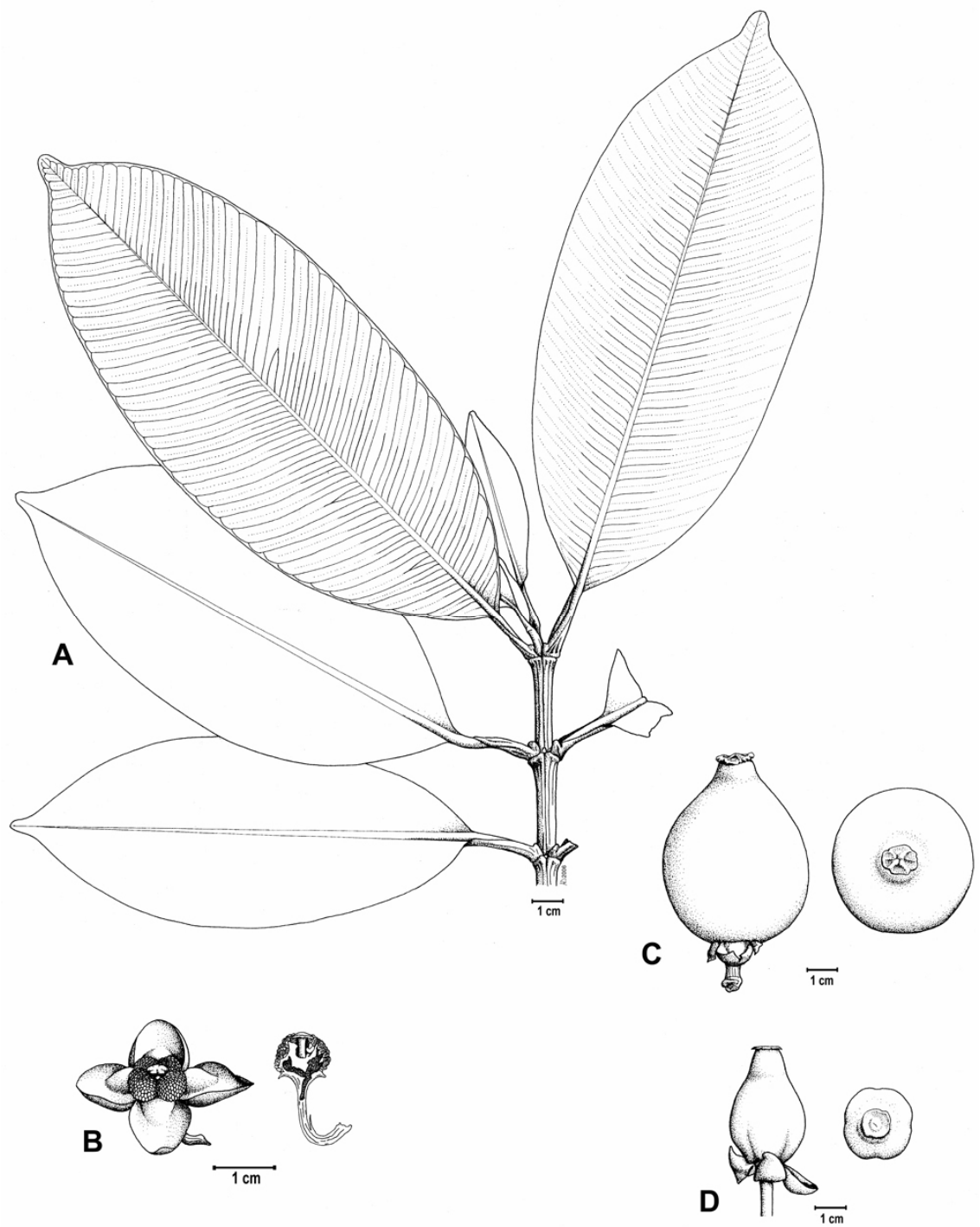


Figure 6.7. *Garcinia discoidea* Nazre. **A.** habit (female); **B.** male flower; **C, D.** fruits and stigma. [(A, C: *Shamsul SA02-10*, E); (B: *Corner s.n.*, SING); (D: *Kadim KN307*, L)]. Drawn by Anna Dorward.

**6. *Garcinia exiguus* Nazre sp. nov. – Figure 6.8; Map 6.6.**

— Type: Paul Chai S31750 (holo SAR; iso L) Bintulu, Sarawak.

(Etymology: Lat. from small size in all parts of the specimens)

*Tree* to 20 m tall. *Bole*: bark dark brown, flaking, smooth and dippled, inner bark reddish. *Exudate* yellow. *Twigs* angled, stout, longitudinally finely wrinkled when dry, dark brown to pale brownish green, sometimes with very a small leaf-like, terminal bud. *Petiole* finely striate horizontally, dark brown or pale brown, to 0.5 cm long. *Lamina* thickly leathery, pale brown or greenish brown, shiny above, elliptic, 8.1-20.6 x 2.7-9.3 mm; apex acute or bluntly acuminate; base acute to cuneate; midrib raised above and below; margin entire and finely revolute; secondary veins very faint to nearly invisible above, fine and inconspicuous below, brochidodromous; tertiary veins invisible. *Glands* clearly visible below, fine and slightly raised above and raised below, arranged in dark grey-black to greenish interrupted wavy lines, running nearly parallel to the midrib to the margin. *Inflorescences* probably terminal. *Male* flower not seen. *Female* flower not seen. *Fruit* globose or ellipsoid, shiny green turning yellow, wall smooth with finely longitudinal striae, dark maroon to brown, small 9.5-10.5 x 8.5-11.0 mm; stigma sessile or slightly raised, flattened, discoid or convex, weakly 4-lobed or crenate, black rugose, to 4.5 mm across; with four persistent leathery sepals; seeds not known.

Distribution – Borneo: Sarawak and Brunei. Rare species, only represented by two collections.

Habitat & Ecology – Hill forest.

Notes – The fruits and leaves closely resemble those of *G. nitida* Pierre except that they are smaller.

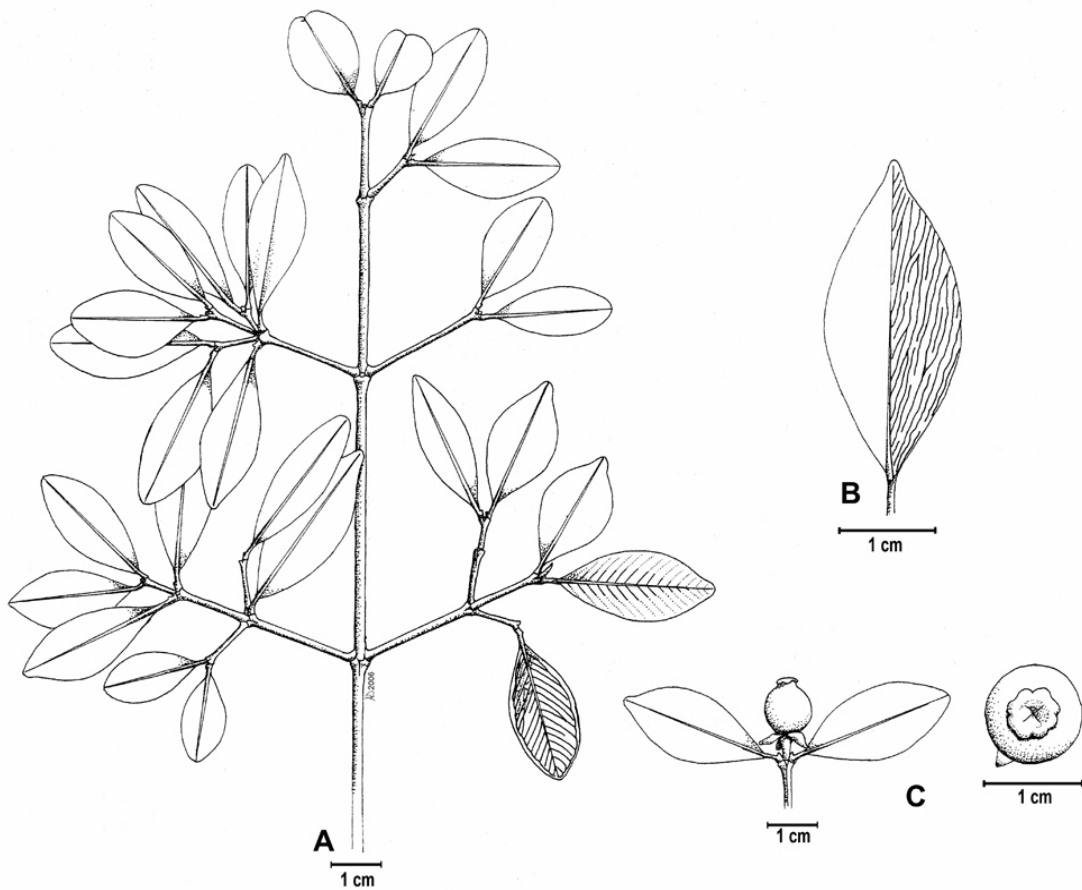
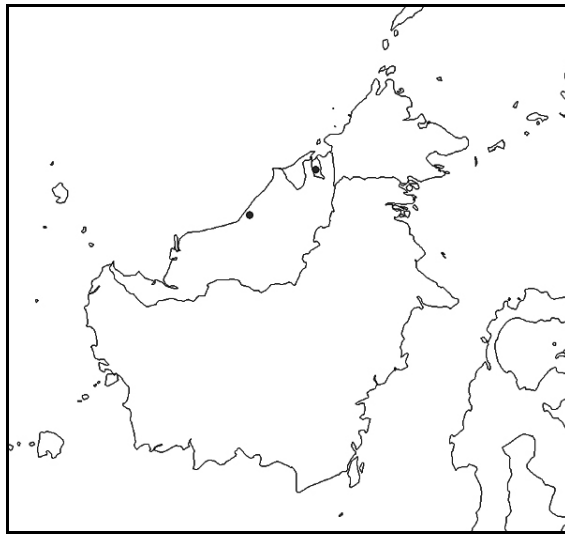


Figure 6.8. *Garcinia exiguus* Nazre. A. habit (female); B. glandular lines; C: fruit and stigma [Chai S31750, SAR]. Drawn by Anna Dorward.



Map 6.6. Distribution of *Garcinia exiguus* Nazre

**7. *Garcinia harmandii* Pierre – Figure 6.9; Map 6.7.**

*Garcinia harmandii* Pierre, Fl. Forest. Cochinch. 1 (1882) 10, pl. 60. — Lectotype (proposed here): *Harmand 349* (P), Laos.

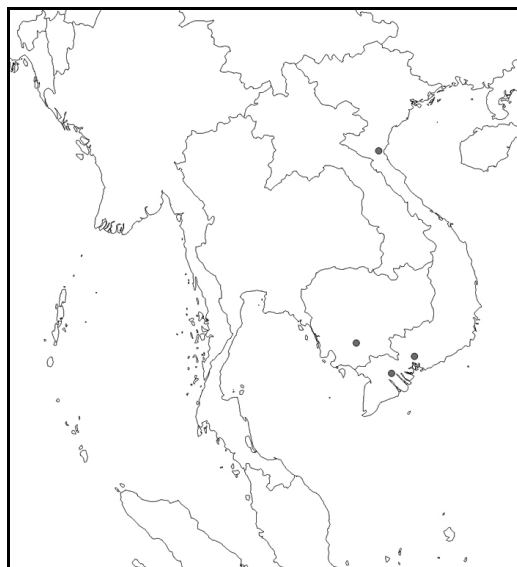
*Small bushy tree, 6-10 m tall. Bole:* no information. *Exudate* yellow. *Twigs* round and slightly angled, longitudinally wrinkled when dry, pale grey-brown to yellowish or blackish. *Petiole* dark brown or blackish, very finely horizontally striate and slightly keeled, stout and short, 0.2-0.9 cm long. *Lamina* leathery, grey-brown or dark brown, spatulate or obovate occasionally elliptic, 3.5-9.2 x 1.3-3.6 cm; apex sharply acuminate to mucronate; base cuneate to acute; margin entire and finely revolute; midrib flattened or slightly sunken above and raised below; secondary veins visible and slightly raised on both surfaces, cladodromous with intra-marginal veins; tertiary veins inconspicuous and fine, a mixture of weakly and transversely ramified and reticulate. *Glands* very fine, inconspicuous to nearly invisible below, invisible above, short wavy lines running across

secondary veins. *Inflorescences* terminal; 2-6 flowers in a cluster, usually triads in males; sepals and petals 4; bracts triangular to 2 mm long. *Male flowers* [in bud] 3 to 6; nearly sessile or on a very short pedicel; sepals concave, orbiculate or obovate, coriaceous, to 4 mm long; petals elliptic to oblong to 8 mm long; stamens 4-lobed, opposite petals; pistillodes fungiform. *Female flowers* solitary or in a triad, pedicel short to 3 mm long; bracteole present; sepals concave, orbiculate to 3.5 mm long; petals ovate, to 4 mm long; ovary sub-globose tipped with a black, rugose stigma. *Fruits* small, globose or ovoid 7-10 x 6-11 mm, turning purplish when mature, wall rough and wrinkled when dry; tipped with sessile, black rugose, discoid or skull-cap-like stigma; seeds with a white, sweet aril.

Distribution – Cambodia, Laos, Vietnam.

Habitat & Ecology – Lowland and hill forest.

Notes – Little information is available for this species which is poorly represented in herbaria. The species is characterised by a spatulate leaf with a short pointed apex.



Map 6.7. Distribution of *Garcinia harmandii* Pierre.

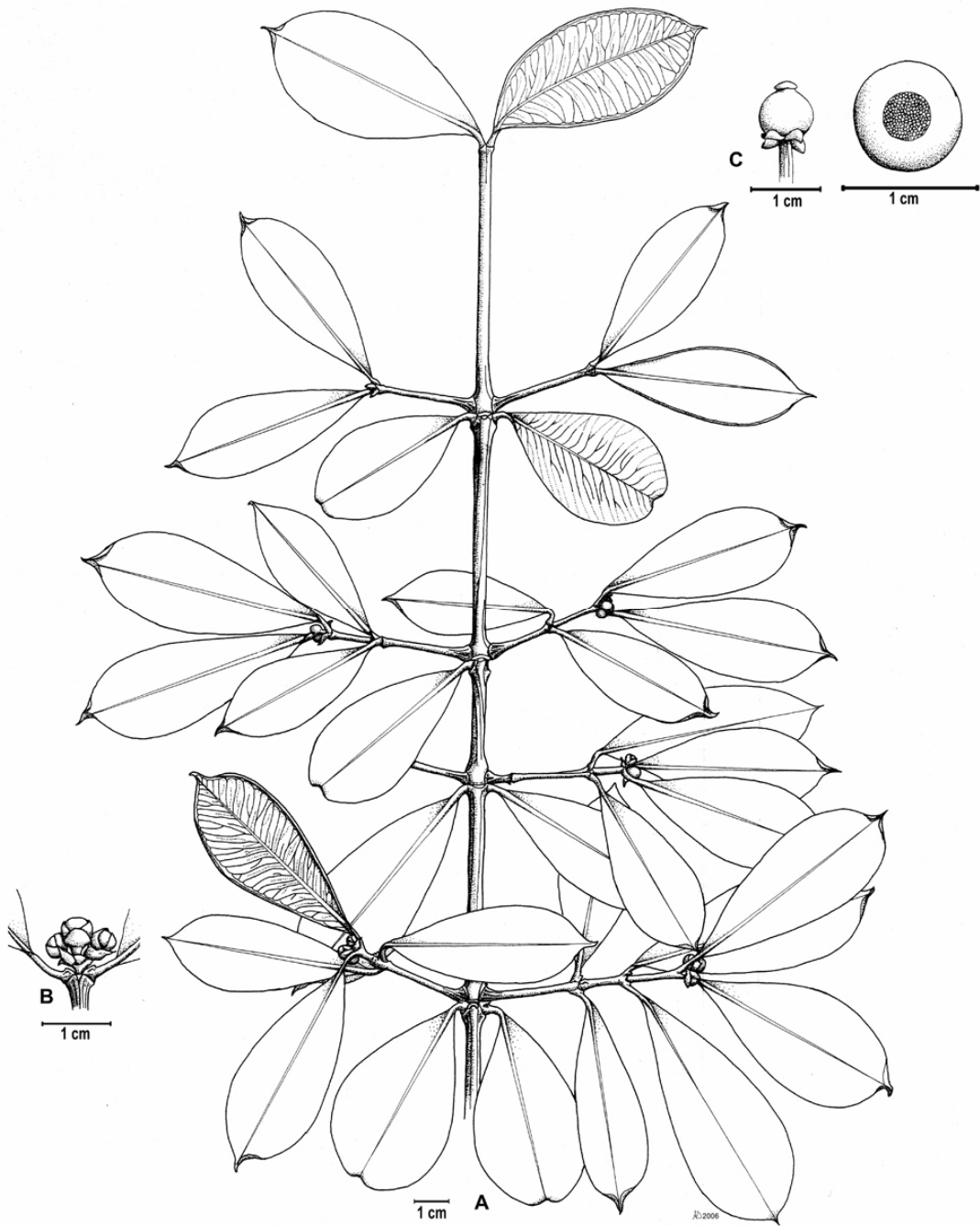


Figure 6.9. *Garcinia harmandii* Pierre. **A:** habit (male); **B:** inflorescence; **C:** fruit and stigma [(A, B: *Pierre 1371*, K); (C: *Thorel 1018*, K)]. Drawn by Anna Dorward.

**8. *Garcinia malaccensis* Hook f. – Map 6.8.**

*Garcinia malaccensis* Hook f., Fl. Brit. India 1 (1872) 261. — Lectotype (proposed here):  
*Maingay 149*[male specimen] (lecto KEP; iso K) Peninsular Malaysia.

*Tree* to 30 m tall. *Bole*: bark dark brownish to pale cream brown, slightly reddish, smooth and minutely dimpled, peeling off with flaking scales; inner bark reddish brown to pink, fibrous or yellowish; sapwood yellowish. *Exudate* yellow. *Twigs* angled, stout, roughly wrinkled longitudinally, sometimes very finely horizontally striate when dry, deep red-brown or pale brown. *Petiole* stout, finely horizontally striate, 0.4-3.0 cm long. *Lamina* thickly leathery, pale brown above and brown or sometimes reddish-brown below, shiny on the upper surface, variable in shape, mainly elliptic to narrowly elliptic, broadly elliptic, ovate or oblanceolate, 9.2-25.5 x 7.0-9.0 cm; apex sharp or bluntly acute to acuminate, sometimes slightly attenuate; base acute to cuneate or obtuse; midrib raised above and strongly raised below; secondary veins fine, visible on both sides, weakly raised above, raised below, fine and closely arranged, brochidodromous with two intra-marginal veins (mainly in female); tertiary veins fine, visible below, randomly reticulate. *Glands* fine but visible below (not in older specimens) and not visible above, dark to grey black wavy lines, closely arranged, running across the secondary veins from the midrib toward the margin. *Inflorescences* terminal in a cluster of 3-5 male flowers and solitary in the female, to 3.3 cm long; sepals and petals 4; bracts thickly leathery, triangular. *Male flowers*: pedicel stout and angular, to 3.3 cm long; sepals yellowish, thickly leathery, concave, orbiculate, broadly elliptic or ovate, 5-8 x 9-13 mm; petals pinkish, elliptic to broad elliptic, 9-15 x 5-9 mm; stamens in a single mass, 4-angled or nearly conical to

cylindrical, surrounding the base of the pistillode, to 20 mm long; pistillode very small, to 1 mm long, placed on top of stamen bundles, nearly sessile, fungiform with a small, black, rough, crenate, convex cap, or sometimes pistillode absent. *Female flowers:* pedicel stout, up to 20 mm long; sepals thickly leathery, yellowish or whitish, orbiculate or broadly ovate, 13-17 x 20-25 mm; petals smaller in size, orbiculate, widely obovate, 6-8 x 9-12 mm; ovary surrounded by irregular free staminodes attached at the base; stigma segregated or fused and sunken at the middle, (5-)6-8 closely arranged bundles, the number of bundles usually equalling the number of ovules. *Fruits* green turning yellow or reddish, big, ovoid or pear-shaped, ellipsoid or sometimes globose, 3.0-5.0 x 3.6-7.0 cm, tipped with the 6-8 segregated or fused, rugose, hardened, dark brown or blackish, nearly sessile or sessile stigmas, which is square or nearly wedge-shaped with an irregular blunt dentate margin; sepals persistent, thickly leathery, concave, orbicular or ovate; seeds with a white aril.

Distribution – Peninsular Malaysia to Singapore, Sumatra and Borneo.

Habitat & Ecology – Lowland to hill forest.

Notes – A possible parent species of *G. mangostana* with which it is easily mistaken, especially when sterile. Sometimes also confused with *G. penangiana*; many specimens from Peninsular Malaysia (e.g. in Tree Flora of Pasoh, Kochummen, 1995) have been incorrectly identified as this species. This species is characterised by having two intra-marginal veins (only in female) leaf as in *G. mangostana*, but the male flowers have a distinct 4-angled stamen with a very small pistillode or none at all. The varieties can only be distinguished in fertile and mature material.

*Key to varieties*

1. a. Stamens 4-angled, nearly conical, more than 6 mm long; fruit ovoid or ellipsoid, stigma raised.....**a.** var. *malaccensis*
- b. Stamens 4-angled, square shaped, less than 6 mm long; fruit globose, stigma sessile.....**b.** var. *pseudomangostana*

**a.** var. **malaccensis** – Figure 6.10.

*Male flower:* stamens in 4-angled, conical mass, with or without pistillode, long and slender, to 10 mm x 4 mm. *Fruits* green turning reddish or maroon, ovoid or ellipsoid; stigma large with neck, to 17 mm across, segregated bundles, fused and sunken in the middle or separate bundles usually when mature, square to nearly wedge-shaped.

Distribution –Peninsular Malaysia, Sumatra and Borneo.

**b.** var. **pseudomangostana** var. nov. Nazre – Figure 6.11.

Type: Madani 50575 (holo SAN; iso L) Sandakan, Sabah.

(Etymology: Lat. resembling mangosteen)

*Male flower:* stamens 4-angled or 4 obscure bundles with or without pistillode, short but wide to 6 mm x 8 mm. *Fruits* green turning yellowish; globose; stigma small without neck, to 5 mm across, bundles fused and weakly sunken in the middle.

Distribution –Borneo (East Coast of Sabah and Kalimantan).

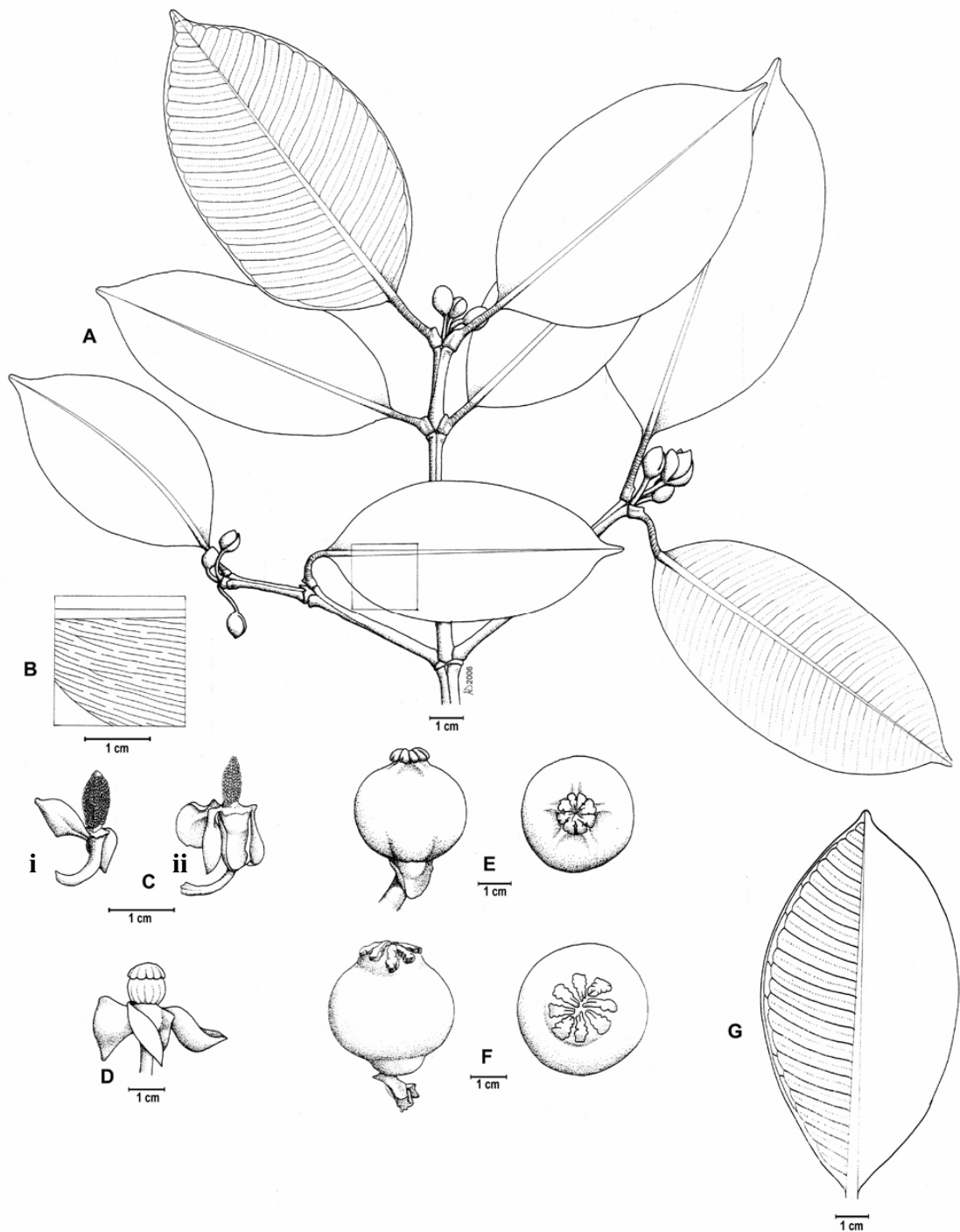


Figure 6.10. *Garcinia malaccensis* var. *malaccensis*. **A.** habit (male); **B.** glandular lines; **C.** male flowers; **D.** female flower; **E., F.** fruit and stigma; **G.** secondary nervation with two intra-marginal veins [(A, B, Cii: Maingay 149, K); (C: Ashton BRUN3392, SAR); (D: Maingay 149/Barcode 489504, L); (E: Wyatt-Smith 80555, L); (F, G: Whitmore FRI8986, L)]. Drawn by Anna Dorward.

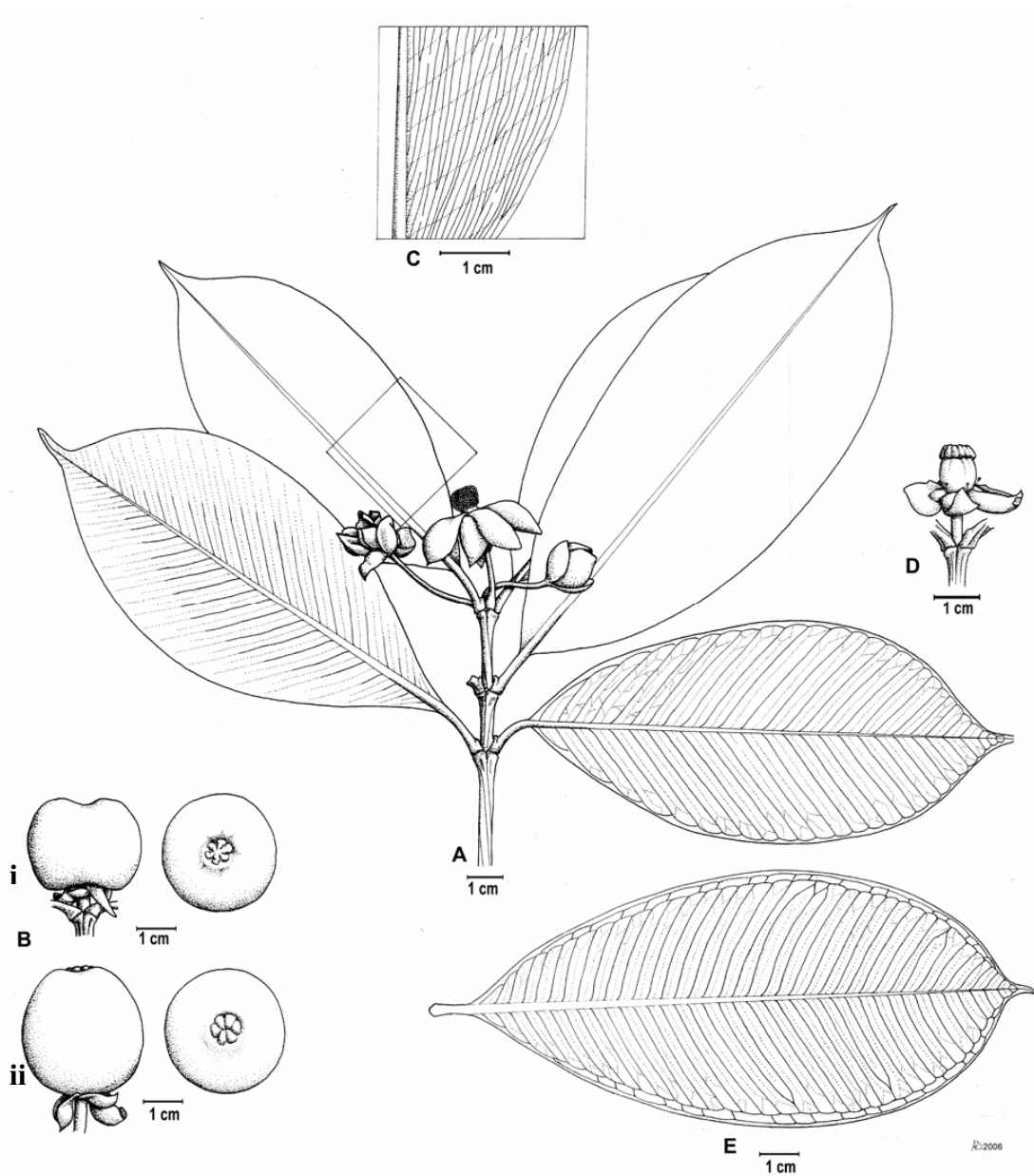
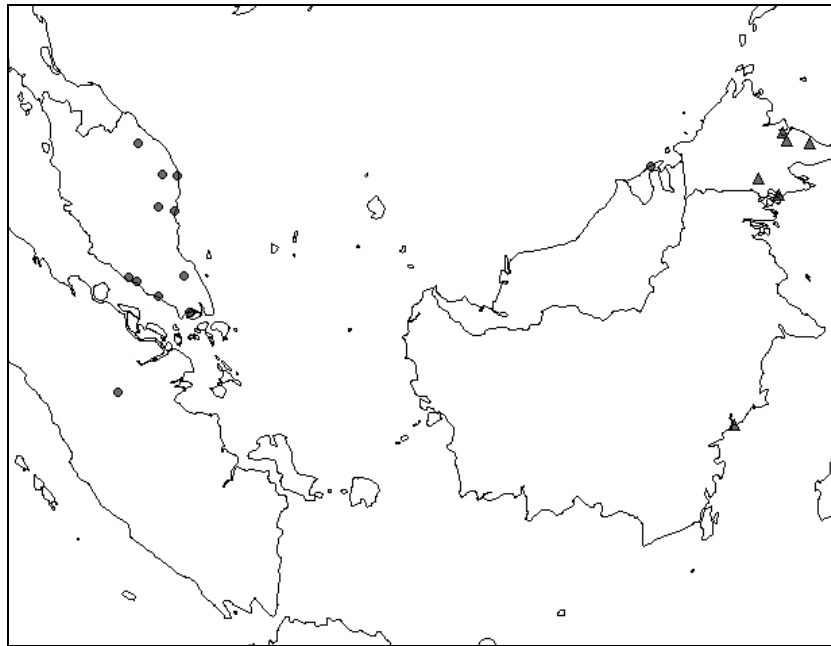


Figure 6.11. *Garcinia malaccensis* var. *pseudomangostana* Nazre. **A.** habit (male); **B.** fruit and stigma; **C.** glandular lines; **D.** female flower; **E.** secondary nervation with two intra-marginal veins [(A, C: *Madani* 50574, L); (Bi: *Meijer* SAN 19236, L); (Bii: *Fidilis & Sumbing* SAN 96472, L); (D, E: *Sigin & Joseph* SAN 97194, L)]. Drawn by Anna Dorward.



Map 6.8. Distribution of *Garcinia malaccensis* Hook. f. *G. malaccensis* var. *malaccensis* (circle); *G. malaccensis* var. *pseudomangostana* (triangle).

### 9. *Garcinia mangostana* L. – Figure 6.12.

*Garcinia mangostana* L., Sp. Pl. 443 (1753); Blume, Bijdr. Fl. Ned. Ind. (1825) 615; Miquel, Fl. Ned. Ind. 1 (1859) 506; Pierre, Fl. Forest. Cochinch. 1 (1882) 11, pl. 54; Boerlage, Cat. Pl. Phan. 2 (1901) 68; Backer & Bakhuizen, Fl. Java (Spermatoph.), (1963) 387. — Lectotype: 'mangoustan' in Garcin, *Philos. Trans.*, 38 (1734) 1. Designated by Hammel in Jarvis et al., *Regnum Veg.* 127 (1993) 28.

*Tree* to 20 m tall. *Bole* brown to dark brown, smooth, scaly and flaking; inner bark orange or red brown to light brown. *Exudate* yellow. *Twigs* clearly 4-angled, stout, roughly wrinkled longitudinally when dry, deep dark yellowish-brown or pale brown. *Petiole* stout, finely horizontally striate, angled, to 3 cm long. *Lamina* thickly leathery, sometimes shiny on the upper surface, pale brown above and reddish-brown below when

dry, elliptic to broadly elliptic, ovate or oblanceolate, 10.0-25.0 x 7.0-9.0 cm; apex acute to obtuse; base cuneate or obtuse; midrib raised above and strongly raised below; secondary veins fine but clearly visible, raised below, brochidodromous with two intramarginal veins; tertiary veins a mixture of reticulate and scalariform, fine and prominent. *Glands* fine, conspicuous below, invisible above, dark to grey black wavy lines, running across the secondary veins, c. 70° from the midrib to the margin. *Inflorescences* terminal in a cluster of 3-4(5) male flowers and usually solitary in the female but sometimes with 2-3 flowers, male flowers smaller than female; sepals and petals 4. *Male flowers* to 3.0 cm across when open; pedicel round or slightly angled, stout, to 15 mm long; petals and sepals leathery, unequal in size, usually inner parts larger than outer, concave to orbicular, 15-20 mm across; stamens square (4-angled) to slightly cylindrical surrounding the base of pistillode, to 8 mm tall; pistillode broadly fungiform in shape, tipped with a convex or mushroom-like cap, margin lobed and crenate, to 5 mm across; anthers 2-thecous with apical slits, nearly sessile or with a short filament. *Female flower* larger than male to 5.0 cm across when open; pedicel as in the male; sepals and petals as in the male but larger to 20 mm across; ovary surrounded by a few to 20 irregular, free staminodes; ovary dwarf fungiform, tipped with a skull-cap stigma, 4-5 lobed. *Fruits*: large, ovoid or globose, to 6 cm across, tipped with the black, sessile stigma, 4-8 lobed or wedge-shaped bundles, fused in the middle, usually the number of lobes or bundles equalling the number of ovules; seeds with white, gelatinous, edible aril.

Distribution – Cultivated unknown origin.

Habitat & Ecology – Planted throughout tropical countries, especially in Southeast Asia.

Notes – Very similar in its vegetative characters to *G. malaccensis* but differing in having thicker and more prominent midribs and venation. Male trees are rarely found and are only recorded from two localities in Peninsular Malaysia. The male flower was correctly described by Salma & Rukayah (1988). Other descriptions of the male flowers by Roxburgh (1832), King (1891) and Maheshwari (1964) fit the description of *G. celebica* L.

Molecular data suggest that the origin of the species might be from multiple hybridisations between *Garcinia malaccensis* varieties either from Peninsular or East Malaysia (Borneo) if mangosteen is an obligate agamosperm species (Chapter 5). If mangosteen is however, a facultative agamosperm a number of *Garcinia* species could be involved in its parentage, besides *G. malaccensis*.

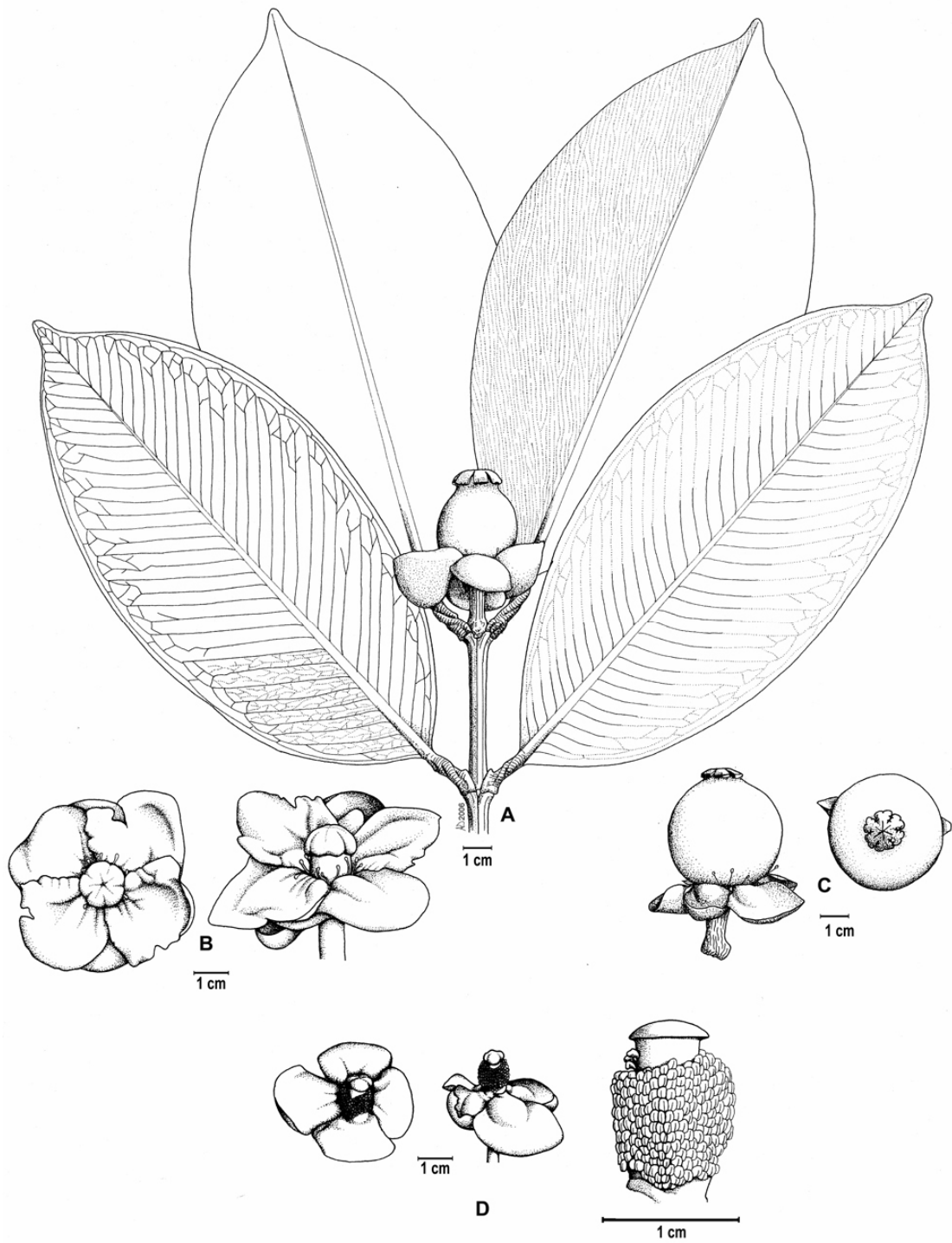


Figure 6.12. *Garcinia mangostana* L. **A:** habit (female); **B:** female flower; **C:** young fruit; **D:** male flower [(A, C: Sibat S23041, L.); (B: Anon, MARDI); (D: Shamsudin s.n., MARDI)]. Drawn by Anna Dorward.

**10. *Garcinia moselleyana* Pierre** – Figure 6.13; Map 6.9.

*Garcinia moselleyana* Pierre, Fl. Forest. Cochinch. 1 (1883) 10. — Lectotype (proposed here):

*Moseley s.n.* (K) Basilan Island, Philippines.

*Small tree* to 3 m tall. *Bole*: no information. *Exudate* yellow. *Twigs* round, longitudinally wrinkled when dry, pale grey-brown to dark-brown. *Petiole* striate longitudinally, same colour as twigs, to 0.6-0.9 cm long. *Lamina* leathery, elliptic or oblong 10.0-15.0 x 3.5-6.0 cm, dark or pale brown; apex acuminate or attenuate; base acute or cuneate; margin weakly wavy or entire and revolute; midrib flattened and inconspicuous to weakly sunken above and strongly raised below; secondary veins clearly visible on both sides, conspicuous and raised below, cladodromous, with fine intramarginal veins; tertiary veins visible below, randomly reticulate. *Glands* not found. *Inflorescences* terminal and axillary, 3-9 flowered, in clusters of variable size and sometimes solitary in females; sepals and petals 5; bracts triangular, to 4.5 mm long. *Male flower* [in bud] nearly sessile or with a short pedicel up to 3 mm long; sepals leathery, concave, broadly ovate or broadly elliptic, two outer, three inner, 4-7 mm long; petals concave, ovate, smaller than the sepals; stamens slightly 4-lobed, enclosing the fungiform pistillode at the centre; pistillode small; anthers 2-theous. *Female flowers*: sepals and petals as in the male; staminodes lacking; ovary globose, tipped by a small, to 3 mm across, skull-cap shaped, black, peltate stigma. *Fruits* smooth, globose 20-33 x 25-35 mm, black when dry; stigma as in the female flowers; no information on seeds.

Distribution – Philippines, endemic.

Habitat & Ecology – Lowland and hill forest near rivers.

Notes – The only species with pentamerous flowers in the section. A distinct species easily identified by the cladodromous nervation and flattened midrib on the adaxial leaf surface.

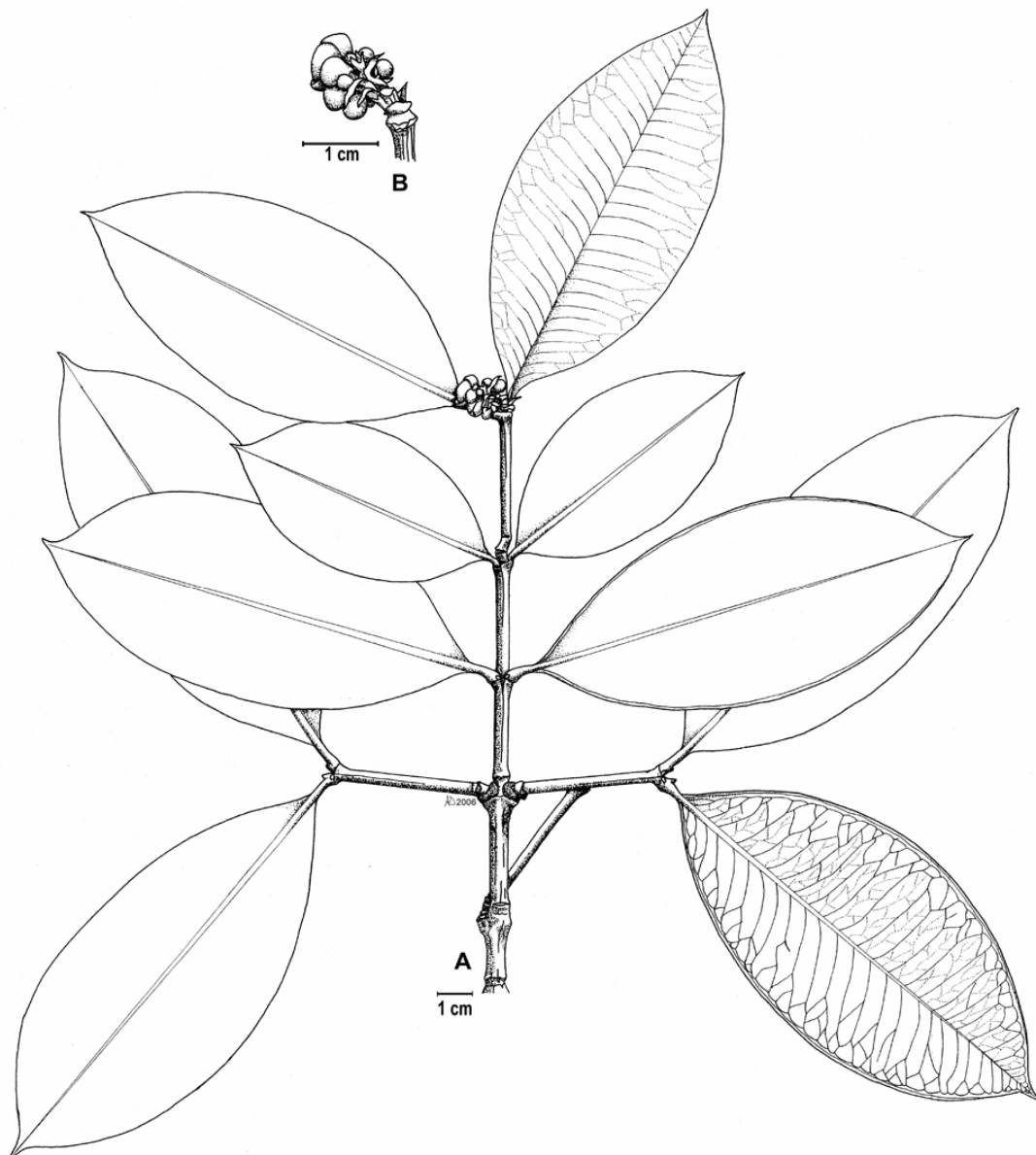
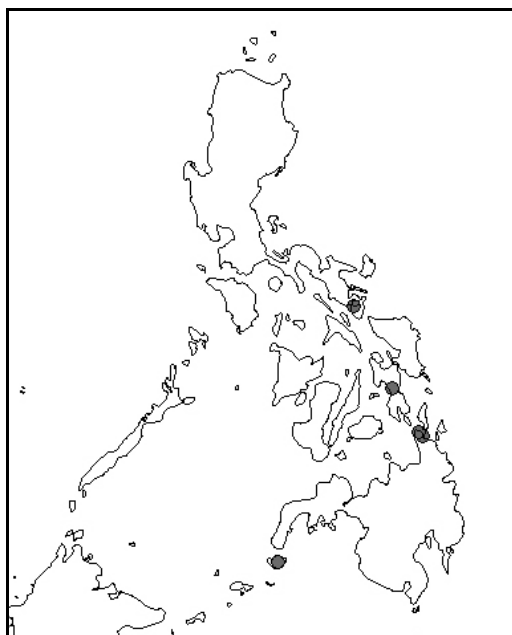


Figure 6.13. *Garcinia moselleyana* Pierre. **A.** habit (male); **B:** inflorescence [*Moseley s.n.*, K]. Drawn by Anna Dorward.



Map 6.9. Distribution of *Garcinia moselleyana* Pierre.

**11. *Garcinia nitida* Pierre** – Figure 6.14; Map 6.10.

*Garcinia nitida* Pierre, Fl. Forest. Cochinch. 1 (1882) 18 & Fl. Forest. Cochinch. 1 (1883) pl.80M & 80N. — Lectotype (proposed here): *Beccari 2104* (lecto K; iso A) Sarawak.

*Tree* to 30 m tall and 45 cm dbh. *Bole*: bark brown, inner bark reddish. *Exudate*: cream or yellow. *Twigs* slightly angled, stout, longitudinally wrinkled when dry, dark pale brown to pale yellowish green. *Petiole* finely striate horizontally, stout, same colour as twigs, 0.3-1.1 cm long. *Lamina* leathery, pale brown to reddish-brown, darker and shiny above, pale below, broadly elliptic, elliptic or sub-orbiculate, 5.0-8.5 x 2.9-3.8 cm; apex to 10 mm long, acute, blunt or blunt-acuminate; base acute to nearly obtuse; midrib raised to nearly square in cross section above, raised below but gradually flattening to the apex; margin entire and finely revolute; secondary veins very fine, sometimes nearly invisible,

inconspicuous above, slightly visible below, brochidodromous, very closely arranged; tertiary veins invisible. *Glands* fine but clearly visible below, especially in younger leaves, black to grey-black or dark greyish brown, of interrupted wavy lines, with a mixture of short and long lines and dots, running c. 70° to the midrib across the secondary veins from midrib towards margins. *Inflorescences* terminal, 1-4 flowers on male, solitary, or rarely, in pairs on female; sepals and petals 4; bracts triangular, to 5 mm long. *Male flower* [in bud]: pedicel to 5 mm long; sepals concave, to 7 mm long; petals narrowly obovate, to 7 mm long; stamens in 4 weak lobes, attached at 1/3 from the base of pistillode, surrounding the fungiform pistillode; anthers 2-theous. *Female flower* not seen. *Fruit* ovoid or flask-shaped to globose, ripening red, pale to dark brown or reddish brown to dark maroon when dry, smooth, thin wall easily ruptured when dry; stigma raised or elongated to 10 mm long, concave with irregular wavy margin to 6 mm across; seeds 4-5 with a white aril.

Distribution – Borneo.

Habitat & Ecology – From lowland to hill forest.

Notes – Closely related to *Garcinia diospyrifolia* and resembling *G. diospyrifolia* var. *minor* but is characterised by being a big tree, the leaves having a mixture of long and short wavy glandular lines, and the fruit with distinct elongated and sunken stigma.

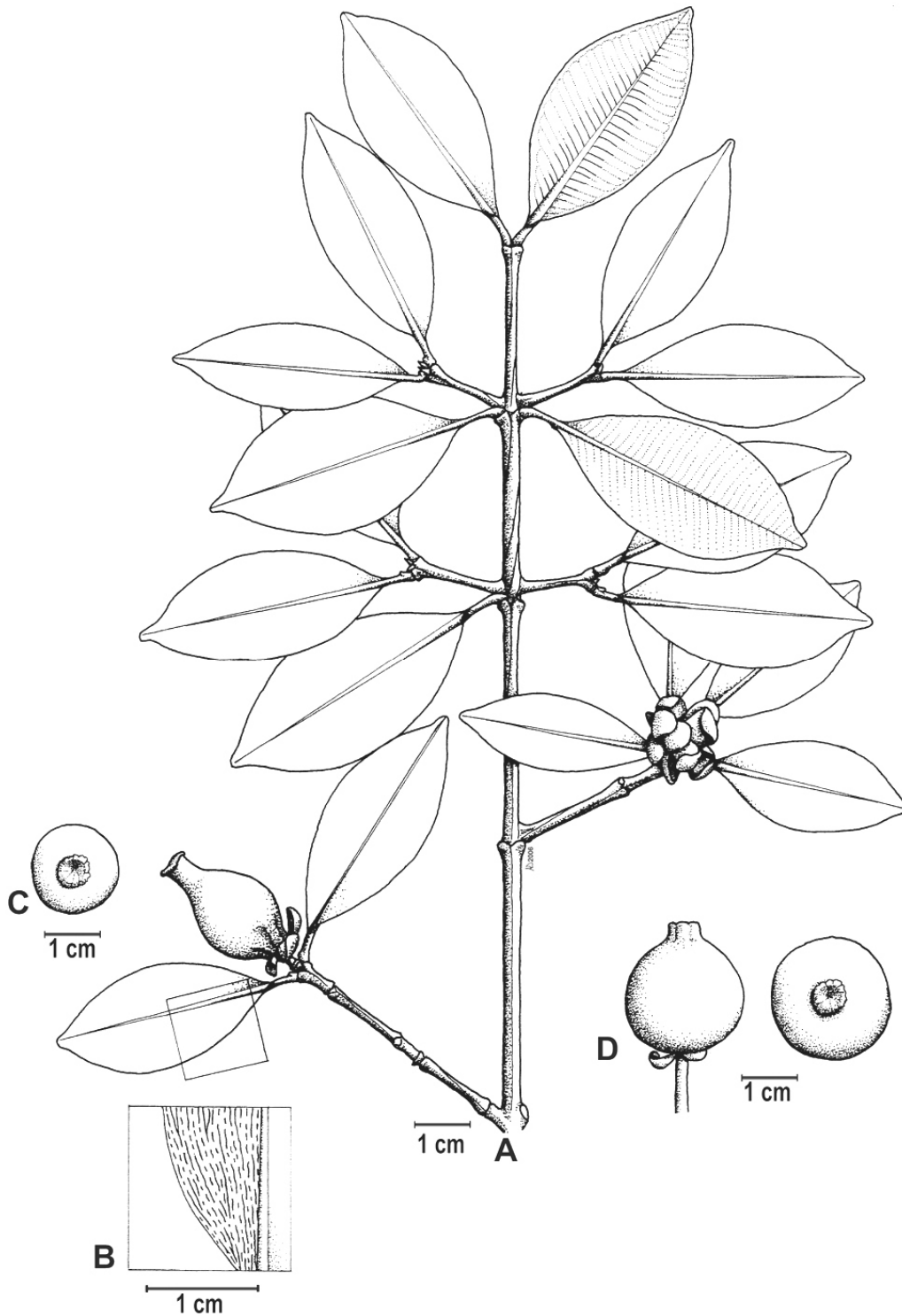


Figure 6.14. *Garcinia nitida* Pierre. **A.** habit (female); **B.** glandular lines; **C.** stigma; **D:** fruit and stigma [(A, C, B: *Argent & Amiril 9318*, K); (D: *Ilias 13317*, K)]. Drawn by Anna Dorward.



Map 6.10. Distribution of *Garcinia nitida* Pierre.

**12. *Garcinia ochraceus* Nazre sp. nov. – Figure 6.15; Map 6.11.**

— Type: *Streimann & Martin LAE 52886* (holo E) West Sepik, Papua New Guinea.

(Etymology: Lat. from the colour of the stamens)

*Small tree* to 6 m tall, slender. *Bole*: no information. *Exudate*: no information. *Twigs* slender, round to angled, longitudinally striate when dry; reddish-brown or brownish yellow. *Petiole* finely striate horizontally, darker colour than twigs, 0.5-0.75 cm long. *Lamina* thinly leathery, pale grey-brown or brownish to yellowish, darker and shiny above, slightly glaucous or opaque yellowish below, elliptic, 6.0-9.5 x 2.1-5.3 cm; apex acute and blunt, sometimes blunt-acuminate; base acute to cuneate; midrib yellowish, raised above, flattened or weakly raised below; margin entire and finely revolute; secondary veins inconspicuous on both sides, fine, brochidodromous, closely arranged; tertiary veins invisible. *Glands* faint to invisible, fine, closely arranged in dark grey-black,

interrupted, wavy lines, running across the secondary veins, nearly parallel to midrib.

*Inflorescences* terminal, 1-2 flowered, female flowers always solitary; bracts not found.

*Male flower* with 4 sepals and petals, leathery, outer usually thicker; pedicel 6 mm long;

sepals ovate or orbiculate to 6 mm across; petals red turning maroon when dry,

oblanceolate or obovate to 10 mm long; stamens yellow, in a mass, weakly 4-lobed

central bundle surrounding the pistillode; pistillode fungiform; anthers 2-theous with a

short filament. *Female* flower not seen. *Fruit* globose, cream turning red, reddish or

maroon brown to brownish when dry, wall smooth, 1.8-2.3 x 1.8-2.1 cm, tipped with a

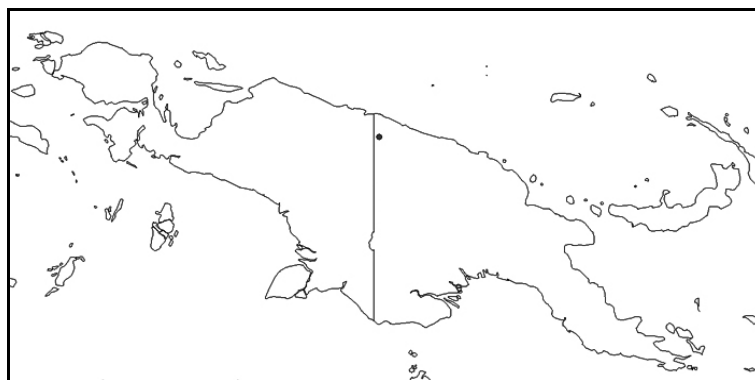
black stigma, sessile, flattened on the surface but slightly sunken in the middle, margin

weakly crenate, to 7 mm across; with thick leathery prominent sepals; unknown seed.

Distribution – New Guinea.

Habitat & Ecology – Lowland rainforest.

Notes – The fruits resemble *G. diospyrifolia* and *G. nitida* but the stigma is rather flat and without lobes. There are also vegetative characters that can differentiate the species: the leaf with faint and nearly invisible venations on the lower surface, and the glandular lines almost invisible.



Map 6.11. Distribution of *Garcinia ochraceus* Nazre.

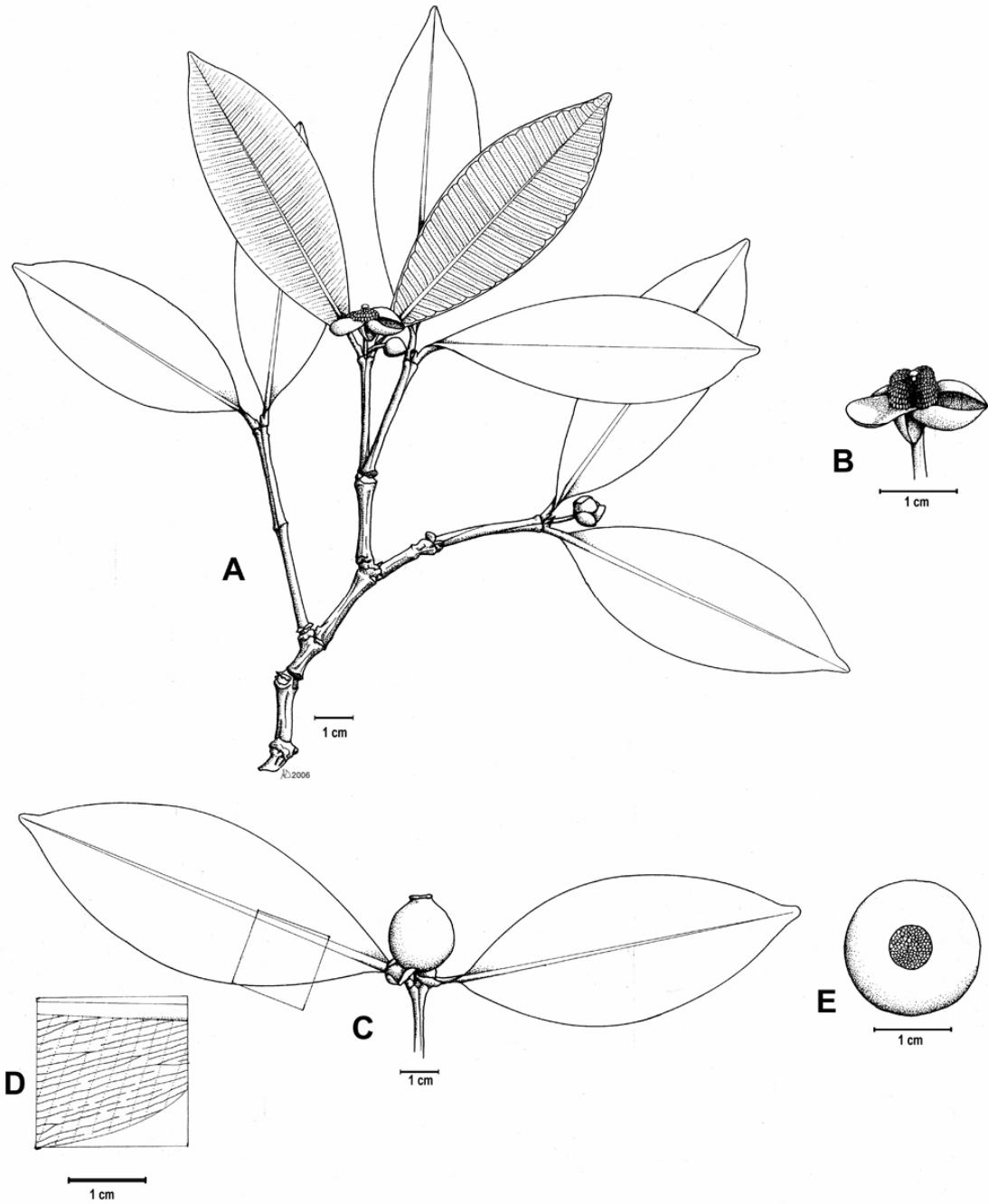


Figure 6.15. *Garcinia ochraceus* Nazre. **A.** habit (male); **B.** male flower; **C, E.** fruits and stigma; **D.** glandular lines [(A, B: Streimann & Martin 52886, E); (C, D, E: Streimann & Martin 52890, E)]. Drawn by Anna Dorward.

**13. *Garcinia penangiana* Pierre** – Figure 6.16; Map 6.12.

*Garcinia penangiana* Wall. ex Pierre, Fl. Forest. Cochinch. 1 (1882) 37. — Lectotype (proposed here): Wallich 4852D (P) Penang, Peninsular Malaysia.

*Garcinia fascicularis* Wall., Numer. List (1831) 4853, nom. nud.

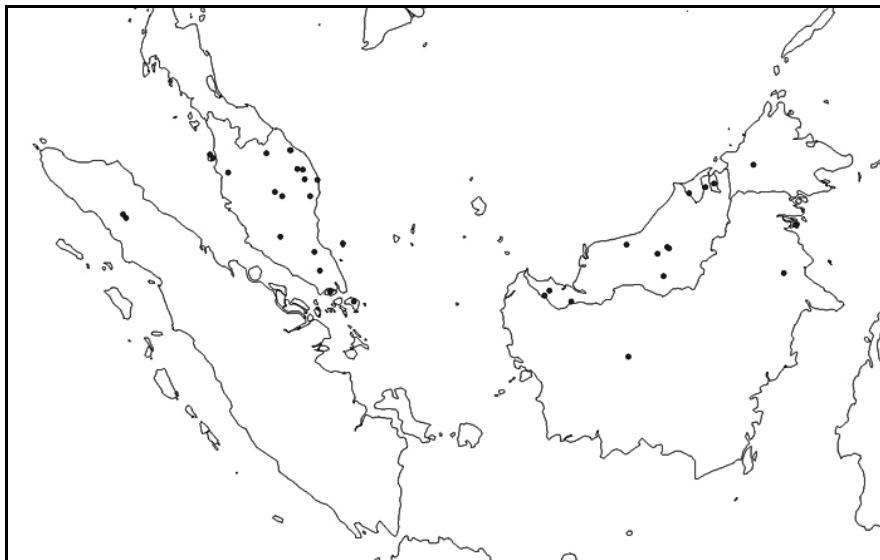
*Small to medium sized tree* to 20 m. *Bole*: bark dark brown to blackish, smooth and flaking; inner bark reddish. *Exudate* white. *Twigs* slender, angular, reddish black or orange-brown and wrinkled longitudinally when dry. *Petioles* stout, finely striate horizontally, 0.7-1.9 cm long. *Lamina* leathery, dark reddish brown above and paler below (distinctly reddish in new specimens), elliptic to narrowly elliptic, 13.2-16.0 x 3.5-6.0 cm; apex bluntly acuminate; base acute; margin entire, finely revolute; midrib slightly raised and square in cross section above, strongly raised below; secondary veins brochidodromous, fine, raised above and below, very closely arranged; tertiary veins very fine, conspicuous below, inconspicuous above, scalariform. *Glands* clearly visible and raised below, a mixture of long and short black wavy lines, running from midrib across secondary veins towards margin. *Inflorescences* terminal, in clusters of 3-10 male flowers of variable size, the females usually solitary but sometimes in pairs; sepals and petals 4. *Male flower*: pedicel slender, 5.0-13 mm long; sepals concave, ovate or orbicular, 3-7 x 4-10 mm; petals oblanceolate or elliptic, 7-15 x 3.2-7 mm; stamens in a cruciform central mass (from above) or slightly 4-angled; pistillodes absent; anthers sessile or on very short filaments, 2-theous. *Female flower*: pedicel slender or stout, to 7 mm long; sepals and petals as in male flower, deep red or light green; ovary globose, tipped by a black, corrugated stigma in 3-4 bundles. *Fruits* globose or ovoid, green, smooth, dark brown or purplish when dry, tipped with nearly sessile blackish stigma, 3-4 wedge-shaped bundles

of many nodule-like surface, segregated or fused (becoming lobed), entire or cracked margin; seeds 3-4(5) with a white aril.

Distribution – Peninsular Malaysia, Sumatra and Borneo.

Habitat & Ecology – Common species in lowland to hill forest sometimes reaching lower montane forest (900 m).

Notes – This species is characterised by the distinct leaf characters: the dry leaf turns reddish, the secondary veins are fine but visible below and closely arranged. Male flowers are particularly distinctive due to the cruciform stamens when dry. However, sterile specimens are always confused with *G. malaccensis* and *G. diospyrifolia*, especially older specimens. Morphologically (also supported by the molecular data) it is closely related to *Garcinia malaccensis* Hook. f., *G. mangostana* L. and *Garcinia venulosa* (Blanco) Choisy.



Map 6.13. Distribution of *Garcinia penangiana* Pierre.

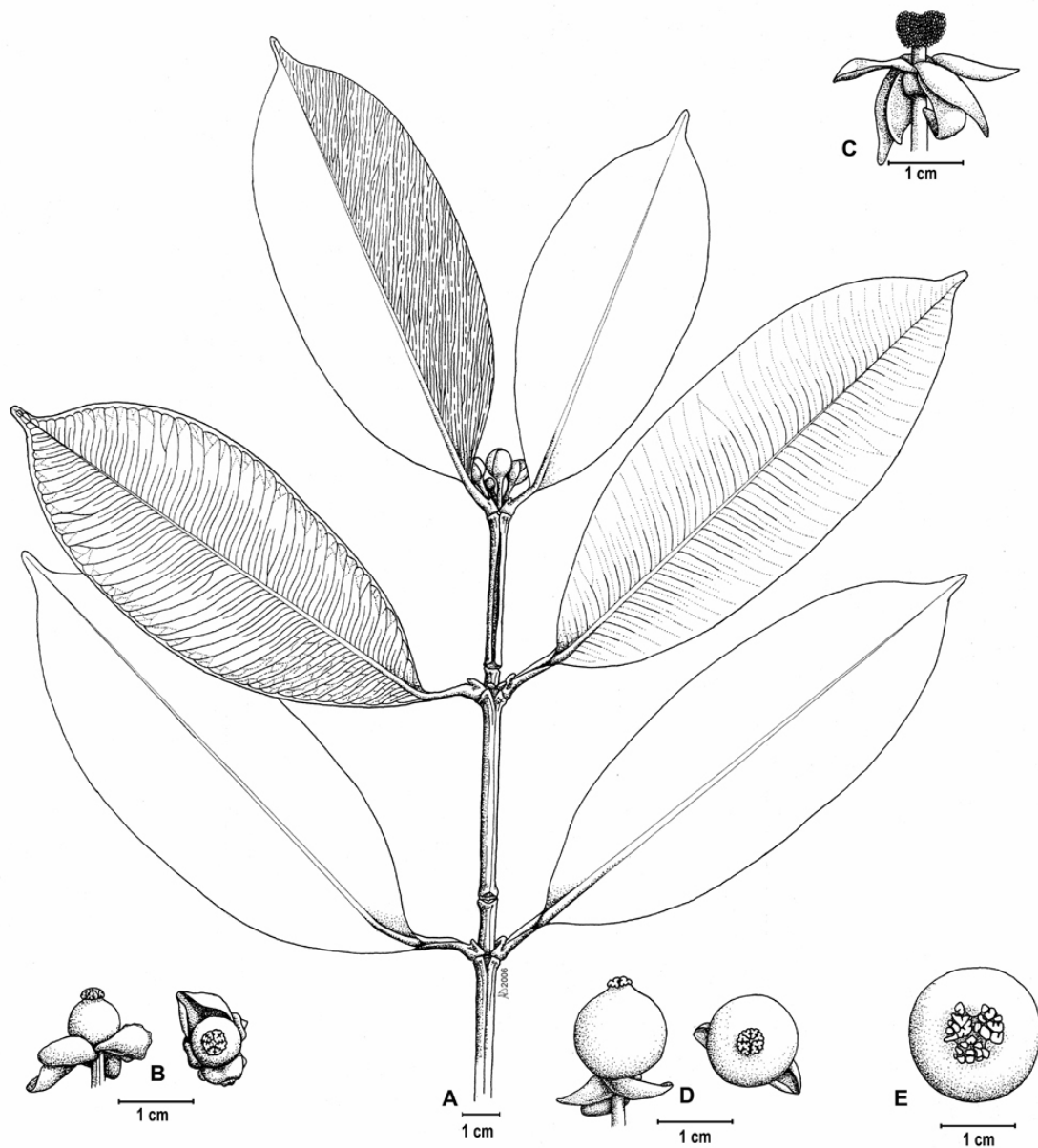


Figure 6.16. *Garcinia penangiana* Pierre. **A**: habit (male); **B**: young fruit and stigma; **C**: male flower; **D**, **E**: mature fruit and stigma [(A, C: Whitmore FRI 20347, L); (B: Sibat S25238, K); (D: Sibat S23622, K); (E: Sinclair SFN 40794, SING)]. Drawn by Anna Dorward.

**14. *Garcinia rigida* Miq. – Figure 6.17; Map 6.14.**

*Garcinia rigida* Miq., Fl. Ned. Ind., Eerste Bijv. (1861) 493. — Lectotype (proposed here):

*Diepenhorst HB646* (lecto BO; iso L) Priaman, Sumatra.

*Garcinia schefferi* Pierre, Fl. Forest. Cochinch. 1 (1882) 12, pl.59, excl. Harmand 2029. —

Lectotype (proposed here): *Harmand 4012* (lecto P; iso K) Condore Island, Vietnam.

*Garcinia porrecta* var. *schizogyna* Boerl., Cat. Hort. Bogor. 2 (1901) 69. — Nomen. invalidum.

*Tree* to 30 m tall. *Bole*: sometimes with buttresses, bark light brown or pale grey-dark brown, occasionally exposing dark cream bark under peeling scales; inner bark reddish; sapwood yellow. *Exudate* white, turning cream or yellow. *Twigs* angled, stout, longitudinally wrinkled when dry, dark brown or brown to greenish. *Petioles* stout, finely striate horizontally, 0.6-2.0 cm long. *Lamina* leathery, narrowly elliptic or ovate, elliptic to broadly elliptic, 0.5-18.0 x 3.0-8.0 cm; apex blunt acute to acuminate; base cuneate to acute; margin entire, weakly and finely revolute; midrib slightly raised above and raised below; secondary veins inconspicuous above and slightly raised below, brochidodromous, closely arranged; tertiary veins invisible below, visible above only in new specimens, randomly reticulate. *Glands* black or dark brown to greenish, fine wavy lines running across secondary veins from midrib towards margin, c. 1 mm apart, clearly visible and weakly raised below (not so in very old specimens). *Inflorescences* probably terminal. *Male flower* not seen. *Female flower* not seen. *Fruit* solitary and terminal; pedicel stout, to 2.0 cm; brown to dark brown and slightly reddish-brown when dry, ovoid or ellipsoid, 2.5-3.2 x 2.0-2.5 cm, thin walled and smooth; stigma raised, discoid but flattened on the surface or slightly sunken in the middle, margin irregularly dentate or weakly 4 lobes; 4 persistent sepals, greenish white, leathery, concave; seeds unknown.

Distribution – Indo-china, Sumatra, Sulawesi and Maluku.

Habitat & Ecology – Lowland, especially near the coast or on islands.

Notes – Specimens cultivated in Bogor Botanic Gardens and labelled as *G. porrecta* var. *schizogyna* (VI.C.145, VI.A.50 & VI.C.235) bear an invalid name and are *G. rigida*. They were collected from Sulawesi and Ambon, Maluku. However, I have not yet found any herbarium material collected from this area from wild individuals of *G. rigida*.

The species is characterised by the fruit: the wall is thin and smooth as in *G.*

*diospyrifolia* but it has a distinct discoid, flattened stigma with a neck. The leaves resemble *G. celebica* but the glandular lines are more distinct and widely spaced.

Molecular analysis clearly shows the monophyly of this species and it is quite distantly related to either *G. celebica* or *G. diospyrifolia*. Incongruence between plastid and nuclear phylogenetic trees indicate that it could have arisen from a hybridisation event with maternal origins from species outside section *Garcinia*.

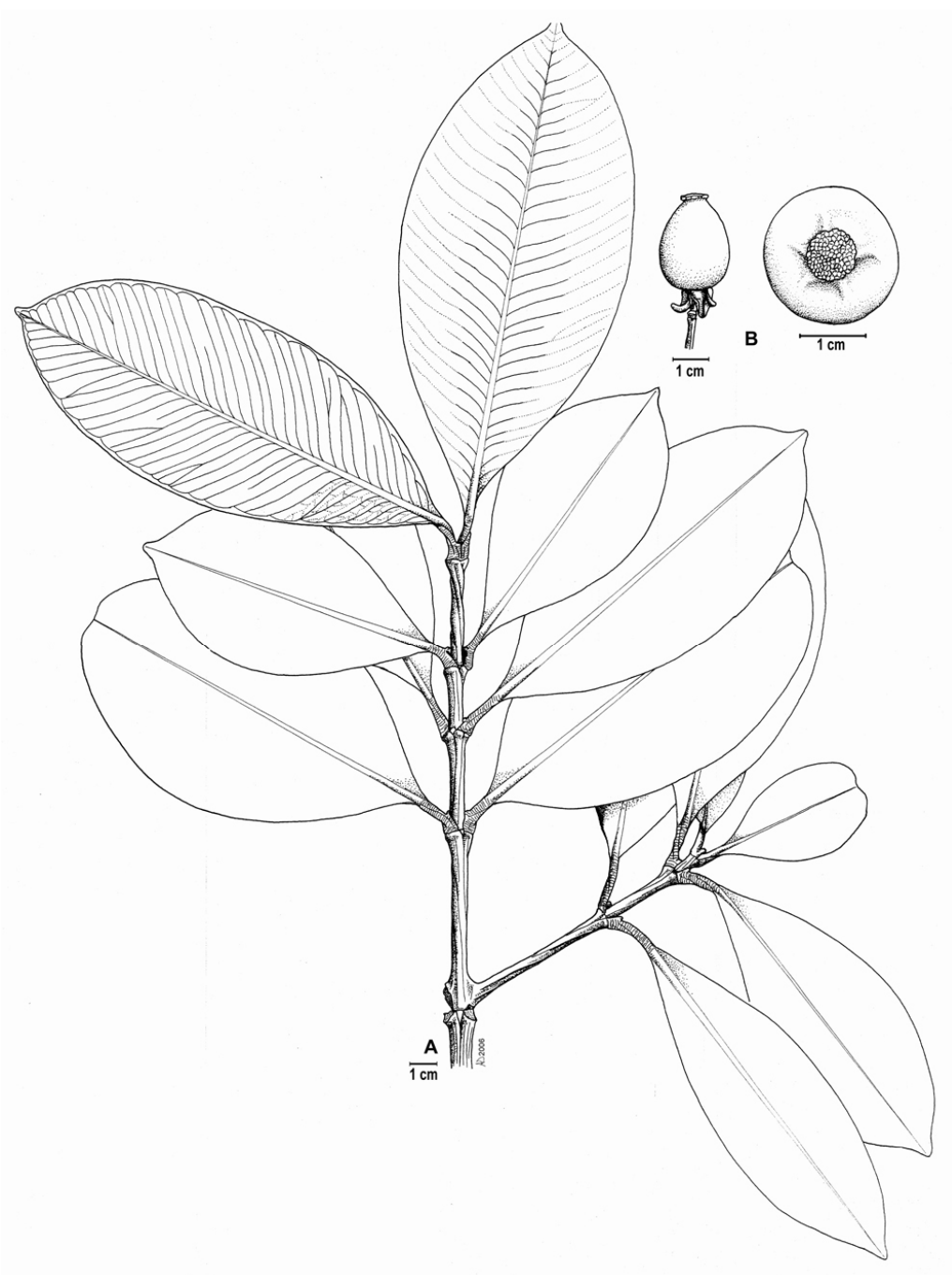
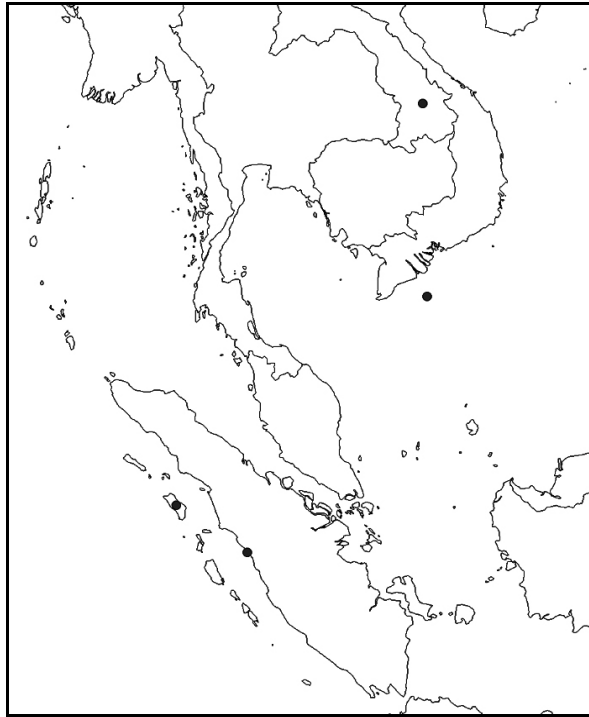


Figure 6.17. *Garcinia rigida* Miq. **A.** habit (female); **B.** fruit and stigma [*Harmand 4012, K*].  
Drawn by Anna Dorward.



Map 6.14. Distribution of *Garcinia rigida* Miq.

**15. *Garcinia sangudsangud* Nazre sp. nov. – Figure 6.18 ; Map 6.15.**

—Type: *Aban Gibot* SAN32977 (holo SAN; iso K, L) Tawau, Sabah, Malaysia  
(Etymology: local name in Sabah)

*Tree* to 20 m tall. *Bole*: bark blackish, with papery scales; inner bark reddish brown, sapwood pale red-brown. *Exudate* white. *Twigs* slightly angled especially towards end; pale greyish brown. *Petiole* stout, finely striate horizontally, 0.5-2.0 cm long. *Lamina* leathery; dark brown or pale brown and shiny above, pale greenish brown or brown below, elliptic or narrowly elliptic, 7.2-15.0 x 2.1-5.0 cm; apex with drip tips, blunt acuminate to 13 mm long; base acute; midrib raised above, nearly square in cross section, strongly raised below; margin entire and finely revolute; secondary veins visible on both

sides, fine, brochidodromous but intra-marginal veins very fine, closely arranged, inconspicuous above and slightly raised below; tertiary veins invisible. *Glands* fine but visible below, dark grey-green to blackish, mixture of interrupted wavy lines running across secondary veins towards the margin, with many fine dashes and dots in between them. *Inflorescences* terminal, males with 1-3 flowers, females solitary, pedicel short, to 5 mm long; petals and sepals 4. *Male flowers*: [in bud], sepals and petals concave; pistillode a dwarf fungiform shape with a large dome-shaped cap; surrounded by stamens in 4 weak lobes attached at the base of pistillode; anthers sessile, 2-theous. *Female flower*: sepals thick, ovate; ovary dwarf fungiform, stigma small, hemispherical, turning flat-topped when mature or slightly sunken; staminodes absent. *Fruit* ovoid or globose, 2.5 x 1.6 cm, with a thick and hardened or stony wall, smooth sometimes shiny surface, brown to dark brown; stigma sessile or weakly raised, to 3 mm long, dark brown or blackish, thinly flattened, weakly convex or sunken on surface; seeds unknown.

Distribution – Sabah, Malaysia.

Habitat & Ecology – Lowland and hill forest up to lower montane forest.

Notes –Closely resembling *G. nitida* in its male flower and leaf but distinguished by the thick and hard fruit wall.

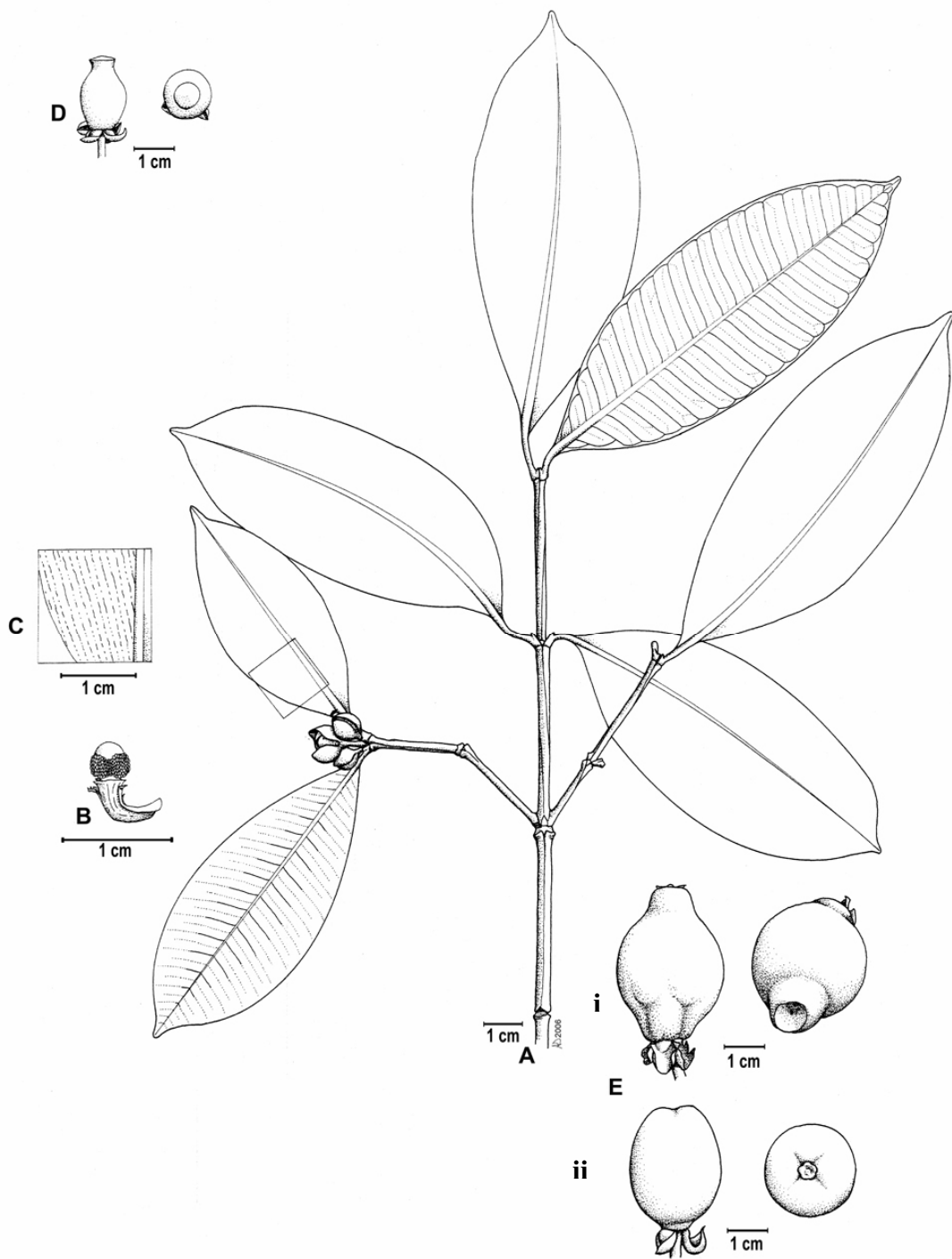


Figure 6.18. *Garcinia sangudsangud* Nazre. **A.** habit (male); **B.** male flower; **C.** glandular lines; **D.** young fruit; **E.** mature fruits and stigma [(A, B, C: *Aban* SAN 32914, L); (D: *Meijer* SAN 22942, L); (Ei: *Muin Chai* SAN 33449, L); (Eii: *Burley* 2660, E)]. Drawn by Anna Dorward.



Map 6.16. Distribution of *Garcinia sangusangud* Nazre

**16. *Garcinia venulosa* (Blanco) Choisy** – Figure 6.19; Map 6.17.

*Garcinia venulosa* (Blanco) Choisy, Mem. Soc. Hist. Phys. Nat. 12: 414 (1850). — *Cambogia venulosa* Blanco, Flora de Filipinas (1837) 435. — Neotype (proposed here): *Cuming 1124* [male specimen] (neo K; iso BM).

*Garcinia cumingiana* Pierre, Fl. Forest. Cochinch. 1 (1882) 11, Fl. Forest. Cochinch. 1 (1883) pl. 78E & 78F. — Lectotype (proposed here): *Cuming 1124* [female specimen] (lecto K; iso BM).

*Tree* to 25 m tall. *Exudate* yellow. *Bole*: no information. *Twigs* angled, stout, longitudinally striate when dry; dark brown or chocolate to pale brownish yellow. *Petiole* finely striate horizontally, stout, with ligule-like appendages at the base, brown, yellowish, dark brown or dark maroon, 0.9-2.2 cm long. *Lamina* thickly or thinly leathery, pale brown or pale grey-brown to greenish, darker and shiny above, elliptic-oblong, broadly elliptic to ovate or obovate, 8.1-20.6 x 2.7-9.3 cm; apex acute, sometimes

acuminate or attenuate; base acute to cuneate or decurrent; midrib yellowish, flattened to slightly raised above, raised and sharp below; margin entire and finely revolute; secondary veins brochidodromous, visible on both sides, closely arranged, slightly raised, occasionally with 2 intramarginal veins especially on female but these are very fine; tertiary veins inconspicuous below, fine, reticulate. *Glands* fine, closely arranged in dark grey-black to greenish interrupted wavy lines, running nearly parallel to midrib across secondary veins from midrib to margin. *Inflorescences* terminal, males with 1-3 (4) flowers in a cluster, females always solitary; bracts triangular, thickly leathery, to 1.0 cm long. *Male flower* with 4 sepals and petals, leathery, outer usually thicker; pedicel 5-10 mm long; sepals ovate; petals lanceolate to ovate; stamens in a massive, weakly 4-angled in central bundle; pistillode absent, anthers 2-thealous, sessile. *Female* flower not seen. *Fruit* globose, dark maroon-purple or dark chocolate, smooth and shiny, 1.4-5.0 x 1.6-5.5 cm, with thick leathery sepals; stigma sessile, with 8-12 crenate, nearly square lobes, black and hardened, to 0.8 cm across; seeds 4-10 with white aril.

Distribution – Luzon, Philippines.

Habitat & Ecology – Lowland and hill forest.

Notes – The fruits closely resemble *G. malaccensis* var. *pseudomangostana* but can be distinguished by the smoother and shiny fruit wall. The leaf also has 2-intramarginal veins but these are not as distinct as in *G. malaccensis*.

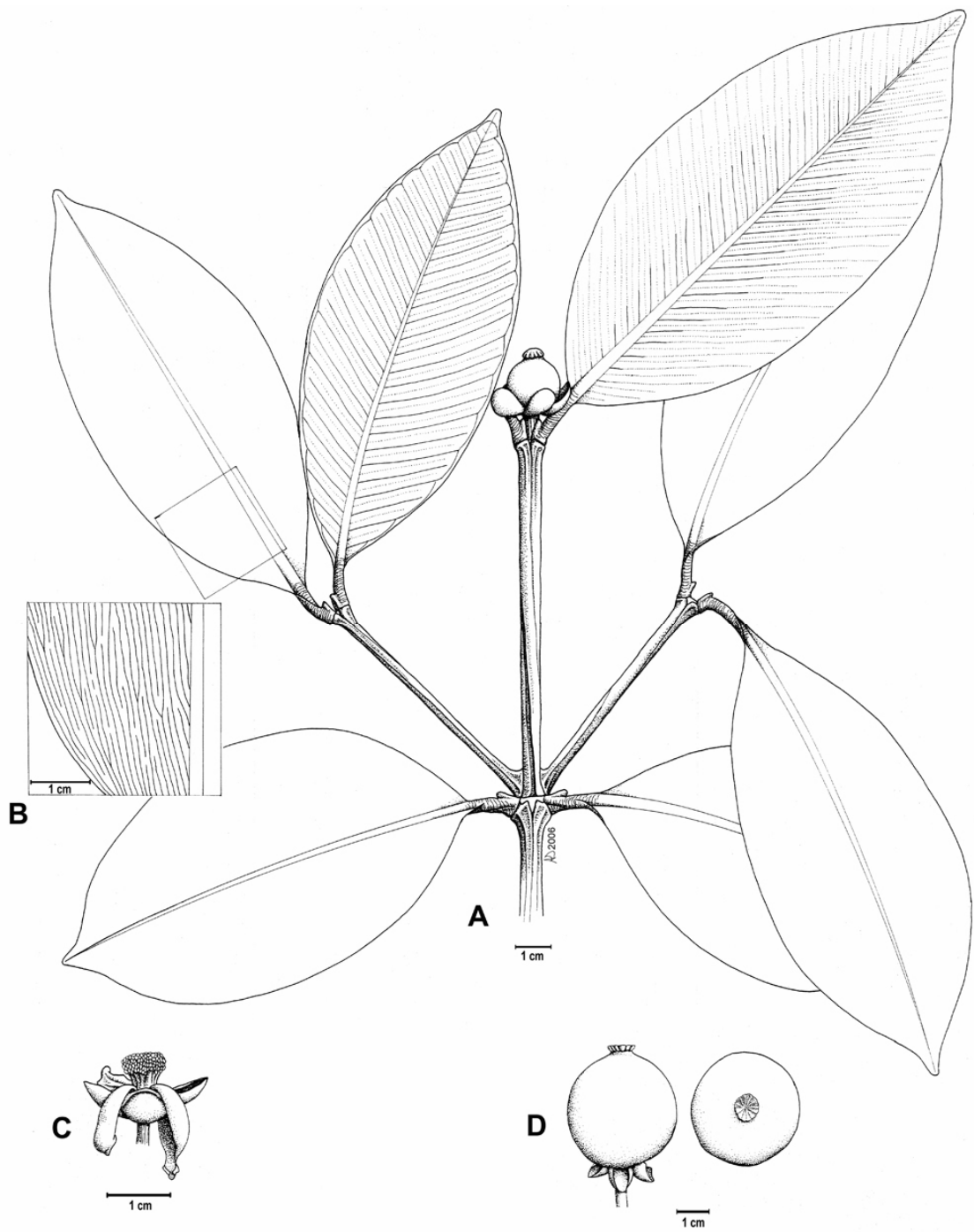
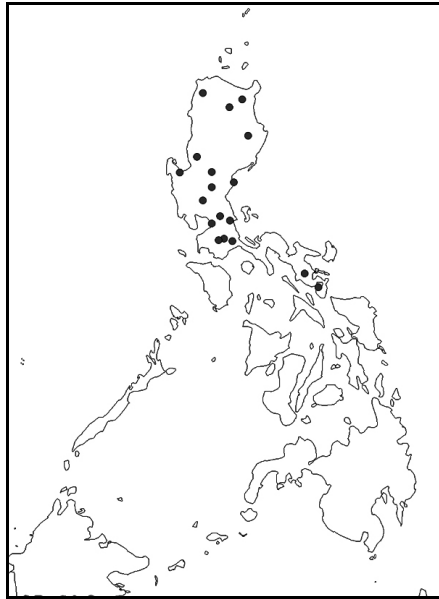


Figure 6.19. *Garcinia venulosa* (Blanco) Choisy. **A.** habit (female); **B.** glandular lines; **C.** male flower; **D.** fruit and stigma [(A, B: *Cuming 1124*, K); (C: *Alvarez 18513*, K); (D: *Curran 17827*, K)]. Drawn by Anna Dorward.



Map 6.17. Distribution of *Garcinia venulosa* (Blanco) Choisy.

#### 6.8.1 *Species with Unknown Status*

The species listed here were originally listed by Jones (1980) as belonging to sect. *Garcinia*. Because of the scarcity and the poor state of the specimens of these species, it is impossible to determine whether they have the morphological characters that would assign them to sect. *Garcinia*. I am classifying them as of unknown status until more specimens can be obtained for study.

#### 1. ***Garcinia blancoi* Pierre**

*Garcinia blancoi* Pierre, Fl. Forest. Cochinch. 1 (1882) 15 & loc. cit (1883) pl.79C. — Lectotype (proposed here): *Gaudichaud-Beaupre* 226 (P) Manila, Philippines.

*Garcinia calleryi* Pierre, Fl. Forest. Cochinch. 1 (1882) 15 & loc. cit (1883) pl.79B. — Lectotype (proposed here): *Callery 56* (P), Palawan Island, Philippines.

*Trees*: no information on the size. *Exudate* yellow *Bole*: no information. *Twigs* 4-angled, longitudinally wrinkled when dry. *Petiole*: stout, 6-9 mm long. *Lamina* leathery, ovate-elliptic, 11-12 x 5 cm; apex acute to obtuse; base acute to acuminate; midrib slightly raised above and raised below; margin entire and finely revolute; secondary veins brochidodromous, fine but distinct, closely arranged, nearly parallel, visible above and slightly raised below. *Inflorescences* terminal, sepals and petals 4; female not found. *Male flower* [in bud] pedicel to 8 mm long; sepals unequal, outer larger than inner, concave, 7.5-8.0 x 5.0-6.0 mm; petals thicker and larger than sepals; stamens in an annular mass surrounding the pistillode; pistillode dwarf fungiform, tipped with a convex cap, 8-10 lobed; anthers sessile, 2-theous. *Fruit*: unknown.

Distribution – Philippines.

Habitat & Ecology – Possibly in the lowlands.

Notes – Most of the description follows that of Pierre (1882). It is only known from two specimens in bud making it difficult to classify.

## **2. *Garcinia erythrosperma* Lauterb.**

*Garcinia erythrosperma* Lauterb. in Bot. Jahrb. Syst. lviii. 26 (1922) — Type: *Lederman 11305*

*Tree* 10-12 m high, slender. *Exudate*: yellow. *Bole*: brown bark grey. *Twigs* subangular, longitudinally wrinkled when dry. *Petiole* 12-15 mm long; coarse towards lamina. *Leaves*

lanceolate, 10-15 x 5-7 cm, coriaceous; apex acuminate and short; base acute; margin sub-revolute; secondary veins brochidodromous, c. 20 pairs, prominent. *Male flower* terminal, solitary, pedicel thick, as long as petiole; sepals 4, persistent, sub-coriaceous, 2 small, lanceolate, sub-acute, 2 larger, concave, widely ovate, subrotundate; staminodes c. 12, free, filament thread like; anthers elliptic, 2-theous, dehiscent longitudinally. *Fruits* yellow, globose, 4.5 cm diameter; pedicel 15 mm long; sepals persistent 7-10 x 4-9 mm; stigma with 8-9 nearly free lobes, 10 mm across; pericarp fleshy; seeds ca. 8, lanceolate or crescent shaped, 12-17 x 4-8 mm long, covered by dark red aril with a reddish black seed when dry.

Distribution – North East New Guinea: Hunsteinspitze 1300 m.

Habitat & Ecology – Mossy montane forest.

Notes – The description of this species is taken from Lauterbach (1922) as I have not seen any specimens. The original type material in Berlin was destroyed during World War II.

Duplicate material was traced in Wroclaw (WRSL) but the specimen was lost in the post to Edinburgh.

### 3. *Garcinia lucens* Pierre

*Garcinia lucens* Pierre, Fl. Forest. Cochinch. 1 (1882) 37. — Lectotype (proposed here): *Korthals s.n.* (lecto BO; iso L) East Kalimantan, Borneo.

*Tree*: size not known. *Exudate*: not known. *Twigs* slightly angled, pale brown to cream.

*Petiole* stout, horizontally striate, angled, 1.6-2.0 cm long. *Lamina* thinly leathery, elliptic

to ovate, 12.5-16.0 x 4.9-7.7 cm; apex acute with a blunt apex; base broadly acute to obtuse; midrib flattened to slightly raised above, raised and pointed below; margin entire and weakly revolute; secondary veins brochidodromous. *Glands* fine, in wavy longitudinal lines, running nearly parallel to the midrib. *Inflorescences* terminal, only males known, in cluster of 3-10 flowers; sepals and petals 4. *Male flower*: [in bud] pedicel longer than petiole, to 25 mm; sepals ovate to broadly elliptic, 3.6- 4.0 x 2.8-3.1 cm; petals oblong-elliptic, 3.0-3.5 x 5.5-6.2 cm; stamens in an annular mass, surrounding half of the pistillode; pistillode a slender fungiform shape, tipped by a 4-lobed cap; anthers 2-theous. *Fruit* Unknown.

Distribution – East Kalimantan.

Habitat & Ecology – Lowland forest.

Notes – This species is represented by multiple collections of male and sterile specimens by Korthals, possibly from one locality in Kalimantan. No other specimens have been found. Although it has the distinct character of a long pedicel, the male flower is still in bud making it difficult to study.

#### **4. *Garcinia moulmeinensis* Pierre ex Vesque**

*Garcinia moulmeinensis* Pierre ex Vesque, Monogr. Phan. 8 (1893) 394. — Lectotype (proposed here): *Wallich 4852E* (P) Moulmein, Burma.

*Size* not known. *Exudate* not known. *Twigs* not known. *Petiole* short, 5-9 mm long.

*Lamina* leathery, elliptic or oblong, 11.7-12.5 x 3.7-6.0 cm, pale brown; apex sharply

acute or not; base broadly acute or nearly round; margin entire and revolute; midrib flattened above and raised below; secondary veins fine, visible below, a mixture of eucamptodromous and cladodromous, running nearly parallel, c.85° from the midrib; glandular lines faint but fine and inconspicuous below. *Inflorescences* possibly terminal, in cluster of 4-6 flowers; sepals and petals 4. *Male flower* not found. *Female flower* nearly sessile or with a short pedicel, to 3 mm long; sepals membranous, broadly ovate; petals concave, to 5 mm long; staminodes absent; ovary short, tipped by six locular, convex stigma. *Fruit* not found.

Distribution – Burma.

Habitat & Ecology – Lowland.

Notes – Represented only by the type specimen of two leaves without twigs. The flowers were detached and kept in a capsule.

##### **5. *Garcinia squamata* Lauterb.**

*Garcinia squamata* Lauterb., Bot. Jahrb. Syst. 58 (1923) 21. — Lectotype (proposed here):  
*Gjellerup 1167* (BO), Arfak Mountains, Papua, Indonesia.

*Small tree* to 4 m tall. *Exudate* yellow. *Twigs* round to slightly angled, pale grey-brown to brown. *Petiole* blackish, weakly channelled on the adaxial surface, to 1.0 cm long.

*Lamina* thickly leathery, obovate, 3.0-5.0 x 1.8-3.0 cm; apex nearly round; base acute; margin entire and revolute; midrib flattened or weakly raised above and raised below, especially near the petiole; secondary veins very fine, inconspicuous below, camptodromous; tertiary veins inconspicuous. *Inflorescences* terminal or axillary, flowers

solitary; sepals and petals 4. *Male flower* not seen. *Female flowers*: pedicel stout, to 9 mm long; sepals unequal in size, two outer larger, concave or nearly round, to 6 mm long; petals yellow, broadly elliptic, to 9 mm long; ovary 4-6 x 4-5 mm; stigma black, rugose; staminodes not present. *Fruit* Unknown.

Distribution – New Guinea.

Habitat & Ecology – Montane at 1900 m.

Notes – Represented only by female specimens in very young fruit which makes it difficult to classify.

### 6.8.2 *Excluded Species*

Listed here are species excluded from sect. *Garcinia* which were treated by Jones (1980) as belonging to the section. No description is given for each species except notes on the morphological characters that separate them from sect. *Garcinia* and, where appropriate, details are given of their phylogenetic position in the analysis of DNA sequence data (Chapter 5).

#### 1. ***Garcinia anomala*** Planch. & Triana.

*Garcinia anomala* Planch. & Triana, Ann. Sci. Nat. ser. 4, 14 (1860) 329; Hook f., Fl. Brit. Ind. 1 (1872) 266; Kurz, J. Asiat. Soc. Bengal, Pt. 2., Nat. Hist. 43 (1874) 87; Kurz, Forest. Fl. Burma (1877) 89; Pierre, Fl. Forest. Cochinch. 1 (1882) 10, Fl. Forest Cochinch. 1 (1883) pl.79L; Vesque, Monogr. Phan. 8 (1893) 369; Engler, Nat. Pflanzenfam, ed. 2, 21 (1925) 223; Maheshwari, Bull. Bot. Surv. India 6 (1964) 117. — Lectotype (proposed here): *Hooker f. & Thomson 14* (lecto E; iso A, L), Khasia, India.

Distribution – Eastern India to northern Thailand.

Habitat & Ecology – Montane to 1500 m.

Notes – Differs from sect. *Garcinia* by the axillary inflorescences in thyrses and the male flowers having glomerate, more or less round bundles of stamens surrounding the pistillode.

**2. *Garcinia chapelieri* (Planch & Triana) H. Perrier.**

*Garcinia chapelieri* (Planch & Triana) H. Perrier, in Mem. Mus. Hist. Nat., Paris, 24 (1948) 97. — *Ochrocarpos chapelieri* Planch. & Triana in Ann. Sc. Nat. Bot. Ser. 4, 13 (1860) 366; Vesque, Monogr. Phan. 8 (1893) 522. — *Garcinia polyphlebia* Baker in Journ. Linn. Soc. 22 (1886) 447.

Distribution – Madagascar.

Habitat & Ecology – Lowland to hill forest.

Notes – No type specimens were available to study this species and the protologue only gave descriptions of the female specimens. However, other material obtained from P included a few male specimens. Differs from sect. *Garcinia* through the male flowers having 4 free bundles (vs. lobes) of stamens surrounding the pistillode.

Pistillode slender, filament-like. The fruit wall has lobes and is tipped by small, 4 fused notch-like stigmas.

**3. *Garcinia costata* Hemsl. ex King.**

*Garcinia costata* Hemsl. ex King, J. Asiat. Soc. Bengal, Pt. 2, Nat. Hist. 59 (1890) 161. —

Lectotype (proposed here): *King's Collector 5375* (lecto K; iso L).

Distribution – Peninsular Malaysia.

Habitat & Ecology – Hill forest to 600 m.

Notes – Vegetatively, the large, cabbage-like and ribbed leaves are distinctly different from sect. *Garcinia*. The inflorescences are thyrses rather than in clusters as in sect. *Garcinia*, the male flower is similar to *G. anomala* by the mass of stamens surrounding the pistillode and the fruit has a lobed or segmented wall. Molecular analysis shows this species to be positioned outside the clade of sect. *Garcinia*, forming a monophyletic group together with *G. maingayi* and *G. trianaii*, closely related to sect. *Brindonia*.

**4. *Garcinia maingayi* Hook f.**

*Garcinia maingayi* Hook f., Fl. Brit. India 1(1872) 267. — Lectotype (proposed here):

*Cuming 161* (lecto K) Peninsular Malaysia.

*Garcinia baillonii* Pierre, Fl. Forest. Cochinch. 1 (1882) 18; loc. cit (1883) pl. 83. —

Lectotype (proposed here): *Beccari 2498* (lecto K) Kuching, Sarawak.

Distribution – Sumatra, Peninsular Malaysia and Borneo.

Habitat & Ecology – Hill forest to lower montane 900 m

Notes – Closely related to *G. costata* but the leaf is smaller and the fruit wall not lobed but smooth with stripes of green and yellow. Molecular analysis positioned *G.*

*maingayi* outside the sect. *Garcinia* clade where it forms a monophyletic group together with *G. trianaii* and *G. costata*.

**5. *Garcinia propinqua* Craib, Kew Bull. 3(viii), 85 (1924).**

*Garcinia propinqua* Craib, Bull. Misc. Inform. Kew (1924) 85. — Lectotype (proposed here): *Kerr 5611* (lecto K), Doi Chieng Dao, Thailand

Distribution – Northern Thailand.

Habitat & Ecology – Montane forest to 1200 m.

Notes – Represented only by a male type specimen. Morphologically it is similar to *G. anomala* but with a smaller stature, leaves and flowers.

**6. *Garcinia pseudo-guttifera* Seem.**

*Garcinia pseudo-guttifera* Seem., Fl. Vit. (1865) 11; A.C. Smith & S. Darwin, J. Arnold Arb., 55 (1974) 245; A.C. Smith, Fl. Vit. Nova (1981) 343-344. — Type: *Seeman 50* (holo K; iso BM) Navua River, Serua Province, Viti Levu (Fiji)

Distribution – New Hebrides, Fiji and Tonga.

Habitat & Ecology – From hill to lower montane forest.

Notes – Differs from sect. *Garcinia* by the inflorescences in thyrses which contain many flowers (to 40).

**7. *Garcinia tonkinensis* Vesque.**

*Garcinia tonkinensis* Vesque, Monogr. Phan. 8 (1891) 379. — Lectotype (proposed here):  
*Balansa* 4359 (lecto P; iso K) Tonkin, Vietnam.

Distribution – Vietnam

Habitat & Ecology – Not known.

Notes – Poorly known with only two female specimens. Differs from sect. *Garcinia*  
by the thyrsoid inflorescences positioned terminally and axillary.

**8. *Garcinia trianaii* Pierre**

*Garcinia trianaii* Pierre, Fl. Forest. Cochinch. 1 (1882) 18; Fl. Forest Cochinch. 1 (1883) pl.  
80L. — Lectotype (proposed here): *Beccari* 360 (lecto K) Sarawak.

Distribution – Borneo.

Habitat & Ecology – Lower to upper montane forest 1900 m.

Notes – Similar morphologically to *G. maingayi* but has smaller leaves and flowers.

Molecular analysis supports close relationship to *G. maingayi*.

**9. *Garcinia vidua* Ridley.**

*Garcinia vidua* Ridley, Kew Bull. 17 (1938) 115. — Lectotype (proposed here) *Haviland*  
2222 (lecto K; iso L, UC) Kuching, Sarawak.

Distribution – Sarawak.

Habitat & Ecology – Lowland to hill forest 600 m.

Notes – Differs from sect. *Garcinia* by having nearly sessile axillary inflorescences, with one to three flowers; stamens in the male flowers are very few in a central mass without a pistillode. The fruits are small and tipped with a sessile and rugose stigma.

Both flower and fruit characters suggest *G. vidua* is a member of sect. *Brindonia* sensu Jones (1980).

## 6.9 List of Exsiccatae

### 1. *G. acuticosta* Nazre

Cockburn, P.F. FRI10559 (L ex. KEP) *Batu Biwa, Terengganu* — Henderson, M.R. 19654 (K) *Kelantan, Malaysia* — Loh, H.S. FRI13410 (L ex. KEP) *Gading Forest Reserve, Selangor* — Nazre, M. BB01 (E) *Bukit Bauk Forest Reserve, Terengganu*.

### 2. *G. cataractalis* Whitmore

Whitmore, T.C. FRI20350 (K ex. KEP) *Sungai Kerbat, Terengganu*.

### 3. *G. celebica* L.

Aban Gibot 54837 (L, SAN), *Pulau Sakar, Lahad Datu, Sabah* — Ag. Amin Sigun SAN104109 (E, SAN) *Membakut, Sabah*, SAN 126779 (E), SAN 98765 (E) — Ahmad Shukor AS107 (SING) *Kijal, Terengganu*, AS29 (SING) *Jason Bay, Johor* — Alvins, V.M. (SING) *Melaka* — Anderson, J.A.R. S30779 (L) & S30790 (E, L) *Gunung Api, Baram, Sarawak* — Anderson, M. 28 (P) *Calcutta, India*.

Backer, C.A. 27885 (L) *Pulau Kangean, Java* — Balgooy, M.M.J.v. 6818 (A) *Pulau Maikoor, Aru Is.* — Bayak 2150 (K) *Jesselton, Sabah* — Beguin 307 (L) *Bengkalis, Sumatra* — Bernard Lee S41863 (L) *Datu Protected Forest, Sarawak* — Blume, C.L. s.n. (L) *Java Island*; 1273 (L); 1669 (L) — Borssum Waalkes, J.v. 321 (L) *Pulau Panaitan, Java* — Boschbouwproefstation bb0945 (L), bb9316 (L ex. BO) *Lampung, Sumatra*; bb10104 (L) *Waii, Ambon Island*; bb5708 (L) *Nias Island, Sumatra* — Burghardt, C.P. bb5855 (L) *Boeton, Sulawesi* — Burkill, H.M. HMB259 (SING) *Pangkor Island, Perak*; HMB 999 (SING, L) *Pulau Angsa, Singapore*.

Castro & Melegrito 1499 (UC) *Banggi Island, Sabah* — Celestino & Ramos 23055 (L) *Sagpangan, Aborlan* — Cenabre, A.L. 29179 (A) & 29229 (A) *Puerto Princesa*

- City, Palawan Islands* — Chew, W.L. CWL1058 (A, L, SING) *Sungai Melinau, Baram* — Collins, D.J. s.n. (E, K, L), 1693 (L), 898 (L) *Sriracha, Thailand* — Congdon, G., Hamilton, C. 444 (A) *Haad Yai, Thailand*. — Coode, M.J.E., Wong, K.M. 6856 (A) *Tutong, Brunei* — Corner, E.J.H. s.n. (SING) *Sungai Sedili Kecil, Johor*; SFN 25766 (SING) *Kuala Bebar, Pahang* — Croft, G.W. 5684 (UC) *Indo-China* — Cuming, H. 2296 (K) *Melaka* — Curran, H.M. 3787 (K) *Palawan Islands* — Curtis, C. s.n. (SING) & 690 (SING) *Telok Bahang Forest, Pulau Pinang*; 2453 (SING) *Pulau Tengah, Pulau Pinang*; 3438 (SING) *Beruas, Perak*.
- Diepenhorst HB2152 (L) *Priaman, Sumatra Barat*.
- Eberhardt 3072 (P) *Tinh Thua Thien-Hue, Vietnam* — Edano, G. 77389 (SING) *Palawan Islands* — Elmer, A.D.E. 12617 (E, L), 12711 (A) *Addison Peak, Palawan*; 18480 (L, UC) *Puerto Princesa City, Palawan Islands*.
- Falconer, H. (GH, L) *Molamyaing, Burma* — Flerry, F. 30203 (P) *Thu Dau Mot, Vietnam* — Forbes, H.O. 539 (L) *Sogeri Island, Papua New Guinea* — Foreman, D.B. LAE52091 (L) *Tavanatangir Harbour, Papua New Guinea*, LAE52129 (L) *Rabaul District, Papua New Guinea* — Foxworthy, F.W. 12577 (SING) *Telok Bahang Forest Reserve, Pulau Pinang*; 1741 (SING) *Pulau Pangkor, Perak*
- Goodenough, J.S. (SING) *Changi, Singapore* — Griffith, W. s.n. (K), 857 (K, L, P) *Melaka*; s.n. (L) *Molamyaing, Burma*; Gwee, Paul, Samsuri Ahmad, Saifuddin, Jacky GAT141 (SING) *Tanjung Chek Java, Singapore*.
- Hamilton, F.B. 1113 (E) *Calcutta, India* — Harmand 1074 (P) *Laos* — Harreveld, P.v. (L) *Java* — Haviland, G.D. 2329 (K, SING) *Bintulu, Sarawak* — Helfer, J.W. 21 (A, E, L) — Henderson, M.R. 18412 (SING), *Pulau Tioman, Pahang* — Holmberg, P.J. 868 (SING) *Malacca* — Holttum, R.E. (SING) *Singapore*; 17673 (SING) *Kuala Terengganu, Terengganu* — Hombron, J.B. (P) *Peninsular Malaysia* — Hoogerwerf, A. 132 (L) *Pulau Java* — Hooker, J.D., Thomson, T. s.n. (GH, L) — Huc., R. RH522 (A, L) *Wai Kambas, Palembang*.
- Ilias Paie S36331 (L) *Katibas, Kapit*.
- Kairo, A. NGF45063 (L) *New Guinea* — Karta 235 (L, BO), 380 (L) *Semarang, Pulau Karimunjawa* — Kerenga, K. LAE74243 (L) *New Britain, Bismarck Archipelago*; LAE74246 (L) *Papua New Guinea* — Kerr, A.F.G. 1073 (L), 8572 (K) *Thailand*; 11179 (K, L) *Kaw Tao Surat, Thailand*; 12752 (L) *Surat Thani, Thailand*; 14078 (L) *Adang Island, Thailand*; 20261 (K) *Chaiyapum, Thailand*; 8513 (K, UC) *Chaiyaburi, Thailand* — King's Collector s.n. (K) *Andaman Is.*; 483 (K) *Nicobar Is.* — Koelz, W. 24900 (UC) *Garro Hills Assam* — Koorders, S.H. 21958B (L), 30833 (L, BO), 33366B (L) *Banyumas Java*; 23646 (L), 2937B (L), S.H. 2960B (L, BO), 34266B (L) *Java Island* — Korthals, P.W. 1313a[4601] (P) *Banjarmasin Kalimantan* — Kostermans, A.J.G.H. 193A (L) *Manokwari, Irian Jaya*; 24028 (A) *Sri Lanka*; BW15509 (L) *Teluk Irian Jaya* — Kostermans, A.J.G.H., Kuswata Kartawinata 49 (A) *Ujung Kulon Nature Reserve Peutjang Is.* — Kostermans, A.J.G.H., Wirawan 661 (L) *Pergunungan Ruteng Lesser Sunda Is.*; 821 (A) *Flores Lesser Sunda Is.* — Kurz, W.S. 24 (P) *Andaman Is.*
- Lace, J.H. 4675 (E) *Myanmar* — Lai, J. LJ154 (SING) *Pulau Ubin, Singapore*; Lai, J. LJ7 (SING) *Sentosa Island, Singapore* — Lam, H.J. (L) *Java Is.* — Laman, T., Ismail A Rachman, Edi Mirmanto TL545 (L) *Gunung Palung, West Kalimantan*;

- TL559 (A ex. BO) *West Kalimantan* — Lesmy Tipot FRI33980 (L) *Pantai Aceh Forest Reserve, Pulau Pinang*; FRI34940 (L) *Pulau Pangkor Perak* — Liew, K.C. 168 (L ex. KLU) *Pulau Redang Terengganu*.
- M.C. Lakshnakara 522 (L) Chantabun, Thailand — Mahamud 14983 (E) Rompin Pahang — Mahamud 14983 (SING) Rompin Pahang — Maingay, A.C. 150 (K) Melaka, Malaysia — Manalo, A. 7430 (K) — Palawan Islands — Maxwell, J.F. 85-167 (A) Songkhla, Thailand — Maxwell, J.F. 85-300 (A, L) Nakhon Si Thammarat Peninsular; 85-474 (E, A) Na Mom Songkhla; 86-68 (A) Songkhla; 88-190 (L) Chiang Mai Thailand — Mohamed Haniff, Mohd Noor 7570 (SING) Pulau Chupak, Kedah — Mohd Noor (UC ex. SING) Botanic Gardens Singapore — Mohd Noor, Samsuri Ahmad MN25 (E, A) Pulau Pemanggil, Johor — Muin Chai SAN29720 (L) Lahad Datu, Sabah.
- N.G. Nair 3526 (L) *Sawai, Nicobar Is.* — Nengah Wirawan 392 (L), 394 (L) *Ujung Kulon Nature Reserve Peutjang Island* — Neth. Ind. For. Service bb30737 (L), bb30792 (L), bb30816 (L), bb30890 (L) *Serui, Irian Jaya*; bb30940 (L) *New Guinea*; bb32514 (L) *Sulawesi*; Cel./II-112 (L) *Malili, Sulawesi* — Noltee, A.C. 4022 (L ex. BO) *Pekalongan Java*.
- P. Siriruga 1013 (A) *Songkhla Thailand* — Phuang van dier 36 (K ex. P) *Ba Ria Vietnam* — Pierre, J.B.L. 3633 (K ex. P) *Phuoc Le, Vietnam*; 3637 (P) *Laos*; 3695 (P); *Xa Phuoc Le, Vietnam*; 4607 (P) *Java Is.*; 700 (A) *Vietnam*; 700 (K) *Cochinchina*; 700 (P) *Ho Chi Minh City Vietnam* — Poilane, M.E. 14205 (K) *Cambodia*; 14781 (K ex. P) *Kampot, Cambodia*; 24088 (P) *Do Mai Vietnam*; 27208 (P) *Siemreab Cambodia*.
- Rahim Ismail 97929 (L) *Kuantan Pahang* — Ramlanto, Zainal Fanani 721 (L) *Tolitoli Sulawesi* — Ridley, H.N. (SING) *Dinding Perak*; 1096B (SING) *Kuala Pahang, Pahang*; 1962 (SING) *Bukit Timah, Singapore*; 4791 (K, SING) *Pulau Ubin, Singapore*; 7966 (SING) *Pulau Pangkor, Perak*; 9023 (K, SING) — Ridsdale, C.E. SMHI188 (K, L, UC), SMHI195 (A, L) *Puerto Princesa City, Palawan Islands* — Rossum, H.I.P. 171 (L) *Pulau Belitung Sumatra* — Royen, P.v. 5464 (A) *Waigeo Island Irian Jaya*
- Seidenfaden, G. 2585 (SING) *Tha Chang, Thailand* — Sidek bin Kiah S366 (SING) *Naka Kedah*; S77 (L) *Pulau Pawai, Singapore* — Sinclair, J. 5791 [SFN38548] (E ex. SING) *Machua Khali, Bangladesh* — Smith, J.E. (K) *Calcutta India* — Smythies, B.E. 14052 (L) *Sungai Satok Kuching* — Soepadmo, E. 311 (L) *Suaka Mergasatwa Ujungkulon Peutjang Island* — Steenis, C.G.G.J. bb28917 (L) *Papua New Guinea* — Stevens, P., Lelean, Y. LAE58551 (L) *Hoskins, Papua New Guinea* — Stone, B.C. 14477 (L ex. KLU) *Pantai Aceh Forest Reserve, Pulau Pinang* — Suppiah, T. FRI14761 (L) *Pahang, Malaysia*.
- T & P KL2835 (SING) *Dungun Terengganu* — Teijsmann, J.E. s.n. (A, BO, A) *Hutumuri Ambon Island*; s.n. (L) *Tanjung Pandan Belitung*; 11843 (L) *Pangkadjene, Sulawesi*; 12392 (L) *Sulawesi*; 8045 (L) *Mampawah, Kalimantan*; 8960 (L ex. BO) *Timor Lesser Sunda Is* — Treub, M. 4169 (K) *Pulau Bangka, Sumatra*.
- Vethevelu, P. 29696 (L) *Menchali Forest Reserve Pahang* — Vidal y Soler, S. 2152 (A, K) *Iwahig, Palawan* — Villamil, A. 342 (A) Sabah — Vogel, E.F. 1390 (L) *Suaka Mergasatwa, Ujungkulon Peutjang Island*.

Wallich, N. 4852 (K) *Penang, Malaysia*; 4854 (K) *Sylhet* — Whitmore, T.C. FRI 15746 (L ex. KEP) *Hulu Perak, Perak*; FRI 20509 (L) *Perlis, Malaysia* — Womersley, J.S. NGF24961 (L) *Morobe, Papua New Guinea* — Wood, D.D. 2451 (UC) *North Borneo* — Wyatt-Smith, J. 80555 (L) — *Pisang Is., Johor*.  
Yahya SFN21419 (SING) *Penang, Malaysia*.

#### 4. *G. diospyrifolia* Pierre

##### i. *G. diospyrifolia* Pierre var. *diospyrifolia*

Burley, J.S., Tukirin 703 (A) *Sungai Kahayan, Central Kalimantan*. — Chan, Y.C. FRI19844 (L ex. KEP) *Lesong Forest Reserve, Pahang*; FRI25052 (KEP, K, L) *Bukit Bauk Forest Reserve, Terengganu*. — Clemens, J., Clemens, M.S. 22104 (K ex. SAR) *Sungai Rejang, Sarawak* — Cockburn, P.F. FRI 11066 (L ex. KEP) *Chini Forest Reserve, Pahang* — Corner, E.J.H. s.n. (K ex. SING) *Sedili, Mersing*; E.J.H. SFN28653 (L) *Johore, Malaysia*.  
Jacobs, M. 5125 (K ex. L) *Kuching, Sarawak* — Kochummen, K.M. 77731 (L) *Mersing, Johor* — Korthals, P.W. s.n. (BO) *Borneo*; s.n. (L) *Banjarmasin, Kalimantan*; s.n. (L) *Sekubang, Kalimantan* — Laman, T., Ismail A Rachman, Edi Mirmanto TL1204 (A), TL987 (A) *Gunung Palung National Park, Kalimantan* — Laumonier, Y. YL6414 (L) *Siak, Sumatra*.  
Maskuri 155 (L) *Tanah Merah, Kalimantan* — Mohamad Shah bin Mohamad Nur 1560 (SING) *Bukit Terom, Pahang* — Ridley, H.N. 9005 (K, SING) *Siak, Sumatra* — Samsuri Ahmad, Ahmad Shukor SA412 (A, SING) *Lesong Forest Reserve, Pahang* — Soengeng Reksodihardjo 47 (L ex. BO) *Samarinda, Kalimantan* — Soepadmo, E. 170 (E) *Pekanbaru, Riau* — Zainal Ariffin 914 (A, K) *Wain, Kalimantan*.

##### ii. *G. diospyrifolia* Pierre var. *arborea* Nazre

Aban Gibot SAN31195 (L) *Sungai Serudong, Tawau* — George Mikil SAN31886 (L ex. SAN) *Tenom, Sabah* — Ilias Paie S17017 (K) *Sabal Forest Reserve, Sarawak* — King's Collector 5460 (K, UC), 10958 (P, L), 7232 (K, KEP, P) *Larut, Perak*.  
Majawat SAN102415 (K, SAN) *Kinabatangan, Sabah* — Kuswata Kartawinata 856 (L, BO), 942 (L) *Jaro, South Kalimantan* — Martin, P.J. S37905 (K, L) *Gunung Gading, Sarawak* — Pereira, J.T., A. Berhaman, J. Sugau, J., Madani, L., Baraham B JTP65 (E) *Penampang, Sabah* — Vogel, E.F. 2777 (L) *Sumatra Island*.

##### iii. *G. diospyrifolia* Pierre var. *minor* Nazre

Nazre, M. BB08 (E) *Bukit Bauk Forest Reserve, Terengganu* — Rosemary Gianni 541 (L) *Tasik Bera, Pahang* — Wong, K.M. 32216 (A ex. KEP) *Pasoh Forest Reserve, Negeri Sembilan*.

**5. *G. discoidea* Nazre**

- Burkill, H.M. HMB1854 (L ex. SING) *Sedili, Mersing* — Chew, W.L. CWL843 (KEP, SING, UC) *Cameron Highlands, Pahang* — Corner, E.J.H. s.n. (SING), 34940 (E, L, SING) *Mawai, Johor*; SFN31499 (SING) *Kuala Nerang, Kedah*.  
Kochummen, K.M. FRI2756 (A) *Bukit Arang, Selangor* — Nazre, M. BB02 (E) *Bukit Bauk Forest Reserve, Terengganu* — Ng, F. FRI5017 (L) *Bukit Fraser, Pahang* — Samsuri Ahmad, Ahmad Shukor SA462 (SING) *Pahang, Malaysia*.

**6. *G. exiguus* Nazre**

- Coode, M.J.E., Dransfield, J., Kirkup, D., Idris M Said 7886 (A ex. BRUN) *Ulu Belalong, Brunei* — Paul Chai, P.P.K. S31750 (L) *Bintulu, Sarawak*.

**7. *G. harmandii* Pierre**

- Harmand 349 (P) Laos — Pierre, J.B.L. 1371 (A, K) *Thanh Ho Bien Ho, Vietnam*; 776 (K, P) *Cambodia* — Poilane, M.E. 17388 (P) *Kampong Spoe, Cambodia* — Thorel, M. 1018 (K ex. P) *Mekong River, Cambodia*.

**8. *G. malaccensis* Hk. f.**

**i. *G. malaccensis* var. *mangostana***

- Alvins, V.M. 2742 (SING) *Merlimau, Melaka* — Ashton, P.S. BRUN3392 (SAR ex. BRUN) *Belalong, Brunei* — Chelliah, S. 104374 (KEP, L) *Bukit Bauk Forest Reserve, Terengganu* — Cockburn, P.F. FRI10887 (L) *Sungai Keliu, Pahang* — Cockburn, P.F. FRI8181 (KEP, L) *Sungai Kemia, Besut* — Kiah Hj. Salleh (SING) *Mandai Road Village*.  
Kochummen, K.M. 76634 (L) *Sungai Menyala Forest Reserve, Negeri Sembilan* — Lindong, Santiago, A. KEP71525 (L) *Banang Forest Reserve, Johor* — Maingay, A.C. 149 (GH, K, L); 1643 (K) *Malacca, Malaysia* — Mohd Noor 1529 (SING) *Bukit Timah, Singapore*.  
Neth. Ind. For. Service bb30045 (L) *Indragiri Hulu, Riau* — Nazre, M. BB 9 (E) *Bukit Bauk Forest Reserve, Terengganu* — Ridley, H.N. (SING) *Chua Chu Kang, Singapore* — Whitmore, T.C. FRI8682 (KEP, L) *Sungai Kahang, Kluang*; FRI8986 (L) *Sungai Loh, Terengganu*.

**ii. *G. malaccensis* var. *pseudomangostana* Nazre**

- Arbainsyah AA1004 (BO) *Balikpapan, Kalimantan* — Elmer, A.D.E. 21113 (K, L, SING) *Tawau, Sabah* — Fidilis Krispinus, Sumbing SAN96472 (L) *Luasong, Kalabakan*.  
James Ahwing 38173 (L) *Gomantong Forest Reserve, Sandakan* — Madani, L. 50574 (L) *Sandakan, Sabah* — Meijer, W. SAN19236 (L) *Tawau, Sabah* — Nazre, M. S1

(E) *Forest Research Centre, Sandakan* — Sigin, Joseph SAN97194 (L) *Kretam Forest Reserve, Sandakan*.

**9. *G. mangostana* L.**

Anon. 1667 (SING) *Malacca, Malaysia*; 7 (L) *Java*.

Anderson, J.A.R., Ilias Paie 15371 (L) *Gunung Gading, Sarawak* — Bakhhuizen van den Brink Sr., R.C. 3417 (L ex. BO) *Gunung Kembang, Java* — Blume, C.L. (L) *Java* — Bradley, M.C. (GH) *Thailand* — Brass, L.J. 32410 (A) *Kassam, Papua New Guinea*

Daud, Tachun bin Baba SFN36093 (L) *Gunung Gading, Sarawak* — Fidilis Krispinus, Sumbing SAN96472 (E) — *Luasong, Kalabakan* — Forest Department A1069 (SING) *Sandakan* — Fretes, d. (L) *Ambon Island*

Furtado, C.X.D.R. 34866 (A) *Botanic Gardens, Singapore* — Hasan Pukol 5726 (L, SING ex. BRUN) *Muara, Brunei* — Ilias Paie S15825 (L) *Labang Forest Reserve, Bintulu* — Junghuhn, F. (L) *Java* — Lai, S.T. S68180 (SAR) *Sungai Engkari, Sarawak*

Laman, T., Ismail A Rachman, Edi Mirmanto TL1300 (A) *West Kalimantan* — Maxwell, J.F. 86-499 (A) *Patalung, Thailand* — Mohamad Shah bin Mohamad Nur, Mohd Noor MS1782 (L, A, SING) *Pahang* — Mohamed Haniff 15649 (SING) *Teluk Intan, Perak* — Mohd Noor 146 (A) *Botanic Gardens, Singapore*

Nixon, A.B. 18690 & 18695 (A) *Tavoy, Burma* — Othman Ismawi S32968 (L) *Kuching*; S43603 (L) *Sungai Pakau, Belaga* — Sibat Luang S23041 (L) *Bukit Mentagai, Sarawak* — Sinclair, J. 6842 (E) *Jurong, Singapore* — Stevens, P., Streimann, H. LAE54735 (A, L) *Morobe, Papua New Guinea* — Stomps, T.J. (L) *Berastagi, Sumatra*

Telado 1922 (A) *Papar, Sabah* — Wong, K.M. WKM1597 (A) *Bukit Patoi, Temburong Brunei* — Wray, L. 5587 (SING) *Taiping, Perak* — Zollinger, H. 779 (L) *Java Island*.

**10. *G. moselleyana* Pierre**

Elmer, A.D.E. 7187 (K) *Leyte Island, Philippines* — Moseley, H.N. (K) *Basilan, Philippines* — Wenzel, C.A. 3036 (K, UC) *Placer, Philippines*.

**11. *G. nitida* Pierre**

Argent, G., Amiril Saridan 9318 (A, E, K) *Central Kalimantan* — Beccari, O. 1179 (A, K) *Sarawak*; 2104 (A) *Gunung Berumput, Sarawak* — Blicher, U. S59879 (L) *Kapit, Sarawak* — Church, A.C., Mahyar, U.W., Indah, Ismail, Hamzah 1299 (E ex. BO) *Central Kalimantan* — Clemens, J., Clemens, M.S. 40402 (UC); 40620 (A, K) *Kinabalu National Park*

Fidilis Krispinus, Sumbing SAN113356 (A) *Sungai Mantuluk, Keningau* — Haviland, G.D. 2339 (K) *Kuching* — Ilias Paie 13317 (L, SAR) *Perigi Mountain, Sarawak* — J.K. Lajangah 44731 (K, L SAN) *Kinabalu National Park, Sabah* — Sibat

Luang S22499 (A, K, SAR) *Anap Forest Reserve, Sarawak* — Zainal Ariffin AA914 (A) *Wain, Kalimantan*.

**12. *G. ochraceus*** Nazre

Streimann, H., Martin, N. LAE52886 (E); LAE52890 (E) *Amanab, New Guinea*.

**13. *G. penangiana*** Pierre

- Brunig, E.F. S1142 (K ex. SAR) *Temburong, Brunei* — Burbidge, F.W. (K) *Borneo* — Burkill, I.H., Mohamed Haniff 15716 (SING) *Kuala Lipis, Pahang* — Chan, Y.C. FRI25061 (L ex. KEP) *Bukit Bauk, Terengganu* — Church, A.C., Mahyar, U.W., Afriastini 1762 (A, E, BO) *Sungai Serawai, Kalimantan*; 1840 (A) *West Kalimantan* — Church, A.C., Mahyar, U.W., Indah, Ismail, Hamzah 1216 (A) *Central Kalimantan* — Cockburn, P.F. FRI7877 (L ex. KEP) *Labis Forest Reserve, Johor*; FRI8242 (L ex. KEP) *Besut, Terengganu* — Corner, E.J.H. (SING) *Kemaman, Terengganu* — Curtis, C. s.n. (SING), 1549 (SING) *Telok Bahang Forest Reserve, Pulau Pinang*; 1430 (SING) *Prince of Wales, Malaysia*.
- Daud, Tachun bin Baba SFN36067 (L, SING) *Sarawak* — de Wilde, W.J.J.O., de Wilde-Duyfjes, B.E.E. 12678 (K) *Aceh, Sumatra*; 18011 (K); 18028 (K) *North Sumatra* — Foxworthy, F.W. 1185 (SING) *Johor* — Galau S14977 (K ex. SAR) *Kuching* — Henderson, M.R. 21689 (K, SING) *Pulau Tioman, Pahang*; 24836 (SING) *Ketil, Kelantan* — Ilias Paie, James Dawos Mamit S28974 (E) *Banjaran Bungoh, Sarawak*.
- Jacobs, M. 5444 (K) *Belaga, Sarawak* — Kadim, Mohd Noor KN591 (A, K, L, SING) *Pulau Tioman, Pahang* — King's Collector 3583 (L ex. KEP), 6769 (L ex. KEP), 7565 (SING) *Larut, Perak* — Kirkup, D., Wong, K.M., Dransfield, S., Niga 265 (K ex. BRUN) *Brunei* — Maurus, Heya, Suali SAN72470 (A) *Sook, Sabah* — Meijer, W. 2531 (A) *Pulau Tarakan, Kalimantan* — Othman Ismawi S29670 (E ex. SAR) *Kapit, Sarawak*.
- Paul Chai, P.P.K., Wright, E. S32334 (L) *Sungai Medamit, Limbang* — Sibat Luang S23622 (K ex. SAR), S23667 (A) *Arip River, Sarawak*; Sibat Luang S25238 (K) *Semengoh, Sarawak* — Sidek bin Kiah, Maulod bin Elin SK533 (SING) *Pulau Tioman, Pahang* — Sidiyasa, K., Ambriansyah, Zainal Ariffin 1296 (A) *East Kalimantan* — Sinclair, J., Kiah Hj. Salleh SFN40794 (E, L, SING) *Gunung Tebu Forest Reserve, Terengganu*.
- Teijsmann, J.E. (L ex. BO) *Pulau Bintan, Riau* — Vethevelu, P. FRI32926 (L ex. KEP) *Pasoh Forest, Negeri Sembilan* — Wallich, N. 4848 (E) *Singapore*; 4852D [Pierre4604] (P) *Prince of Wales, Malaysia* — Whitmore, T.C. FRI20282 (A, L, KEP), FRI20286 (L ex. KEP) *Trengan, Pahang*; FRI20347 (L) *Sungai Kerbat, Terengganu*; FRI8959 (L ex. KEP) *Sungai Loh, Terengganu* — Yeob 5839 (SING) *Johor, Malaysia* — Yii, P.C. S40775 (L ex. SAR) *Sabal Forest Reserve, Sarawak*; S53588 (SAR) *Batang Balui, Sarawak*.

**14. *G. rigida* Miq.**

Diepenhorst HB646 (L) *Priaman, Sumatra Barat* — Harmand 4012 (K) *Pulau Kondor, Vietnam* — Maskuri 193 (L) *Nias Island, Sumatra* — Poilane, M.E. 13735 (K) *Muang Saravane, Laos*; 17783 (K, P) *Cambodia*.

**15. *G. sangudsangud* Nazre**

A.Bakar, Meijer, W. SAN28681 (K, L) *Sungai Balung, Sabah* — Aban Gibot 60730 (L ex. SAN) *Ranau, Sabah*; SAN29564 (K, L, SAN), SAN32914 (L ex. SAN), SAN32977 (L, K) *Tawau, Sabah*; SAN30021 (K ex. SAN) *Quoin Hill, Tawau*; SAN34020 (L ex. SAN) *Merotai Besar, Sabah*.

Meijer, W. SAN22942 (L) *Sandakan, Sabah* — Muin Chai SAN29368 (K), SAN29720 (K, L, SAN), SAN33449 (K, L) *Lahad Datu, Sabah* — Wood, G.H.S., Wyatt-Smith, J. SANA4297 (L ex. SAN) *Mostyn estate, Lahad Datu*.

**16. *G. venulosa* (Blanco) Choisy**

Alvarez, R.J. 18513 (K) *Cagayan, Philippines* — Anon. 979 (L) *Arayat, Luzon* — Cuming, H. 1124 (K) — Curran, H.M. 10313 (K) *Tayabas, Luzon Island*; 17827 (E, K, L) *Cagayan, Philippines* — Edano, G. 5270 (A) *Nueva Ecija, Luzon Island* — Elmer, A.D.E. 18303 (L, UC) *Mount Makiling, Laguna*; 7308 (K) *Sorsogon, Luzon Island*; 8043 (E), 9151 (E) *Luchan, Luzon Island*; 8783 (E) *Baguio, Luzon Island*.

Fenix, E. 29968 (L) *Pangasinan* — Juliano, J.B. 23 (UC) *Laguna, Luzon Island* — Loher, A. 12655 (UC) *Montalban, Philippines*; 71 (K) *Luzon Island* — M. Azurin 29361 (A); 29361 (UC) *Isabela, Luzon Island* — Merrill, E.D. 4020 (K) *Tayabas, Luzon Island*

Quisumbing, E. 8065 (L) *Baler, Luzon Island* — Ramos, M. 33088 (L, SING) *Ilocos Norte, Luzon Island*; 77073 (SING) *Cagayan, Philippines* — Ramos, M., Edano, G. 47195 (SING) *Isabela, Luzon Island* — Teresita B. Banaga 33382 (L) *Mount Makiling, Laguna* — Vidal y Soler, S. 2110 (K), 2112 (K) *Manila, Philippines*; 2115 (K), 638 (K) *Tayabas, Luzon Island*.

## CHAPTER 7

### GENERAL CONCLUSIONS AND RECOMMENDATIONS

#### 7.1 Delimitation and Taxonomy of Section *Garcinia*

From a total of 43 species distributed in Madagascar, and India to Fiji, originally listed by Jones (1980) in sect. *Garcinia*, 19 species are reduced to synonyms, nine species are found not to belong in the section and five species do not have enough characters to classify them to a section. *Garcinia celebica* is found to have the most synonyms (13) while one species, *G. cataractalis*, previously unlisted in Jones's (1980) classification is newly included in sect. *Garcinia*. Five species, *G. acuticosta*, *G. discoidea*, *G. exiguus*, *G. ochraceus* and *G. sangudsangud*, and three varieties, *G. diospyrifolia* var. *arborea*, *G. diospyrifolia* var. *minor* and *G. malaccensis* var. *pseudomangostana* are newly described, which makes a total of 16 species in sect. *Garcinia* (see Appendix I for a full listing). The distribution of sect. *Garcinia* is now centred in SE Asia, stretching from eastern India to Papua New Guinea.

In sectional delimitation, important morphological characters are from the inflorescence, the orientation of the male flower, and from the fruit and leaves. Section *Garcinia* has a terminal inflorescence (except *G. moselleyana*) in clusters of 2-14 flowers, and the male flowers have 4-lobed or 4-angled stamen bundles with 2-theous anthers.

The fruits are berries, borne terminally, usually solitary, with a smooth and thick or thin wall when dry, and containing 2-12 seeds covered with a soft, edible translucent or white aril. The calyces are persistent and the stigmas are distinct with enough variation for species identification. For species circumscription, the inflorescence characters become less important, and characters from the male flower, fruit and leaf are most significant. Vegetative characters are usually sufficient for species identification.

## 7.2 Molecular Study of Sect. *Garcinia*

Parsimony and Bayesian analyses show that nuclear ITS data are more informative and provide better phylogenetic resolution in sect. *Garcinia* than the plastid *trnS-G* and *trnD-T* regions. Most of the sections of *Garcinia* delimited by Jones (1980) proved to be monophyletic but both ITS and plastid regions show that sect. *Garcinia* sensu Jones (1980) is not. This is because the clade consisting of *G. maingayi*, *G. costata* and *G. trianii* is nested outside of the clade of sect. *Garcinia* and is resolved as closely related to sect. *Brindonia* sensu Jones (1980). These three species differ from the species in sect. *Garcinia* and should be excluded from it on the basis of having thyrsoid inflorescences, stamen bundles without lobes, a ribbed fruit wall and stout secondary venation.

ITS phylogenetic trees allow sect. *Garcinia* to be divided into four groups of which all, except one of *G. nitida*, *G. cataractalis* and *G. diospyrifolia*, are monophyletic.

Characters that support these groups are the shape of stamen bundles, the shape of the fruit and stigma, and the fruit wall. Within sect. *Garcinia*, however, there is striking incongruence in the tree topology between the ITS and plastid trees. The main source of incongruence is due to the placement of *G. rigida*. In the ITS tree, *G. rigida* is nested within the sect. *Garcinia* clade but in the plastid trees, especially those based upon *trnS-G* data, *G. rigida* is placed outside sect. *Garcinia*. *Garcinia rigida* is believed to be of hybrid origin resulting in chloroplast capture, an introgression of the cytoplasmic genome of one species into the nuclear background of another. Plastid data from the species sampled are insufficient, however, to determine the maternal parent species of *G. rigida*.

Another species, *G. mangostana* (mangosteen), a widely cultivated fruit species in South East Asia that is thought to be of hybrid origin (Richards, 1990c), does not show any discordance between the placement of accessions in ITS and plastid trees. The phylogenies indicate that mangosteen arose from crosses between varieties of *G. malaccensis* if it is confirmed as an obligate agamosperm. However, if mangosteen is a facultative agamosperm, beside *G. malaccensis*, another *Garcinia* species could be involved in its parentage.

Most species of sect. *Garcinia* sampled in this study are distributed to the west side of Wallace's Line. Species from the eastern side of Wallace's Line are found nested within species from the west which suggests an origin on the western side of the line. Age estimation shows that the crown of *Garcinia* is 61.26 $\pm$ 1.58 MY while accessions of *G. rigida* from east of Wallace's Line split from their most common recent ancestor on the

west no later than 21.58 $\pm$ 2.90 MY. This is consistent with the theory of dispersal of species across this biogeographic division from west to east after the Sahul and Sunda shelves converged c. 20 MY ago (Morley, 1998).

### 7.3 Future Work and Recommendations

Future work is needed to clarify several areas of uncertainty raised by the research presented in this thesis:

1. The most morphologically heterogeneous and agamospermous species, *G. celebica*, needs additional study at the population level, especially from some islands (e.g. Andaman, Nicobar and Indonesia) and Indo-china where few collections have been made. Study of breeding systems (embryology, sex ratio, whether multiple seedlings emerge from a single seed) as outlined by Richards (1990a) needs to be carried out on these populations to determine if they are agamospermous, and if so whether this agamospermy is obligate or facultative. To investigate the correlation between breeding system and morphological variation, morphometric study could be carried out using herbarium specimens and compared with studies of genetic variation. Genotypic variation throughout a wide geographical range of *G. celebica* could be investigated by DNA fingerprinting (e.g. AFLP & RAF) techniques as used by Yapwattanaphun et al. (2003) and Ramage et al. (2004). Both AFLP and RAF might give sufficient intraspecific resolution to determine if morphologically distinct populations are genetically different, and therefore whether they represent distinct agamospermous lines.

2. Taxonomic placement of a group of species omitted from sect. *Garcinia* (*G. maingayi*, *G. trianaii* and *G. costata*), needs to be considered relative to sect. *Brindonia* (sensu Jones). It is not currently clear whether this clade of *G. maingayi*, *G. trianaii* and *G. costata* should stand as a new section or part of sect. *Brindonia*. To investigate this further, more detailed molecular phylogenetic study of sect. *Brindonia* (77 species) is needed with greater sampling of representative species. DNA sequence data from the regions used in this study of ITS, *trnS-G* and *trnD-T* could be used to see if the phylogenetic placement of the *G. maingayi*, *G. trianaii* and *G. costata* clade is within sect. *Brindonia*. If they nest within the clade of sect. *Brindonia*, they should be considered as part of the section. If the group is placed outside of sect. *Brindonia*, morphological investigation, especially focusing on type of inflorescence, shape of stamens and fruit characters, needs to be made to determine whether *G. maingayi*, *G. trianaii* and *G. costata* might be justifiably described as a new section.
3. Many more herbarium collections are needed to determine the placement of species not currently assigned to a section of *Garcinia*. Collection of specimens with male flowers is especially important. However, because finding male flowers is difficult, specimens with mature fruit could be sufficient because as proven in this study, fruit characters (e.g. fruit wall and stigma in *G. acutiscosta* and *G. cataractalis*) could give enough characters for sectional placement. Future field collection should prioritise poorly collected areas where unassigned species are found: Myanmar (*G. moulmienensis*), Kalimantan (*G. lucens*), the Philippines (*G.*

*blancoi*), and eastern and western parts of Papua (*G. squamata* and *G. erythrosperma*).

4. Any future molecular phylogenetic analysis in sect. *Garcinia* should sample more species with multiple accessions because it will give a clearer understanding of species relationships and possible hybrid origins. Apart from the ITS region, other nuclear markers such as the ribosomal external transcribed spacer (ETS), or perhaps low copy number nuclear genes (e.g. *PgiC*, *Adh*, GBSSI, G3PDH, PHY) that are less subject to concerted evolution, could be used to infer phylogeny because they may give additional phylogenetic resolution, and provide species-specific markers to enable the identification of parents of hybrid species. For plastid regions, additional potentially more variable markers such as *rpoB-trnC*, *trnT-trnL* and *trnS-trnfM* will allow better identification of the source of conflicting phylogenetic signal between plastid and nuclear phylogenetic trees as shown for example in *G. rigida*, and may provide species-specific markers to identify the maternal parents of hybrids.
5. Further research on breeding systems is required to confirm the nature of agamospermy in mangosteen throughout its range and especially in the Southeast Asian region. In order to do this, it will be necessary to investigate more detailed aspects of the embryology of populations, particularly of those from Borneo and east Indonesia because few studies have been carried out in this area. More field investigation in Southeast Asia is needed to locate male trees which will enable studies of pollination biology and the potential viability of the male flowers. Such

studies may indicate that outbreeding in mangosteen is occurring and that it is therefore not an obligate agamospermous species.

6. a) Undoubtedly the nature of agamospermy in mangosteen (as outlined in 5 above) is important to unravel its hybrid origin, but more data are needed especially on the ploidy level and the possible existence of cytological variation to understand its parentage. At present, cytological information of mangosteen has a range of published chromosome numbers from 56 to 96 (Chapter 3), but it is not certain whether this simply reflects difficulties in making cytological preparations of *Garcinia*'s tiny chromosomes. Establishing the true variation in chromosome number in mangosteen is important because although it is conceivable that an obligate agamosperm might have a variable chromosome number, such variation might also reflect multiple origins through hybridisation or perhaps a facultatively agamospermous breeding system.
- b) A more resolved phylogeny is needed to pinpoint parental species, and whether mangosteen has originated once or multiple times. These phylogenetic studies will need multiple mangosteen accessions, multiple samples of *G. malaccensis* that this study implicates as a possible parent, and other species closely related to mangosteen (*G. penangiana* and *G. venulosa*). It will also be necessary to sample other species of sect. *Garcinia* not included in this study (e.g. *G. exiguus*, *G. harmandii*, *G. moselleyana*, *G. ochraceus*, *G. sangudsangud* and *G. venulosa*) as any could be a parent of mangosteen. Sampling should concentrate in Malesia, particularly in Indo-china, Peninsular Malaysia, Sumatra, Borneo, Philippines, Sulawesi and Papua where these species are found.

c) More variable nuclear and plastid regions are needed to give species-specific markers to enable the identification of possible parental species in mangosteen. The markers indicated in (4) above could be variable enough, but fingerprinting techniques such as AFLPs might be another promising means of generating species-specific markers given their greater potential variability.

## BIBLIOGRAPHY

- Adams, C. D. 1970. Miscellaneous additions and revisions to the flowering plants of Jamaica. *Phytologia* 20 (5): 312.
- Adanson, M. 1763. *Familles des plantes* 2. Paris.
- Ahluwalia, V. K. & A. K. Tehim. 1984. Synthesis of some 1,3,6-trioxygenated isopentenylated xanthenes: Constitution of garcinone A. *Tetrahedron* 40(17): 3303-3312.
- Allem, A. C. 2004. Optimization theory in plant evolution: An overview of long-term evolutionary prospects in the angiosperms. *The Botanical Review* 69(3): 225-251.
- Anderson, T. 1874. Guttiferae. In: J. D. Hooker (ed.), *Flora of British India*: 258-278.
- APG 2003. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG II. *Botanical Journal of Linnean Society* 141: 399-436.
- Appanah, S. 1985. General flowering in the climax rain forests of South-East Asia. *Journal of Tropical Ecology* 1(3): 225-240.
- Appanah, S. & G. Weinland. 1993. A preliminary analysis of the 50-hectare Pasoh demography plot: I. Dipterocarpaceae. Research Pamphlet No. 112. Forest Research Institute of Malaysia, Kepong.
- Arnheim, N., M. Krystal, R. Schmickel, G. Wilson, O. Ryder & E. Zimmer. 1980. Molecular evidence for genetic exchanges among ribosomal genes on nonhomologous chromosomes in man and apes. *Proceedings of the National Academy of Sciences of the United States of America* 77(12): 7323-7327.
- Asai, F., H. Tosa, T. Tanaka & M. Iinuma. 1995. A xanthone from pericarps of *Garcinia mangostana*. *Phytochemistry* 39(4): 943-944.
- Ashton, P. S. 1988a. Manual of the non-Dipterocarp trees of Sarawak II. Dewan Bahasa dan Pustaka, Sarawak.
- Ashton, P. S. 1988b. Dipterocarp biology as a window to the understanding of tropical forest structure. *Annual Review of Ecology and Systematics* 19: 347-370.
- Ashton, P. S. 1988c. Systematics and ecology of rain forest trees. *Taxon* 37(3, Symposium Tropical Botany: Principles and Practice): 622-629.
- Asker, S. E. & L. Jerling. 1992. *Apomixis in plants*. CRC Press.
- Babcock, E. B. & G. L. Stebbins. 1938. The American species of *Crepis*: their relationships and distribution as affected by polyploidy and apomixis. Publication No. 504. Carnegie Institute, Washington.
- Backer, C. A. & R. C. J. Bakhuizen. 1963. *Flora of Java (Spermatophytes only)*. Vol.1. N.V. P. Noordhoff, Groningen.
- Baillon, H. 1877. *Histoire des plantes*. Volume 6 Libraire Hachette et Cie., Paris.
- Baldwin, B. G., M. J. Sanderson, J. M. Porter, M. F. Wojciechowski, C. S. Campbell & M. J. Donoghue. 1995. The ITS region of nuclear ribosomal DNA: A valuable source of evidence on angiosperm phylogeny. *Annals of the Missouri Botanical Garden* 82(2): 247-277.
- Bamps, P., N. Robson & B. Verdcourt. 1978. *Flora of tropical East Africa: Guttiferae*. Crown Agents for Overseas Government and Administration, London.

- Baum, D. A. 1992. Phylogenetic species concepts. *Trends in Ecology and Evolution* 7: 45-67.
- Bawa, K. S. 1980. Evolution of dioecy in flowering plants. *Annual Review of Ecology and Systematics* 11: 15-39.
- Bawa, K. S. 1998. Conservation of genetic resources in the Dipterocarpaceae. In: S. Appanah and J. S. Turnbull (eds.), *A review of Dipterocarps: Taxonomy, ecology and silviculture*: 45-56. CIFOR.
- Benthams, G. 1862. Guttiferae. In: G. Benthams and J. D. Hooker (eds.), *Genera Plantarum ad exemplaria imprimis in Herbariis Kewensibus servata definita* 1: 167-177.
- Berg, M. E. V. D. 1979. Revisao das especies brasileiras do genero *Rheedia* L. (Guttiferae). *Acta Amazonica* 9: 43-47.
- Blume, C. L. v. 1825. *Bijdragen tot de flora van Nederlandsch Indie*. Ter Lands Drukkeri, Batavia.
- Borhidi, A. 1980. New names and new species in the Flora of Cuba, II. *Acta Botanica Academicae Scientiarum Hungaricae* 26 (3-4): 255-275.
- Burkill, I.H. 1935. *Dictionary of economic products of the Malay Peninsula* 1. Governments of the Straits Settlements and Federated Malay States.
- Carman, J. G. 1997. Asynchronous expression of duplicate genes in angiosperms may cause apomixis, bispority, tetraspority, and polyembryony. *Biological Journal of the Linnaean Society*. 61: 51-94.
- Chairungsrilerd, N., K. Furukawa, T. Ohta, S. Nozoe & Y. Ohizumi. 1996. Histaminergic and serotonergic receptor blocking substances from the medicinal plant *Garcinia mangostana*. *Planta Medica* 62(5): 471-472.
- Chan, H. T. 1981. Reproductive biology of some Malaysian dipterocarps, III. Breeding systems. *Malaysian Forester* 44: 28-36.
- Chen, S. X., M. Wan & B. N. Loh. 1996. Active constituents against HIV-1 protease from *Garcinia mangostana*. *Planta Medica* 62(4): 381-382.
- Choisy, J. D. 1823. Mémoire sur un nouveau genre de Guttifères, et sur l'arrangement methodique de cette famille. *Mémoires de la Société d'Histoire Naturelle de Paris* 1(1): 210-232.
- Choisy, J. D. 1824. Guttiferae. In: A. P. D. Candolle (ed.), *Prodromus systematis naturalis regni vegetabilis*: 557-564. Treutel & Wurtz, Paris.
- Choisy, J. D. 1851. Description des Guttiferes de l'Inde. *Mémoires de la Société de Physique et d'Histoire Naturelle de Geneve* 12: 381-440.
- Chomnawang, M. T., S. Surassmo, V. S. Nukoolkarn & W. Gritsanapan. 2005. Antimicrobial effects of Thai medicinal plants against acne-inducing bacteria. *Journal of Ethnopharmacology* 101(1-3): 330-333.
- Clausen, J. 1954. Partial apomixis as an equilibrium system in evolution. *Caryologia* (Vol. Suppl.): 469-479.
- Corlett, R. T. 1993. Reproductive phenology of Hong Kong shrubland. *Journal of Tropical Ecology* 9(4): 501-510.
- Cracraft, J. 2000. Species concepts in theoretical and applied biology: A systematic debate with consequences. In: Q. D. Wheeler and R. Meier (eds.), *Species concepts and phylogenetic theory: a debate*: 3-14. Columbia University Press, New York

- Craib, W. G. 1931. *Florae Siamensis enumeratio*. Volume 1 Polypetalae. Siam Society, Bangkok.
- Crepet, W. L. & K. C. Nixon. 1998. Fossil Clusiaceae from the Late Cretaceous (Turonian) of New Jersey and implications regarding the history of bee pollination. *American Journal of Botany* 85(8): 1122-1133.
- Cronquist, A. 1978. Once again, what is species? In: J. A. Romberger (ed.), *Biosystematic in agriculture*: 3-20. Allenheld, Osmun & Co., New Jersey.
- Cruz, F. S. D. J. 2001. Status report on genetic resources of mangosteen (*Garcinia mangostana* L.) in Southeast Asia. IPGRI.
- Cunningham, C. W. 1997. Is congruence between data partitions a reliable predictor of phylogenetic accuracy? Empirically testing an iterative procedure for choosing among phylogenetic methods. *Systematic Biology* 46: 464-478.
- Davis, P. H. & V. H. Heywood. 1963. *Principles of angiosperm taxonomy*. Oliver & Boyd, Edinburgh and London.
- Demesure, B., N. Sodji & R. J. Petit. 1995. A set of universal primers for amplification of polymorphic non-coding regions of mitochondrial and chloroplast DNA in plants. *Molecular Ecology* 4: 129-131.
- Davis, J. I. & K. C. Nixon 1992. Populations, genetic variation, and the delimitation of phylogenetic species. *Systematic Biology* 41(4): 421-435.
- Doyle, J. J. & J. L. Doyle. 1990. Isolation of plant DNA from fresh tissue. *Focus* 12: 13-15.
- Doyle, J. J., J. L. Doyle & A. H. Brown. 1999. Incongruence in the diploid B-genome species complex of *Glycine* (Leguminosae) revisited: histone H3-D alleles versus chloroplast haplotypes. *Molecular Biology and Evolution* 16(3): 354-362.
- Engler, A. 1893. Guttiferae. In: A. Engler and K. A. E. Prantl (eds.), *Natürlichen Pflanzenfamilien*: 194-242. Engelman, Leipzig.
- Engler, A. 1925. Guttiferae. In: A. Engler and K. A. E. Prantl (eds.), *Natürlichen Pflanzenfamilien* 2nd. ed.: 154-237. Leipzig.
- Farris, J. D., M. Källersjö, A. G. Kluge & C. Bult. 1994. Testing the significance of incongruence. *Cladistics* 10: 315-319.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39: 783-791.
- Fitch, W. M. 1971. Toward defining the course of evolution: minimum change for specific tree topology. *Systematic Zoology* 20: 406-416.
- Ford, V. S. & L. D. Gottlieb. 1999. Molecular characterization of PgiC in a tetraploid plant and its diploid relatives. *Evolution* 53: 1060-1067.
- Friesen, N., R. M. Fritsch, S. Pollner & F. R. Blattner. 2000. Molecular and morphological evidence for an origin of the aberrant genus *Milula* within the Himalayan species of *Allium* (Alliaceae). *Molecular Phylogenetics and Evolution* 17: 209-218.
- Fuertes Aguilar, J., J. A. Rossello & G. Nieto Feliner. 1999. Nuclear ribosomal DNA (nrDNA) concerted evolution in natural and artificial hybrids of *Armeria* (Plumbaginaceae). *Molecular Ecology* 8(8): 1341-1346.

- Garcin, L. 1733. The settling of a new genus of plants, called after the Malayans, mangostans; By Laurentius Garcin, M. D. and F. R. S. Translated from the French by Mr. Zollman, F. R. S. *Philosophical Transactions* 38: 232-242.
- Ge, S., T. Sang, B.-R. Lu & D.-Y. Hong. 1999. Phylogeny of rice genomes with emphasis on origins of allotetraploid species. *Proceedings of the National Academy of Sciences of the United States of America* 96(25): 14400-14405.
- Goh, S. H., I. Jantan, A. I. Gray & P. G. Waterman. 1992. Prenylated xanthenes from *Garcinia opaca*. *Phytochemistry* 31(4): 1383-1386.
- Gopalakrishnan, C., D. Shankaranarayanan, L. Kameswaran & S. K. Nazimudeen. 1980. Effect of mangostin, a xanthone from *Garcinia mangostana* L. in immunopathological & inflammatory reactions. *Indian Journal of Experimental Biology* 18(8): 843-846.
- Gopalakrishnan, G. & B. Balaganesan. 2000. Two novel xanthenes from *Garcinia mangostana*. *Fitoterapia* 71(5): 607-609.
- Gopalakrishnan, G., B. Banumathi & G. Suresh. 1997. Evaluation of the antifungal activity of natural xanthenes from *Garcinia mangostana* and their synthetic derivatives. *Journal of Natural Products* 60(5): 519-524.
- Govindachari, T. R., P. S. Kalyanaraman, N. Muthukumaraswamy & B. R. Pai. 1971. Xanthenes of *Garcinia mangostana* L. *Tetrahedron* 27(16): 3919-3926.
- Graham, R. 1837. Remarks on the gamboge tree of Ceylon and the character of *Hebradendron*, a new genus of the Guttiferae and that to which the tree belongs. *Companion to the Botanical Magazine* 2 (19): 193-200.
- Grant, V. 1971. *Plant speciation*. Columbia University Press, New York & London.
- Gustafson, A. 1946-1947. Apomixis in higher plant. *Lunds Universitets Arsskrift Avdelningen* 42-43: 1-370.
- Gustafsson, M. H. G., V. Bittrich & P. F. Stevens. 2002. Phylogeny of Clusiaceae based on *rbcL* sequences. *International Journal of Plant Sciences* 163 (3): 1045-1054.
- Ha, C. O., V. E. Sands, E. Soepadmo & K. Jong. 1988. Reproductive patterns of selected understorey trees in Malaysian rain forest: the apomitic species. *Botanical Journal of Linnaean Society*. 97: 317-331.
- Hahn, W. J. 2002. A phylogenetic analysis of the Arecoideae line of palms based on plastid DNA sequence data. *Molecular Phylogenetics and Evolution* 23: 189-204.
- Hall, T. A. 1999. Bioedit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95-98.
- Hamilton, M. B., J. M. Braverman & D. F. Soria-Hernanz. 2003. Pattern and relative rate of nucleotides and insertion/deletion evolution at six chloroplast intergenic regions in New World species of the Lecythidaceae. *Molecular Biology and Evolution* 20: 1710-1721.
- Hammel, B. 1989. New combinations and taxonomies in Clusiaceae. *Annals of the Missouri Botanical Garden* 76(3): 927-929.
- Hardig, T. M., P. S. Soltis & D. E. Soltis 2000. Diversification of the North American shrub genus *Ceanothus* (Rhamnaceae): conflicting phylogenies from nuclear ribosomal DNA and chloroplast DNA. *American Journal of Botany* 87(1): 108-123.

- Harris, J. G. & M. W. Harris. 2004. Plant identification terminology. An illustrated glossary. 2nd Edition. Spring Lake Publishing, Utah.
- Hasskarl, J. K. 1844. Catalago plantarum in Horto Bogoriensi. Lands Drukkerij, Batavia.
- Hennig, W. 1966. Phylogenetic systematic. University of Illinois Press, Urbana.
- Hickey, L. J. 1979. A revised classification of the architecture of dicotyledonous leaves. In: C. R. Metcalfe & L. Chalk (eds.), Anatomy of the dicotyledons: 25-39. Clarendon Press, Oxford.
- Hijmans, R. J., L. Guarino, M. Cruz & E. Rojas. 2001. Computer tools for spatial analysis of plant genetic resource data: 1. DIVA-GIS. Plant Genetic Resource Newsletter No. 127: 15-19.
- Ho, C. K., Y. L. Huang & C. C. Chen. 2002. Garcinone E, a xanthone derivative, has potent cytotoxic effect against hepatocellular carcinoma cell lines. *Planta Medica* 68(11): 975-979.
- Hooker, J. D. 1875. *Garcinia*. In: J. D. Hooker (ed.), Flora of British India 1: 259. L. Reeve & Co., London.
- Huang, Y. L., C. C. Chen, Y. J. Chen, R. L. Huang & B. J. Shieh. 2001. Three xanthenes and a benzophenone from *Garcinia mangostana*. *Journal of Natural Products* 64(7): 903-906.
- Huelsensbeck, J. P., B. Larget, R. E. Miller & R. Ronquist. 2002. Potential applications and pitfalls of Bayesian inference of phylogeny. *Systematic Biology* 51(5): 673-688.
- Hume, E. P. & M. Cobin. 1946. Relation of seed size to germination and early growth of mangosteen. *Proceedings of the American Horticultural Science* 48: 293-302.
- Huxley, T. H. 1868. On the classification and distribution of the Alectoromorphae and Heteromorphae. *Proceedings of the Zoological Society of London*: 294-319.
- Idris, S. & A. Rukayah. 1987. Description of male mangosteen (*Garcinia mangostana* L.) discovered in Peninsular Malaysia. *Mardi Research Bulletin* 15(1): 63-66.
- Iinuma, M., H. Tosa, T. Tanaka, S. Kanamaru, F. Asai, Y. Kobayashi, K. Miyauchi & R. Shimano. 1996. Antibacterial activity of some *Garcinia* benzophenone derivatives against methicillin-resistant *Staphylococcus aureus*. *Biological and Pharmaceutical Bulletin* 19(2): 311-314.
- Jones, S. W. 1980. Morphology and major taxonomy of *Garcinia* (Guttiferae). Ph. D. Thesis. University of Leicester. Unpublished.
- Jung, H. A., B. N. Su, W. J. Keller, R. G. Mehta & A. D. Kinghorn. 2006. Antioxidant xanthenes from the pericarp of *Garcinia mangostana* (Mangosteen). *Journal of Agricultural and Food Chemistry* 54(6): 2077-2082.
- Jussieu, A. L. 1789. *Genera plantarum*. Viduam Herissant. Paris.
- K-Sen, A., K. K. Sarkar, P. C. Mazumder, N. Banerji, R. Uusvuori & T. A. Haset. 1980. A xanthone from *Garcinia mangostana*. *Phytochemistry* 19(10): 2223-2225.
- Kaur, A., C. O. Ha, K. Jong, V. E. Sands, H. T. Chan, E. Soepadmo & P. S. Ashton. 1978. Apomixis may be widespread among trees of the climax rain forest. *Nature* 271: 440-442.
- Kenicer, G. J., T. Kajita, R. T. Pennington & J. Murata. 2005. Systematics and biogeography of *Lathyrus* (Leguminosae) based on internal transcribed spacer and cpDNA sequence data. *American Journal of Botany* 92: 1199-1209.

- King, G. 1891. Materials for flora of Malay Peninsular. Journal of the Asiatic Society of Bengal, Part 2, Natural History 60 (2, No. 1): 147-249.
- Koch, M. A., C. Dobes & T. Mitchell-Olds. 2003. Multiple hybrid formation in natural populations: Concerted evolution of the internal transcribed spacer of nuclear ribosomal DNA (ITS) in North American *Arabidopsis thaliana* (Brassicaceae). Molecular Biology and Evolution 20(3): 338-350.
- Kochummen, K. M. 1997. Tree flora of Pasoh. Forest Research Institute, Kepong.
- Kochummen, K. M., J. V. LaFrankie & N. Manokaran. 1990. Floristic composition of Pasoh Forest Reserve, a lowland rain forest in Peninsular Malaysia. Journal of Tropical Forest Science 3: 1-13.
- Koorders, S. H. & T. Valeton. 1903. Tot de kennis der boomsoorten op Java. Bijdrage No.9. G. Kolff & Co., Batavia.
- Kostermans, A. J. G. H. 1962. Miscellaneous botanical notes 3. Reinwardtia 6(2): 155-187.
- Kostermans, A. J. G. H. 1976. Notes on the Clusiaceae of Sri Lanka and reductions of *Pentapthalangium* Warb. Ceylon Journal of Science. (Biological Science) 12: 55-71.
- Kovarik, A., J. C. Pires, A. R. Leitch, K. Y. Lim, A. M. Sherwood, R. Matyasek, J. Rocca, D. E. Soltis & P. S. Soltis. 2005. Rapid concerted evolution of nuclear ribosomal DNA in two *Tragopogon* allopolyploids of recent and recurrent origin. Genetics 169(2): 931-944.
- Krishnaswamy, N. & V. S. Raman 1949. A note on the chromosomes numbers of some economic plants of India. Current Science 18: 376-378.
- Kurz, W. S. 1870. On some new or imperfectly known Indian plants. Journal of the Asiatic Society of Bengal. Part 2 no. 2, 39: 61-90.
- Kurz, W. S. 1877. Forest flora of Burma I. Office of the Superintendent of Government Printing, Calcutta.
- Kyndt, T., B. Van Droogenbroeck, E. Romeijn-Peeters, J. P. Romero-Motochi, X. Scheldeman, P. Goetghebeur, P. V. Damme & G. Gheysen. 2005. Molecular phylogeny and evolution of Caricaceae based on rDNA internal transcribed spacers and chloroplast sequence data. Molecular Phylogenetics and Evolution 37(2): 442-459.
- Lambert, F. & A. G. Marshall. 1991. Keystone characteristics of bird-dispersed *Ficus* in Malaysian lowland rain forest. Journal of Ecology 79: 793-809.
- Lanessan, J. M. A. 1872. Memoire sur le genre *Garcinia* (Clusiacees) et sur l'origine et les proprietes de la gomme-gutte. J.-B. Bailliere et fils. Paris.
- Lee, H. S., P. S. Ashton, T. Yamakura, S. Tan, S. J. Davies, A. Itoh, E. O. K. Chai, T. Ohkubo & J. V. LaFrankie. 2002. The 52-ha forest research plot at Lambir Hills, Sarawak, Malaysia: tree distribution maps, diameter tables and species documentation. Forest Department Sarawak, The Arnold Arboretum-CTFS Asia Program and The Smithsonian Tropical Research Institute.
- Leighton, M. & D. R. Leighton. 1983. Vertebrate response to fruiting seasonality within a Bornean rain forest. In: S. L. Sutton, T. C. Whitmore and A. C. Chadwick (eds.), Tropical Rain Forest: Ecology and Management: 181-196. Blackwells, Oxford.

- Lim, A. L. 1984. The embryology of *Garcinia mangostana* (Clusiaceae). Gardens' Bulletin Singapore 37(1): 93-104.
- Linnaeus, C. 1753. Species Plantarum 1. Stockholm.
- Linnaeus, C. 1754. Herbarium Amboinense. In, amoenitates academici seu dissertationes 2. Exc. L.M. Hojer, Upsaliae.
- Linnaeus, C. 1774. Systema vegetabilium secundum classes ordines genera species cum characteribus et differentiis. Gottingae.
- Liogier, A. H. 1986. Novitates antillanae XII. Phytologia 61 (6): 353-360.
- Lorenz-Lemke, A. P., V. C. Muschner, S. L. Bonatto, A. C. Cervi, F. M. Salzano & L. B. Freitas. 2005. Phylogeographic inferences concerning evolution of Brazilian *Passiflora actinia* and *P. elegans* (Passifloraceae) based on ITS (nrDNA) variation. Annals of Botany 95(5): 799-806.
- Loureiro, J. D. 1890. Flora Cochinchinensis. Typis, et Expensis Academicis, Lisbon.
- Love, A. 1960. Biosystematics and classification of apomictics. Feddes Repertorium 63: 136.
- Luckow, M. 1995. Species concepts: Assumptions, methods, and applications. Systematic Botany 20(4): 589-605.
- Lughadha, E. N. & C. Proenca. 1996. Survey of reproductive biology of the Myrtoideae (Myrtaceae). Annals of the Missouri Botanical Garden 83(4): 480-503.
- Maheshwari, J. K. 1960. Taxonomic study of Indian Guttiferae III. The genus *Garcinia* L. s.l. Bulletin of the Botanical Survey of India 6: 107-135.
- Mason-Gamer, R. J. 2001. Origin of North American *Elymus* (Poaceae: Triticeae) allotetraploids based on granule-bound starch synthase gene sequences. Systematic Botany 26(4): 757-768.
- Matsumoto, K., Y. Akao, E. Kobayashi, K. Ohguchi, T. Ito, T. Tanaka, M. Iinuma & Y. Nozawa. 2003. Induction of apoptosis by xanthones from mangosteen in human leukaemia cell lines. Journal of Natural Products 66(8): 1124-1127.
- Mayr, E. 1970. Populations, species, and evolution. The Belknap Press of Harvard Univ. Press, Massachusetts.
- Merrill, E. D. 1908. The Philippines species of *Garcinia*. Philippine Journal of Science. Section C, Botany 3: 359-368.
- Merrill, E. D. 1923. An enumeration of Philippine flowering plants. Volume 3. Bureau of Printing, Manila.
- Mes, T. H. M., J. Van Brederode & H. T. Hart. 1996. Origin of the woody Macaronesian Sempervivoideae and the phylogenetic position of the east African species of *Aeonium*. Botanica Acta 109: 441-506.
- Metcalfe, I. 1998. Palaeozoic and Mesozoic geological evolution of the SE Asian region: multidisciplinary constraints and implications for biogeography. In: R. Hall and J. D. Holloway (eds.), Biogeography and geological evolution of SE Asia: 25-41. Backhuys Publishers, Leiden.
- Miquel, F. A. W. 1859. Flora Indiae Batavae. Vol.1, Part 2. C.G. van der Post, Amsterdam.
- Miquel, F. A. W. 1860. Flora Indiae Batavae. Supplementum primum : prodromus florae Sumatranæ. C.G. van der Post, Amsterdam.

- Moongkarndi, P., N. Kosem, S. Kaslungka, O. Luanratana, N. Pongpan & N. Neungton. 2004a. Antiproliferation, antioxidation and induction of apoptosis by *Garcinia mangostana* (mangosteen) on SKBR3 human breast cancer cell line. *Journal of Ethnopharmacology* 90(1): 161-166.
- Moongkarndi, P., N. Kosem, O. Luanratana, S. Jongsomboonkusol & N. Pongpan. 2004b. Antiproliferative activity of Thai medicinal plant extracts on human breast adenocarcinoma cell lines. *Fitoterapia* 75(3-4): 375-377.
- Morley, R. J. 1998. Palynological evidence for Tertiary plant dispersals in the SE Asian region in relation to plate tectonics and climate. In: R. Hall and J. D. Holloway (eds.), *Biogeography and Geological Evolution of SE Asia*: 211-234. Backhuys Publishers, Leiden.
- Morley, R. J. 2000. *Origin and evolution of tropical rain forest*. John Wiley & Sons, Ltd., Sussex.
- Murray, A. J. 1789. *Commentatio de arboribus gummi-guttae fundentibus. Commentationes Societatis Regiae Scientiarum Gottingensis*. Gottingen 9: 169-184.
- Nabandith, V., M. Suzui, T. Morioka, T. Kaneshiro, T. Kinjo, K. Matsumoto, Y. Akao, M. Iinuma & N. Yoshimi. 2004. Inhibitory effects of crude alpha-mangostin, a xanthone derivative, on two different categories of colon preneoplastic lesions induced by 1, 2-dimethylhydrazine in the rat. *Asian Pacific Journal of Cancer Prevention* 5(4): 433-438.
- Nakatani, K., M. Atsumi, T. Arakawa, K. Oosawa, S. Shimura, N. Nakahata & Y. Ohizumi. 2002. Inhibitions of histamine release and prostaglandin E2 synthesis by mangosteen, a Thai medicinal plant. *Biological and Pharmaceutical Bulletin* 25(9): 1137-1141.
- Nazre, M. 2000. *The ecology and taxonomy of Garcinia (F. Guttiferae) in 50-ha plot of Pasoh Forest Reserve*. Unpublished M. Sc. Thesis. Botany Department, Universiti Kebangsaan Malaysia.
- Ng, F. S. P. 1992. *Manual of forest fruits, seeds and seedlings*. Malayan Forest Record No. 34. Forest Research Institute Malaysia, Kepong.
- Ng, K. K. S., S. L. Lee & C. L. Koh. 2004. Spatial structure and genetic diversity of two tropical tree species with contrasting breeding systems and different ploidy levels. *Molecular Ecology* 13(3): 657-669.
- Nilar & L. J. Harrison. 2002. Xanthenes from the heartwood of *Garcinia mangostana*. *Phytochemistry* 60(5): 541-548.
- Nilar, L.-H. D. Nguyen, G. Venkatraman, K.-Y. Sim & L. J. Harrison. 2005. Xanthenes and benzophenones from *Garcinia griffithii* and *Garcinia mangostana*. *Phytochemistry* 66(14): 1718-1723.
- Nixon, K. C. & Q. D. Wheeler 1990. An amplification of the phylogenetic species concept. *Cladistics* 6: 211-223.
- Normah, M. N. 2000. In vitro techniques for multiplication and conservation of *Garcinia mangostana*, *Lansium domesticum* and *Baccaurea polyneura*. *Agro-food Industry Hi-Tech* 11 (4): 2-4
- Nylander, J. A. A. 2004. MrModeltest v2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.

- Oliver, D. 1868. Flora of tropical Africa. Vol.1 : Ranunculaceae to Connaraceae. L. Reeve & Co., London.
- Olson, M. E. 2002. Combining data from DNA sequences and morphology for phylogeny of Moringaceae (Brassicales). *Systematic Botany* 27: 55-73.
- Osario, D. & M. Vorobyev. 1996. Colour vision as an adaptation to frugivory in primates. *Proceedings of the Royal Society of London B* 263: 593-599.
- Page, R. D. M. 1996. TREEVIEW: An application to display phylogenetic trees on personal computers. *Computer Applications in the Biosciences* 12: 357-358.
- Palmer, J. D., R. K. Jansen, H. J. Micheals, M. W. Chase & J. R. Manhart. 1998. Chloroplast DNA variation and plant phylogeny. *Annals of the Missouri Botanical Garden* 75: 1180-1206.
- Parveen, M. & N. U.-D. Khan. 1988. Two xanthenes from *Garcinia mangostana*. *Phytochemistry* 27(11): 3694-3696.
- Parveen, M., N. U.-D. Khan, B. Achari & P. K. Dutta. 1991. A triterpene from *Garcinia mangostana*. *Phytochemistry* 30(1): 361-362.
- Pham-Hoang, H. 1991. Cay co Viet Nam: an illustrated flora of Vietnam Quyen I. Nha Xuat Ban Tre.
- Pierre, J. B. L. 1882-1885. Flore forestiere de la Cochinchine 1-2. pl. 54-98. Paris.
- Pitard, L. J. 1910. Guttiferae. In: M. H. Lecomte (ed.), *Flora generale de l' Indochine* 1: 292-330. Paris.
- Planchon, J. E. & J. Triana. 1860. *Garcinia*. *Annales des Sciences Naturelles Botanique*: 169-206.
- Ramage, C. M., L. Sando, C.P. Peace, B.J. Carroll & R.A. Drew. 2004. Genetic diversity revealed in the apomictic fruit species *Garcinia mangostana* L. (mangosteen). *Euphytica* 136: 1-10.
- Renner, S. S. & J. P. Feil. 1993. Pollinators of Tropical Dioecious Angiosperms. *American Journal of Botany* 80(9): 1100-1107.
- Richards, A. J. 1983. *Plant Breeding Systems*. George Allen & Unwin, London.
- Richards, A. J. 1990a. Studies in *Garcinia*, dioecious tropical forest trees: agamospermy. *Botanical Journal of the Linnean Society* 103: 233-250.
- Richards, A. J. 1990b. Studies in *Garcinia*, dioecious tropical forest trees: the phenology, pollination biology and fertilization of *G. hombroniana* Pierre. *Botanical Journal of the Linnean Society* 103: 251-261.
- Richards, A. J. 1990c. Studies in *Garcinia*, dioecious tropical forest trees: the origin of mangosteen. *Botanical Journal of the Linnean Society* 103: 301-308.
- Richards, A. J. 2003. Apomixis in flowering plants: an overview. *Philosophical Transactions of the Royal Society B: Biological Science* 358: 1085-1093.
- Richardson, J. E. , M. F. Fay, Q. C. Cronk, D. Bowman, M. W. A. Chase. 2000. A phylogenetic analysis of Rhamnaceae using *rbcL* and *trnL-F* plastid DNA sequences. *American Journal of Botany* 87(9):1309-1324.
- Richardson, J. E., R. T. Pennington, T. D. Pennington & P. M. Hollingsworth. 2001a. Rapid diversification of species-rich genus of Neotropical rain forest trees. *Science* 293: 2242-2245.

- Richardson, J. E., F. M. Weitz, M. F. Fay, Q. C. B. Cronk, H. P. Linder, G. Reeves & M. W. Chase. 2001b. Rapid and recent origin of species richness in the Cape Flora of South Africa. *Nature* 413: 181-183.
- Ridley, H. N. 1922. *Flora of Malay Peninsula 1*. Lovell Reeve, London.
- Rismita-Sari 2000. Review of *Garcinia* (Clusiaceae) Based on Molecular Systematics. A Phylogenetic study of molecular data of *Garcinia* spp. Unpublished M. Sc. Thesis. Department of Tropical Plant Science, School of Tropical Biology., James Cook University.
- Robson, N. K. B. 1958. New and little-known species from Flora Zambesiaca area VI. *Boletim da Sociedade Broteriana Series 2*, 32: 151-173.
- Robson, N. K. B. 1961. Guttiferae. In: A. W. Exell and H. Wild (eds.), *Flora Zambesiaca. Volume 1: Mozambique, Federation of Rhodesia and Nyasaland, Bechuanaland Protectorate* 378-404. Crown Agents for Overseas Government, London.
- Ronquist, F. & J. P. Huelsenbeck. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572-1574.
- Ronsted, N., M. W. Chase, D. C. Albach & M. A. Bello. 2002. Phylogenetic relationships within *Plantago* (Plantaginaceae): evidence from nuclear ribosomal ITS and plastid trnL-F sequence data. *Botanical Journal of Linnean Society*. 139(4): 323-338.
- Rosen, D. E. 1978. Vicariant patterns and historical explanation in biogeography. *Systematic Zoology* 27:159-188.
- Rosen, D. E. 1979. Fishes from the uplands and intermontane basins of Guatemala: Revisionary studies and comparative geography. *Bulletin of the American Museum of Natural History* 162: 267-375.
- Roxburgh, W. 1798. *Plants of the Coast of Coromandel. Vol.2*.
- Roxburgh, W. 1814. *Hortus Bengalensis*. Calcutta.
- Rozas, J. & R. Rozas 1999. DNASP. *Bioinformatics* 15: 174-175.
- Rukachaisirikul, V., A. Adair, P. Dampawan, W. C. Taylor & P. C. Turner. 2000. Lanostanes and friedolanostanes from the pericarp of *Garcinia hombroniana*. *Phytochemistry* 55(2): 183-188.
- Rukachaisirikul, V., P. Pailee, A. Hiranrat, P. Tuchinda, C. Yoosook, J. Kasisit, W. C. Taylor & V. Reutrakul. 2003. Anti-HIV-1 protostane triterpenes and digeranylbenzophenone from trunk, bark and stems of *Garcinia speciosa*. *Planta Medica* 69(12): 1141-1146.
- Rukachaisirikul, V., S. Saelim, P. Karnsomchoke & S. Phongpaichit. 2005. Friedolanostanes and lanostanes from the leaves of *Garcinia hombroniana*. *Journal of Natural Products* 68(8): 1222-1225.
- Rumphius, G. E. 1741. *Herbarium Amboinenese, purimas complectens arbores, fructices, herbas, plantas terrestres & aquaticas, quae in Amboina ed adjacentibus Reperiuntur Insulia 1*. Amsterdam.
- Sakagami, Y., M. Iinuma, K. G. Piyasena & H. R. Dharmaratne. 2005. Antibacterial activity of alpha-mangostin against vancomycin resistant *Enterococci* (VRE) and synergism with antibiotics. *Phytomedicine* 12(3): 203-208.
- Sanderson, M. J. 1997. A nonparametric approach to estimating divergence times in the absence of rate constancy. *Molecular Biology and Evolution* 14(12): 1218-1231.

- Sanderson, M. J. 2003. r8s; inferring absolute rates of evolution and divergence times in the absence of a molecular clock. *Bioinformatics* 19: 301-302.
- Sang, T. & D. Zhang. 1999. Reconstructing hybrid speciation using sequences of low copy nuclear genes: hybrid origins of five *Paeonia* species based on *Adh* gene phylogenies. *Systematic Botany* 24(2): 148-163.
- Schumann, K. & M. U. Hollrung. 1889. *Flora von Kaiser Wilhelms Land*. Berlin.
- Shankaranarayan, D., C. Gopalakrishnan & L. Kameswaran 1979. Pharmacological profile of mangostin and its derivatives. *Archives Internationales de Pharmacodynamie et de Therapie* 239(2): 257-269.
- Shaw, J., E. B. Lickey, J. T. Beck, S. B. Farmer, W. Liu, J. Miller, K. C. Siripun, C. T. Winder, E. E. Schilling & R. L. Small. 2005. The tortoise and the hare II: Relative utility of 21 noncoding chloroplast DNA sequences for phylogenetic analysis. *American Journal of Botany* 92(1): 142-166.
- Simmons, M. P. & H. Ochoterena. 2000. Gaps as characters in sequence-based phylogenetic analyses. *Systematic Biology* 49: 369-381.
- Small, R. L., R. C. Cronn & J. F. Wendel. 2004. Use of nuclear genes for phylogeny reconstruction in plants. *Australian Systematic Botany* 17: 145-170.
- Soepadmo, E. 1989. Contribution of reproductive biological studies towards the conservation and development of Malaysian plant genetic resources. In: A. H. Zakri (ed.) *Genetic Resources of Under-utilised Plants in Malaysia*. 1-41. Malaysian National Committee of Plant Genetic Resources. Kuala Lumpur
- Soltis, D. E. & R. K. Kuzoff. 1995. Discordance between Nuclear and Chloroplast Phylogenies in the *Heuchera* Group (Saxifragaceae). *Evolution* 49(4): 727-742.
- Soltis, D. E., P. S. Soltis, T. G. Collier & M. L. Edgerton. 1991. Variation within and among genera of the *Heuchera* group: evidence of chloroplast capture and paraphyly. *American Journal of Botany* 78: 1091-1112.
- Sprecher, M. A. 1919. Étude sur la semence et la germination du *Garcinia mangostana* L. *Revue Generale de Botanique* 31: 513-531.
- Stace, C. A. 1998. Species recognition in agamosperms - the need for a pragmatic approach. *Folia Geobotanica* 33: 319-326.
- Stevens, P. F. 2001 onwards. Angiosperm Phylogeny Website. Version 6. <http://www.mobot.org/MOBOT/research/APweb/>. May 2006.
- Suh, Y., L. B. Thien, H. E. Reeve & E. A. Zimmer. 1993. Molecular evolution and phylogenetic implications of internal transcribed spacer sequences of ribosomal DNA in Winteraceae. *American Journal of Botany* 80(9): 1042-1055.
- Suksamrarn, S., N. Suwannapoch, W. Phakhodee, J. Thanuhiranlert, P. Ratananukul, N. Chimnoi & A. Suksamrarn. 2003. Antimycobacterial activity of prenylated xanthenes from the fruits of *Garcinia mangostana*. *Chemical and Pharmaceutical Bulletin* 51(7): 857-859.
- Suksamrarn, S., N. Suwannapoch, P. Ratananukul, N. Aroonlerk & A. Suksamrarn. 2002. Xanthenes from the green fruit hulls of *Garcinia mangostana*. *Journal of Natural Products* 65(5): 761-763.
- Sundaram, B. M., C. Gopalakrishnan, S. Subramanian, D. Shankaranarayanan & L. Kameswaran. 1983. Antimicrobial activities of *Garcinia mangostana*. *Planta Medica* 48(1): 59-60.

- Swofford, D. L. 2003 PAUP\*. Phylogenetic Analysis using Parsimony (\*And Other Methods) Version 4. Sinauer Associates, Massachusetts.
- Swofford, D. L. & G. J. Olsen. 1990. Phylogeny reconstruction. In: D. M. Hillis and C. Moritz (eds.), *Molecular Systematics*: 411-501. Sinauer Associates, Massachusetts.
- Thomas, S. C. 1997. Geographic parthenogenesis in a tropical forest tree. *American Journal of Botany* 84(7): 1012-1015.
- Thompson, J. D., T. J. Gibson, F. Plewniak, F. Jeanmougin & D. G. Higgins. 1997. The CLUSTAL X Windows: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acid Research* 25(4): 4876-4882.
- Thouars, L. M. A. D. P. 1805. *Brindonia*. In: G. F. Cuvier (ed.), *Dictionnaire des sciences naturelles, dans lequel on traite methodiquement des differens etres de la nature* V: 339.
- Tixier, P. 1960. Données cytologiques sur quelques Guttiferales au Laos. *Revue Cytologique Biologique Végétal* 22: 65-70.
- Tootil, E. (ed.) 1984. *Dictionary of Botany*. Penguin Book, Middlesex.
- Treub, M. 1911. Le sac embryonnaire et l'embryon dan les angiospermes. *Nouvelle série de recherches. Annales du Jardin Botanique de Buitenzorg* 24: 1-17.
- Turesson, G. 1929. Zur natur und begrenzung der arteinheiten. *Hereditas* 12: 323.
- Turner, I. M. & P. F. Stevens. 1999. The transfer of *Tripetalum cymosum* K. Schum. (Guttiferae) to *Garcinia*. *Gardens' Bulletin Singapore* 51 (2): 175-177.
- Verheij, J. W. M. 1991. *Garcinia mangostana* L. In: J. W. M. Verheij and R. E. Coronel (eds.), *Plant Resources of Southeast Asia No. 2. Edible fruits and nuts*: 177-181. PROSEA, Bogor.
- Vesque, J. 1889. Epharמושis, sive materia ad instruendam anatomiam systematis naturalis. Pars 2 : Genitalia foliaque Garciniearum et Calophyllearum (tab. i-clxii). Celapierre, Vincennes.
- Vesque, J. 1893. Guttiferae. In: A. deCandolle and C. deCandolle (eds.), *Monographiae phanerogamarum prodromi nunc continuato, nunc revisio vol. VIII. Sumptibus G. Masson, Paris*.
- Vieira, L. M., A. Kijjoa, A. M. Silva, I. O. Mondranondra, S. Kengthong, L. Gales, A. M. Damas & W. Herz. 2004. Lanostanes and friedolanostanes from the bark of *Garcinia speciosa*. *Phytochemistry* 65(4): 393-398.
- Voravuthikunchai, S. P. & L. Kitpipit. 2005. Activity of medicinal plant extracts against hospital isolates of methicillin-resistant *Staphylococcus aureus*. *Clinical Microbiology and Infection* 11(6): 510-512.
- Wakasugi, T., M. Sugita, T. Tsudzuki & M. Sugiura. 1998. Updated gene map of tobacco chloroplast DNA. *Plant Molecular Biology Reporter* 16: 231-241.
- Wallace, A. R. 1863. On the physical geography of the Malay archipelago. *Journal of the Royal Geographical Society of London* 33: 217-234.
- Wallich, N. 1828-1849. *A Numerical List of Dried Specimens of plant, in the East India Company's Museum*.
- Warburg, O. 1891. Beitrage zur kenntnis der Papuanischen flora. *Botanische Jahrbücher für Systematik* 13: 382.

- Watt, G. 1890. A dictionary of the economic products of India. Volume 3: *Dacrydium to Gordonia*. Superintendent of Government Printing, Calcutta.
- Weber, H. E. 1998. (46) Proposal concerning the names of apomictic species aggregates. *Taxon* 47(2): 495.
- Wheeler, Q.D. & Platnick, N.I. 2000. The phylogenetic species concept (sensu Wheeler and Platnick). In: Q. D. Wheeler & R. Meier (eds), *Species concepts and phylogenetic theory: a debate*: 55–69. Columbia University Press, New York.
- White, T. J., T. Bruns, S. Lee & J. Taylor. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: D. G. M. Innis, J. Snisky, T. White (ed.), *PCR Protocols: A Guide to Methods and Amplifications*: 315-322. Academic Press, San Diego.
- Whitmore, T. C. 1973. Guttiferae. In: T. C. Whitmore (ed.), *Tree flora of Malaya*: 162-236. Longman Malaysia, Kuala Lumpur.
- Whitmore, T. C. 1975. *Tropical rain forest of the Far East*. Clarendon, Oxford.
- Whitmore, T. C. 1988. *An introduction to tropical forest*. Oxford University Press, New York.
- Wight, R. 1840. *Icones plantarum Indiae orientalis* 1 (Illustrations of Indian botany 1). J.B. Pharoah, Madras.
- Williams, P., M. Ongsakul, J. Proudfoot, K. Croft & L. Beilin. 1995. Mangostin inhibits the oxidative modification of human low density lipoprotein. *Free Radical Research* 23(2): 175-184.
- Wolseley, P., L. Ellis, A. Harrington & C. Moncrieff. 1996 Epiphytic cryptograms at Pasoh Forest Reserve, N. Sembilan, Malaysia - quantitative and qualitative sampling in logged and unlogged plots. In: S. S. M. Lee, D. Y. Gauld, I. D. and Bishop, J. (ed.) *Conservation, Management and Development of Forest Resources. Proceedings of the Malaysia-U.K. Programme Workshop*: 61-83. Forest Research Institute of Malaysia, Kepong.
- Xu, D. H., A. J. Sakai, M. Kanazawa, A. Shimamoto & Y. Shimamoto. 2000. Sequence variation of non-coding regions of chloroplast DNA of soybean and related wild species and its implications for the evolution of different chloroplast haplotypes. *Theoretical and Applied Genetics* 101: 724-732.
- Yapwattanaphun, C., S. Kanzaki, K. Yonemori & S. Subhadrabandhu. 2003. Genetic variation among mangosteen (*G. mangostana* L.) revealed by AFLP analysis. *Thai Journal of Agricultural Science* 36 (3): 329-338.
- Yapwattanaphun, C., S. Subhadrabandhu, C. Honsho & K. Yonemori. 2004. Phylogenetic Relationship of Mangosteen (*Garcinia mangostana*) and several wild relatives (*Garcinia* spp.) revealed by ITS sequence data. *Journal of the American Society for Horticultural Science* 129 (3): 368-373.
- Young, N. D. & J. Healy. 2003. GapCoder automates the use of the indel characters in phylogenetic analysis. *BMC Bioinformatics* 6: 6.
- Zappi, D. C. 1993. A new combination in *Garcinia* (Guttiferae). *Kew Bulletin* 48(2): 410.

## APPENDIX I

Scientific names of sect. *Garcinia* in IPNI plus new names described in this study; names in bold currently belong to sect. *Garcinia* as defined in this thesis (Chapter 6) and asterisks denote those originally listed by Jones (1980) including invalid names.

Species	Taxonomic Status	Taxonomic Notes
1. <b><i>G. acuticosta</i> Nazre</b>	<b>Sect. <i>Garcinia</i>/Sp. nov</b>	Newly described species based on the fruit and evidence from molecular data.
2. <i>G. affinis</i> Wall. ex Pierre*	Illegitimate use	Wallich 4854 in Pierre (1882) identified as <i>G. celebica</i> L.
3. <i>G. affinis</i> Wight & Arn.	Syn. <i>G. cambogia</i> Desr.	Wallich 4866 in Wight & Arn. (1834) identified as <i>G. cambogia</i> Desr.
4. <i>G. anomala</i> Planch. & Triana*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> from inflorescences & male flower.
5. <i>G. baillonii</i> Pierre*	Non sect. <i>Garcinia</i> /Syn. <i>G. maingayi</i> Hk. f.	Differ from sect. <i>Garcinia</i> from the inflorescences, male flower, fruit & leaves.
6. <b><i>G. basacensis</i> Pierre*</b>	<b>Syn. <i>G. celebica</i> L.</b>	Shared the same male flower & leaves with <i>G. celebica</i> L.
7. <b><i>G. benthami</i> Pierre*</b>	<b>Syn. <i>G. celebica</i> L.</b>	Shared the same male flower, leaves & fruit with <i>G. celebica</i> L.
8. <i>G. blancoi</i> Pierre*	Unknown	Insufficient material (male specimen in bud).
9. <i>G. calleryi</i> Pierre*	Syn. <i>G. blancoi</i> Pierre	Insufficient material (male specimen in bud).
10. <b><i>G. cataractalis</i> Whitmore</b>	<b>Sect. <i>Garcinia</i></b>	Newly included members based on the fruit & evidence from molecular data.
11. <b><i>G. celebica</i> L.*</b>	<b>Sect. <i>Garcinia</i></b>	The earliest valid name (1754) of many synonyms.
12. <i>G. chapelierii</i> (Planch. & Triana) H.Perrier*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> from male flower, fruit & stigma.
13. <b><i>G. cornea</i> L.*</b>	<b>Syn. <i>G. celebica</i> L.</b>	Described in 1774; similar fruit & male flower as <i>G. celebica</i> L.
14. <i>G. cornea</i> Roxb. ex Sm.	Illegitimate use	= <i>G. cornea</i> L. (Roxburgh, 1832).
15. <i>G. costata</i> Hemsl.ex King*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> from the inflorescences, male

16. <i>G. cumingiana</i> Pierre*	Syn. <i>G. venulosa</i> (Blanco) Choisy	flower & fruit.
17. <i>G. diospyrifolia</i> Pierre var. <i>diospyrifolia</i> *	Sect. <i>Garcinia</i>	Similar male flower in Cumming 2114 with <i>G. venulosa</i> .
18. <i>G. diospyrifolia</i> var. <i>arborea</i>	Sect. <i>Garcinia</i> /Var. nov.	Slender small tree variety.
19. <i>G. diospyrifolia</i> var. <i>minor</i>	Sect. <i>Garcinia</i> /Var. nov.	New variety – medium sized tree.
20. <i>G. discoidea</i> Nazre	Sect. <i>Garcinia</i> /Sp. nov.	New variety – bushy small tree.
21. <i>G. erythrosperma</i> Lauterb.*	Unknown	Newly described species; previously known as <i>G. hombroniana</i> var. <i>A</i> Whitmore.
22. <i>G. exiguus</i> Nazre	Sect. <i>Garcinia</i> /Sp. nov.	Insufficient material & no type specimen available.
23. <i>G. fabrilis</i> Miq.*	Syn. <i>G. celebica</i> L.	Newly described species; share similar fruit & leaves.
24. <i>G. fascicularis</i> Wall*	Syn. <i>G. penangiana</i> L.	Described by Miquel (1861) found in Sumatra.
25. <i>G. ferrea</i> Pierre*	Syn. <i>G. celebica</i> L.	Collected by Wallich (1831) in Penang.
26. <i>G. harmandii</i> Pierre*	Sect. <i>Garcinia</i>	Described by Pierre (1882) found in Cambodia.
27. <i>G. hombroniana</i> Pierre*	Syn. <i>G. celebica</i> l.	Belongs to sect. <i>Garcinia</i> from the male flowers.
28. <i>G. jawoera</i> Pierre*	Syn. <i>G. celebica</i> L.	Described by Pierre (1882) found in Peninsular Malaysia.
29. <i>G. kingii</i> Pierre ex Vesque*	Syn. <i>G. celebica</i> L.	Described by Pierre (1882) found in Java.
30. <i>G. krawang</i> Pierre*	Syn. <i>G. celebica</i> L.	Described by Vesque (1891) found in Andaman.
31. <i>G. kurzii</i> Pierre*	Syn. <i>G. celebica</i> L.	Described by Pierre (1882) found in Kalimantan.
32. <i>G. lucens</i> Pierre*	Unknown	Described by Pierre (1882) found in Nicobar.
33. <i>G. macrophylla</i> Miq.*	Illegitimate /Non Sect. <i>Garcinia</i>	Insufficient material (male in buds).
34. <i>G. macrophylla</i> T.Anders. ex Hook.f.	Illegitimate/ Non Sect. <i>Garcinia</i>	Name was used earlier by Martius (1841)/ identified as <i>G. griffithii</i> T. Anders.
35. <i>G. maingayi</i> Hook.f.*	Non Sect. <i>Garcinia</i>	Name was used earlier by Martius (1841)/ identified as <i>G. griffithii</i> T. Anders.
36. <i>G. malaccensis</i> Hook. f. var. <i>malaccensis</i> *	Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> from the inflorescences, male flower & fruit.
37. <i>G. malaccensis</i> var. <i>pseudomangostana</i> Nazre	Sect. <i>Garcinia</i> /var. nov.	Varieties based on male flower and fruit.
38. <i>G. mangostana</i> L.*	Sect. <i>Garcinia</i>	A new varieties based on male flower and fruit.
39. <i>G. moselleyana</i> Pierre*	Sect. <i>Garcinia</i>	Type for genus and for the section.
40. <i>G. moulmienensis</i> Pierre ex Vesque*	Unknown	Belongs to sect. <i>Garcinia</i> from the male flowers.
41. <i>G. nitida</i> Pierre*	Sect. <i>Garcinia</i>	Insufficient material (female flower).
		Belongs to sect. <i>Garcinia</i> from male flowers, fruit and leaves.

<b>42. <i>G. ochraceus</i> Nazre</b>	<b>Sect. <i>Garcinia</i>/Sp. nov.</b>	Newly described species & members of sect. <i>Garcinia</i> based on male flowers & fruit.
<b>43. <i>G. opaca</i> King*</b>	<b>Syn. <i>G. diospyrifolia</i> Pierre</b>	Described by King (1890) found in Peninsular Malaysia.
<b>44. <i>G. opaca</i> var. <i>dumosa</i> Whitmore</b>	<b>Syn. <i>G. diospyrifolia</i> var. <i>diospyrifolia</i></b>	Described by Whitmore (1973) found in Peninsular Malaysia.
<b>45. <i>G. penangiana</i> Pierre*</b>	<b>Sect. <i>Garcinia</i></b>	Belongs to sect. <i>Garcinia</i> from male flowers, fruit and leaves.
46. <i>G. porrecta</i> Laness.	Invalid/Syn. <i>G. cornea</i> L.	Used by Lanessan (1872) for synonym of <i>G. cornea</i> L.
47. <i>G. porrecta</i> Wall.*	Nom. nudum	Used by Wallich (1831); cultivated tree in BO.
48. <i>G. porrecta</i> var. <i>schizogyna</i> Boerl.	Invalid/Syn. <i>G. rigida</i>	Used by Boerlage (1901); cultivated tree in BO.
49. <i>G. propinqua</i> Craib*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> from inflorescences & male flower.
50. <i>G. pseudo-guttifera</i> Seem.*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> from inflorescences & male flower.
<b>51. <i>G. riedeliana</i> Pierre*</b>	<b>Syn. <i>G. celebica</i> L.</b>	Described by Pierre (1882) found in Sulawesi.
<b>52. <i>G. rigida</i> Miq.*</b>	<b>Sect. <i>Garcinia</i></b>	Belongs to sect. <i>Garcinia</i> from fruit and molecular evidence.
<b>53. <i>G. rumphii</i> Pierre*</b>	<b>Syn. <i>G. celebica</i> L.</b>	Described by Pierre (1882) found in Bangka Is., Java.
<b>54. <i>G. schefferi</i> Pierre*</b>	<b>Syn. <i>G. rigida</i> Pierre</b>	Shared the same fruit with <i>G. rigida</i> Pierre.
<b>55. <i>G. speciosa</i> Wall.*</b>	<b>Syn. <i>G. celebica</i> L.</b>	Described by Wallich (1832) found in Burma.
<b>56. <i>G. sangudsangud</i> Nazre</b>	<b>Sect. <i>Garcinia</i>/Sp. nov.</b>	Newly described species & members of sect. <i>Garcinia</i> based on male flowers & fruit.
57. <i>G. squamata</i> Lauterb.*	Unknown	Insufficient material (female flower).
58. <i>G. tonkinensis</i> Vesque*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> based on the inflorescences, male flower & fruit.
59. <i>G. trianaii</i> Pierre*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> based on the inflorescences, male flower & fruit.
<b>60. <i>G. venulosa</i> Choisy*</b>	<b>Sect. <i>Garcinia</i></b>	Belongs to sect. <i>Garcinia</i> from inflorescences, fruits & male flowers.
61. <i>G. vidua</i> Ridl.*	Non Sect. <i>Garcinia</i>	Differ from sect. <i>Garcinia</i> based on the inflorescences, male flower & fruit.

## APPENDIX II

Species, accessions and origin of samples used in molecular analysis for this study.

	Species (synonym)	Accession No.	Collectors	Voucher	Locality Collected [Origin]	ITS	Sequence	
							<i>trnS-G</i>	<i>trnD-T</i>
1	<i>G. acutacosta</i>	BB01	M. Nazre et al.	BB01	Bkt. Bauk Fst. Res., Compt. 22			
2	<i>G. balica</i>	021	M. Nazre et al.	VI.C.365	Bogor Bot. Gdns., Java [Maluku Is.]			-
3	<i>G. cataractalis</i>	077	M. Nazre et al.	FRIM01	FRIM Kepong			
4	<i>G. celebica</i>	002	M. Nazre et al.	CR01	Cape Rechado, Melaka, Malaysia.			
5	<i>G. celebica (G. hombroniana)</i>	003	M. Nazre et al.	Nur03	Kepong, Malaysia			-
6	<i>G. celebica (G. hombroniana)</i>	005	M. Nazre et al.	Nur01	Kepong, Malaysia.		-	-
7	<i>G. celebica (G. hombroniana)</i>	009	M. Nazre et al.	Mardi01	MARDI, Selangor, Malaysia.			-
8	<i>G. celebica</i>	023	M. Nazre et al.	VI.C.18	Bogor Bot. Gdns., Java [Borneo Is.:Kalimantan]			
9	<i>G. celebica (G. benthami)</i>	027	M. Nazre et al.	IV.C.106	Bogor Bot. Gdns., Java [Vietnam]			-
10	<i>G. celebica</i>	028	M. Nazre et al.	VI.C.273	Bogor Bot. Gdns., Java [W. Java]			-
11	<i>G. celebica</i>	033	M. Nazre et al.	VI.C.335	Bogor Bot. Gdns., Java [Borneo]			
12	<i>G. celebica</i>	038	M. Nazre et al.	VI.A.16	Bogor Botanic Gdns, Java [Sulawesi]		-	-
13	<i>G. celebica</i>	043	M. Nazre et al.	-	Bogor Botanic Gdns, Java [Java]		-	-
14	<i>G. celebica</i>	053	M. Nazre et al.	VI.A.153	Bogor Botanic Gdns, Java [Sumatra]		-	-
15	<i>G. celebica</i>	054	M. Nazre et al.	VI.C.335a	Bogor Bot. Gdns., Java [Borneo]		-	-
16	<i>G. celebica (G. hombroniana)</i>	059	M. Nazre et al.	CR02	Cape Rechado, Melaka, Malaysia.			-
17	<i>G. celebica</i>	062	M. Nazre et al.	VI.A.4	Bogor Bot. Gdns., Java [P. Malaysia]			-
18	<i>G. costata</i>	BL03	M. Nazre et al.	BL03	Bkt. Larut, behind Rumah Delima			-
19	<i>G. costata</i>	BL06	M. Nazre et al.	BL06	Bkt Larut, 900m besides main road.		-	-
20	<i>G. cowa</i>	004	M. Nazre et al.	Nur02	Kepong, Malaysia		-	-
21	<i>G. cowa (misid. as G. dulcis)</i>	013	M. Nazre et al.	Nur04	Kepong, Malaysia		-	-
22	<i>G. cymosa</i>	037	M. Nazre et al.	XXIV.A.16	Bogor Botanic Gdns, Java [Irian]			
23	<i>G. cymosa aff.</i>	042	M. Nazre et al.	VI.C.338	Bogor Botanic Gdns, Java [Maluku]			-
24	<i>G. diospyrifolia var. arborea</i>	BB02	M. Nazre et al.	BB02	Bkt. Bauk Fst. Res.		-	-
25	<i>G. diospyrifolia var. arborea (G.</i>	BB05	M. Nazre et al.	BB05	Bkt. Bauk Fst. Res., Compt. 22		-	-

	<i>opaca</i> )							
26	<i>G. diospyrifolia</i> var. <i>diospyrifolia</i> ( <i>G. opaca</i> var. <i>dumosa</i> )	078	M. Nazre et al.	MN78	Rompin, Pahang			
27	<i>G. diospyrifolia</i> var. <i>minor</i>	BB06	M. Nazre et al.	BB06	Bkt. Bauk Fst. Res., Compt. 22			
28	<i>G. discoidea</i>	SL01	M. Nazre et al.	SL01	Sg. Loh, near logging camp			
29	<i>G. discoidea</i>	SL02	M. Nazre et al.	SL02	Sg. Loh, main logging road to Compt 33		-	-
30	<i>G. dulcis</i>	050	M. Nazre et al.	IV.C.114	Bogor Bot. Gdns., Java [Sumatra]			-
31	<i>G. echinocarpa</i>	035	M. Nazre et al.	VI.A.36	Bogor Botanic Gdns, Java [Sri Lanka]		-	-
32	<i>G. forbesii</i>	024	M. Nazre et al.	VI.A.58	Bogor Botanic Gdns, Java [Kalimantan]	-		-
33	<i>G. lateriflora</i>	040	M. Nazre et al.	VI.A.17	Bogor Botanic Gdns, Java [Java]			-
34	<i>G. latissima</i>	026	M. Nazre et al.	VI.C.54	Bogor Bot. Gdns., Java [South Papua]		-	-
35	<i>G. maingayi</i>	098	Latiff. et. al	ALM 6120	Rompin, Pahang		-	-
36	<i>G. malaccensis</i>	BB04	M. Nazre et al.	BB04	Bkt. Bauk Fst. Res.		-	-
37	<i>G. malaccensis</i>	BB08	M. Nazre et al.	BB08	Bkt. Bauk Fst. Res., Compt. 22			
38	<i>G. malaccensis</i>	BB09	M. Nazre et al.	BB09	Bkt. Bauk Fst. Res., Compt. 22			
39	<i>G. malaccensis</i> var. <i>pseudomangostana</i>	SP01	M. Nazre et al.	SP01	Sepilok Fst. Reserve, Behind			
40	<i>G. mangostana</i>	096	Shamsul et al.	SK-MY-10	Universiti Putra Malaysia.		-	-
41	<i>G. mangostana</i>	LAO	Newman s.n.	LAO	Laos			
42	<i>G. megaphylla</i>	030	M. Nazre et al.	VI.C.275	Bogor Bot. Gdns., Java [South America]		-	-
43	<i>G. megaphylla</i>	041	M. Nazre et al.	VI.C.277	Bogor Bot. Gdns., Java [South America]		-	
44	<i>G. penangiana</i>	006	M. Nazre et al.	KT01	Kota Tinggi, Johor, Malaysia.			
45	<i>G. penangiana</i>	BB07	M. Nazre et al.	BB07	Bkt. Bauk Fst. Res., Compt. 22		-	-
46	<i>G. picrorhiza</i>	039	M. Nazre et al.	VI.A.27	Bogor Botanic Gdns, Java [Maluku] Ambon			-
47	<i>G. rigida</i>	029	M. Nazre et al.	VI.C.235	Bogor Bot. Gdns., Java [Maluku]			
48	<i>G. rigida</i>	045	M. Nazre et al.	XXIII.A.122	Bogor Bot. Gdns., Java [S. Sulawesi]			
49	<i>G. rigida</i>	049	M. Nazre et al.	VI.A.50	Bogor Bot. Gdns., Java [Maluku]			
50	<i>G. rigida</i>	057	M. Nazre et al.	VI.C.145	Bogor Bot. Gdns., Java [Maluku]			
51	<i>G. rostrata</i>	091	Shamsul et al.	SK-MY-05	Menchali FR, Rompin Pahang			-
52	<i>G. rostrata</i>	BL05-1	M. Nazre et al.	BL05-1	Bkt. Larut, Perak.		-	-

53	<i>G. rostrata</i>	007	M. Nazre et al.	KT02	Kota Tinggi, Johor, Malaysia.			
54	<i>G. scortechinii</i>	012	M. Nazre et al.	Fraser05	Fraser's Hill, Pahang, Malaysia.		-	-
55	<i>G. scortechinii</i>	089	Shamsul et al.	SK-MY-09	Gunung Nuang, Selangor			-
56	<i>G. scortechinii</i>	K04	M. Nazre et al.	K04	Kinabalu Park, Bundu Tuhan Trail			
57	<i>G. sp. B1</i>	008	M. Nazre et al.	Fraser02	Fraser's Hill, Pahang, Malaysia.		-	-
58	<i>G. sp. B1</i>	011	M. Nazre et al.	Fraser04	Fraser's Hill, Pahang, Malaysia.			-
59	<i>G. sp. D1</i>	064	M. Nazre et al.	SULTENG03	Sulawesi Tengah			-
60	<i>G. sp. D1</i>	070	M. Nazre et al.	SULTENG09	G. Hek, Sulawesi Tengah			-
61	<i>G. sp. D2</i>	P01	M. Nazre et al.	P01	Poring Hotspring, trail to canopy walkaway		-	-
62	<i>G. sp. D3</i>	P02	M. Nazre et al.	P02	Poring Hotspring, trail to waterfall			
63	<i>G. sp. D4</i>	BL04	M. Nazre et al.	BL04	Bkt. Larut, behind Rumah Delima		-	-
64	<i>G. sp. D5</i>	BL05-2	M. Nazre et al.	BL05-2	Bkt Larut, 900m besides main road.		-	-
65	<i>G. sp. D6</i>	K12	M. Nazre et al.	K12	Kinabalu Park, Bundu Tuhan Trail		-	-
66	<i>G. sp. M1</i>	025	M. Nazre et al.	-	Bogor Bot. Gdns., Java		-	-
67	<i>G. sp. M2</i>	068	M. Nazre et al.	SULTENG07	G. Hek, Sulawesi Tengah		-	-
68	<i>G. sp. M3</i>	072	M. Nazre et al.	SULTENG11	G. Hek, Sulawesi Tengah		-	-
69	<i>G. sp. M4</i>	022	M. Nazre et al.	IV.C.378	Bogor Bot. Gdns., Java [Sulawesi]		-	
70	<i>G. sp. M5</i>	069	M. Nazre et al.	SULTENG08	G. Hek, Sulawesi Tengah			-
71	<i>G. sp. M5</i>	073	M. Nazre et al.	SULTENG12	G. Hek, Sulawesi Tengah			-
72	<i>G. sp. M6</i>	093	Shamsul et al.	SK-MY-06	Menchali FR, Rompin Pahang			-
73	<i>G. sp. M6</i>	094	Shamsul et al.	SK-MY-08	Apau FR, K. Kelawang NS			-
74	<i>G. sp. X1</i>	052	M. Nazre et al.	VI.C.373	Bogor Botanic Gdns, Java			-
75	<i>G. sp. X2</i>	080	M. Nazre et al.	22B	Sg. Dusun, Selangor			
76	<i>G. spicata</i>	019	M. Nazre et al.	VI.C.111	Bogor Bot. Gdns., Java [Sri Lanka]			-
77	<i>G. trianaii</i>	K02	M. Nazre et al.	K02	Kinabalu Park, Bundu Tuhan Trail			
78	<i>G. urophylla</i>	001	M. Nazre et al.	Fraser01	Fraser's Hill, Pahang, Malaysia.			-
79	<i>G. urophylla</i>	081	M. Nazre et al.	14B	Sg. Dusun, Selangor			-
80	<i>G. xanthochymus</i>	034	M. Nazre et al.	VI.A.52	Bogor Botanic Gdns, Java		-	-
81	<i>G. xanthochymus</i>	BB03	M. Nazre et al.	BB03	Bkt. Bauk Fst. Res.		-	-
82	<i>G. xanthochymus cf.</i>	032	M. Nazre et al.	VI.A.42	Bogor Bot. Gdns., Java Sumatera Is.			-
83	<i>G. xanthochymus?</i>	092	Shamsul et al.	SK-MY-07	Menchali FR, Rompin Pahang		-	-

### APPENDIX III

List of sequences taken from GENBANK used in this study.

	Species [synonym]	Author	Origin	GenBank Accession
1	<i>G. acuminata</i>	Yapwattanaphun et al.	Thailand	AB 110796
2	<i>G. atroviridis</i>	Yapwattanaphun et al.	Thailand	AB 110797
3	<i>G. atroviridis</i>	M. Nazre et al.	P. Malaysia	AF 367211
4	<i>G. celebica</i> [ <i>G. hombroniana</i> ]	Yapwattanaphun et al.	Thailand	AB 110803
5	<i>G. celebica</i> [ <i>G. hombroniana</i> ]	Yapwattanaphun et al.	Thailand	AB 110804
6	<i>G. celebica</i> [ <i>G. hombroniana</i> ]	M. Nazre et al.	P. Malaysia	AF 367214
7	<i>G. celebica</i> [ <i>G. speciosa</i> ]	Yapwattanaphun et al.	Thailand	AB 110816
8	<i>G. costata</i>	Yapwattanaphun et al.	Thailand	AB 110798
9	<i>G. cowa</i>	Yapwattanaphun et al.	Thailand	AB 110799
10	<i>G. diospyrifolia</i> var. <i>minor</i> [ <i>G. opaca</i> var. <i>minor</i> ]	M. Nazre et al.	P. Malaysia	AF 367227
11	<i>G. dulcis</i>	Yapwattanaphun et al.	Thailand	AB 110800
12	<i>G. dulcis</i>	Yapwattanaphun et al.	Thailand	AB 110801
13	<i>G. forbesii</i>	M. Nazre et al.	P. Malaysia	AF 367224
14	<i>G. globulosa</i>	M. Nazre et al.	P. Malaysia	AF 367221
15	<i>G. hanburyi</i>	Yapwattanaphun et al.	Thailand	AB 110802
16	<i>G. latissima</i>	Abdul-Salim, K. and Dick	P. Malaysia	AF 479789
17	<i>G. madruno</i>	Gehrig et al.	S. America	AJ 509215
18	<i>G. malaccensis</i>	Yapwattanaphun et al.	Java, Indonesia	AB 110805
19	<i>G. malaccensis</i>	Yapwattanaphun et al.	Java, Indonesia	AB 110806
20	<i>G. mangostana</i>	Yapwattanaphun et al.	Java, Indonesia	AB 110807
21	<i>G. mangostana</i>	Yapwattanaphun et al.	P. Malaysia	AB 110808
22	<i>G. mangostana</i>	Yapwattanaphun et al.	Thailand	AB 110809
23	<i>G. mangostana</i>	Yapwattanaphun et al.	Thailand	AB 110810
24	<i>G. mangostana</i>	Yapwattanaphun et al.	Thailand	AB 110811
25	<i>G. mangostana</i>	Gehrig et al.	S. America	AJ 509214
26	<i>G. nervosa</i>	M. Nazre et al.	P. Malaysia	AF 367222
27	<i>G. penangiana</i> (misid. as <i>G. malaccensis</i> )	M. Nazre et al.	P. Malaysia	AF 367226
28	<i>G. prainiana</i>	M. Nazre et al.	P. Malaysia	AF 367218
29	<i>G. rostrata</i>	M. Nazre et al.	P. Malaysia	AF 367225

30	<i>G. rostrata</i> (misidentification of <i>G. celebica</i> )	Yapwattanaphun et al.	Thailand	AB 110813
31	<i>G. schomburgkiana</i>	Yapwattanaphun et al.	Thailand	AB 110814
32	<i>G. scortechinii</i>	M. Nazre et al.	P. Malaysia	AF 367219
33	<i>G. sp.</i>	Gehrig et al.	S. America	AJ 312607
34	<i>G. syzygifolia</i> (misidentification of <i>G. rigida</i> )	Yapwattanaphun et al.	Java, Indonesia	AB 110815
35	<i>G. tinctoria</i>	Yapwattanaphun et al.	Australia	AB 110817
36	<i>G. vilersiana</i>	Yapwattanaphun et al.	Thailand	AB 110818
37	<i>G. xanthochymus</i>	Yapwattanaphun et al.	Thailand	AB 110819
38	<i>G. bancana</i>	M. Nazre et al.	P. Malaysia	AF 367212
39	<i>Symphonia globulifera</i>	Gustafsson & Bittrich	-	AY 682919
40	<i>Tripetalum cymosum</i>	Gustafsson & Bittrich	-	AY 145206
41	<i>Clusia rosea</i>	Gehrig et al.	S. America	AJ 509230

**Molecular Lab Protocol**

A. DNA extraction (Doyle and Doyle, 1990)

1. Preheat 1ml CTAB (Cetyltrimmonium bromide) mixed with 2 $\mu$ l 2-mercaptoethanol per sample at 65 $^{\circ}$ C in the waterbath.
2. Harvest approximately 0.50g of leaf material into the 1.5ml eppendorf tube and add 2 spatulas of sand.
3. With the eppendorf lid open, immerse the tube into liquid nitrogen with long handled forceps, being careful not to let any leaf material out of the tube. Hold the tube until it stops making a bubbling noise.
4. Carefully macerate the leaf tissue with a plastic pestle until the material is finely graved. The sample can be re-chilled in liquid nitrogen if difficult.
5. Add 500 $\mu$ l of the pre-heated CTAB buffer and continue to gently grind the sample.
6. Add further 500 $\mu$ l of CTAB buffer with a pinch of PVPP (polyvinylpyrrolidone) and mix the contents by flicking the tube.
7. Incubate the mixture for 30 minute at 65 $^{\circ}$ C in the heated block.
8. Remove the mixture from the heated block and allow to cool to ambient temperature.
9. Add 500 $\mu$ l of wet chloroform (24 chloroform : 1 isoamyl alcohol) and mix gently by hand to obtain a momentary single phase. Transfer the mixture to the orbital shaker and shake on minimum speed for 10-20 minutes.
10. Centrifuge the mixture for 10 minutes at 13,000 rpm.
11. Remove the supernatant (upper aqueous layer) to a clean 1.5ml eppendorf tube and repeat the chloroform extraction (step 9 and 10).
12. Transfer the supernatant to a clean 1.5ml eppendorf tube and precipitate the DNA by adding 2/3 volume of cold isopropanol (-20 $^{\circ}$ C) and mix gently. The mixture can be left overnight to increase the amount of DNA.
13. Centrifuge the mixture for 10 minutes at 13,000 rpm to pellet out the DNA.
14. Remove the supernatant and wash the DNA by adding 1ml wash buffer (76% ethanol, 10mM NH<sub>4</sub> Ac). Leave the DNA for at least 30 minutes.
15. Centrifuge the tube for 5 minutes at 13,000 rpm and discard the supernatant being careful not to dislodge the DNA pellet. Dry the pellet in the vacuum centrifuge for 5 minutes or until the pellet is dry.
16. Dissolve the pellet in 50 $\mu$ l of TE (10mM Tris HCl ph 8.0, 1mM EDTA ph 8.0) buffer.
17. To determine the quality of the DNA, 1% agarose gel electrophoresis was run and DNA concentrated was compared with 1kb+ marker.

B. DNA Extraction - DNeasy® Plant Mini (Qiagen, USA)

1. Preheat a water bath or heating block to 65 $^{\circ}$ C.
2. Grind plant or fungal tissue under liquid nitrogen to a fine powder using a mortar and pestle. Transfer the tissue powder and liquid nitrogen to an appropriately sized tube and allow the liquid nitrogen to evaporate. Do not allow the sample to thaw.

3. Add 400  $\mu$ l of Buffer AP1 and 4  $\mu$ l of RNase A stock solution (100 mg/ml) to a maximum of 100 mg of ground (wet weight) or 20 mg (dried) plant or fungal tissue and vortex vigorously. No tissue clumps should be visible. Vortex or pipet further to remove any clumps. Clumped tissue will not lyse properly and will therefore result in a lower yield of DNA. In the rare case where clumps cannot be removed by pipetting and vortexing, a disposable micropestle may be used. Note: Do not mix Buffer AP1 and RNase A before use.
4. Incubate the mixture for 10 min at 65°C. Mix 2–3 times during incubation by inverting tube. This step lyses the cells.
5. Add 130  $\mu$ l of Buffer AP2 to the lysate, mix, and incubate for 5 min on ice. This step precipitates detergent, proteins, and polysaccharides. Optional: Centrifuge the lysate for 5 min at 20,000 x g (14,000 rpm). After centrifugation, apply supernatant to QIAshredder Mini Spin Column.
6. Apply the lysate to the QIAshredder Mini Spin Column (lilac) placed in a 2 ml collection tube and centrifuge for 2 min at 20,000 x g (14,000 rpm). It may be necessary to cut the end off the pipet tip to apply the lysate to the QIAshredder Mini Column. The QIAshredder Mini Column removes most precipitates and cell debris, but a small amount will pass through and form a pellet in the collection tube.
7. Transfer flow-through fraction from step 4 to a new tube (not supplied) without disturbing the cell-debris pellet. Typically 450  $\mu$ l of lysate is recovered. For some plant species less lysate is recovered.
8. Add 1.5 volumes of Buffer AP3/E to the cleared lysate and mix by pipetting. Buffer AP3/E. Reduce the amount of Buffer AP3/E accordingly if less lysate is recovered. A precipitate may form after the addition of ethanol but this will not affect the DNeasy procedure. Note: It is important to pipet Buffer AP3/E directly onto the cleared lysate and to mix immediately.
9. Apply 650  $\mu$ l of the mixture from step 6, including any precipitate which may have formed, to the DNeasy Mini Spin Column sitting in a 2 ml collection tube (supplied). Centrifuge for 1 min at  $\geq 6000$  x g (corresponds to  $\geq 8000$  rpm for most microcentrifuges) and discard flow-through.\* Reuse the collection tube in step 9.
10. Repeat step 8 with remaining sample. Discard flow-through\* and collection tube.
11. Place DNeasy Mini Spin Column in a new 2 ml collection tube (supplied), add 500  $\mu$ l Buffer AW to the DNeasy Mini Spin Column and centrifuge for 1 min at  $\geq 6000$  x g ( $\geq 8000$  rpm). Discard flow-through and reuse the collection tube in step 11 Note: Ensure ethanol is added to Buffer AW.
12. Add 500  $\mu$ l Buffer AW to the DNeasy Mini Spin Column and centrifuge for 2 min at 20,000 x g (14,000 rpm) to dry the membrane.
13. Transfer the DNeasy Mini Spin Column to a 1.5 ml or 2 ml microcentrifuge tube and pipet 100  $\mu$ l of Buffer AE directly onto the DNeasy membrane. Incubate for 5 min at room temperature (15–25°C) and then centrifuge for 1 min at  $\geq 6000$  x g ( $\geq 8000$  rpm) to elute. Elution with 50  $\mu$ l (instead of 100  $\mu$ l) increases the final DNA concentration in the eluate significantly, but also reduces overall DNA yield. If larger amounts of DNA (>20  $\mu$ g) are loaded, eluting with 200  $\mu$ l (instead of 100  $\mu$ l) increases yield.
14. Repeat step 12 once.

### C. Ethanol Clean Up

1. Make each reaction to 20 $\mu$ l by adding deionised water and transfer to the 0.5ml microfuge tubes.

2. Add 5 $\mu$ l of stop solution (stock B) for each reaction.  
stock B;
  - i. 0.5M EDTA - 0.4 $\mu$ l
  - ii. Sigma Water - 1.6 $\mu$ l
  - iii. 3M NaOAc pH 5.2 - 2.0 $\mu$ l
  - iv. Glycogen (DTCS kit) - 1.0 $\mu$ l
3. Add 60 $\mu$ l of 95-100% ethanol (-20 $^{\circ}$ C).
4. Mix the reactions thoroughly.
5. Centrifuge the reactions at 13,000 rpm for 15 minutes at 4 $^{\circ}$ C.
6. Carefully remove the supernatant with the pipette.
7. Gently add 200 $\mu$ l of 70% ethanol (-20 $^{\circ}$ C).
8. Centrifuge at 13,000 rpm for 5 minutes at 4 $^{\circ}$ C.
9. Carefully remove the supernatant.
10. Repeat the steps 7 – 9.
11. Vacuum dry the pellet for 2-5 minutes until no trace of ethanol remains.
12. Re-suspend each pellet in 35-40 $\mu$ l of Sample Loading Solution (SLS) and mix well.



<i>G. dulcis</i> 050	TCGAAACCTGCC	AGCA	GCACGACCCG	TGAACCTGTTAAC	CACAT	TGACAAG	CGCGTG	TCGTCGGTCC	TCGGATTGGCAACATGC	CCTT	
<i>G. dulcis</i> AB110800	TCGAAACCTGCC	AGCA	GCACGACCCG	TGAACCTGTTTAC	CACAT	TGACAAG	TGCATG	TGGCCATTCC	TCGGATTGGCGACATGC	CCTT	
<i>G. dulcis</i> AB110801	TCGAAACCTGCC	AGCA	GCACGACCCG	TGAACCTGTTTAC	CACAT	TGACAAG	TGCATG	TGGCCATTCC	TCGGATTGGCGACATGC	CCTT	
<i>G. echinocarpa</i> 035	TCGAAACCTGCC	AACA	GCATGACCTG	TGAACCTAGTTTAA	CAAT	TGACAAG	GGTGTG	TGGTCAATCA	CTGGATTGGCAACATTG	CCTT	
<i>G. forbesii</i> AF367224	TCGAAACCTGCC	AATA	GCACGACCCG	TGAACCTGTTTAA	CACAT	TGACAAG	GGTGTG	TGGTCAGTCA	TTGGATTGGCAACATGC	CCCT	
<i>G. forbesii</i> AF67223	TCGAAACCTGCC	AATA	GCACGACCCG	TGAACCTGTTTAA	CACAT	TGACAAG	GGTGTG	TGGTCAGTCA	TTGGATTGGCAACATGC	CCCT	
<i>G. globulosa</i> AF367221	TCGAAACCTGCC	AATA	GCACGACCTG	TGAACCTGTTTAA	CACAT	TGACAAG	GGTGTG	TGGTCAGTCA	TTGGATTGGCAACCTGC	CCTT	
<i>G. hanburyi</i> AB110802	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CACAT	CGACAAG	GGCGTG	TTGTCATCA	TTGGATTGGCAACACGC	CCTT	
<i>G. laterifolia</i> 040	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CACAT	CGACAAG	GGCGTG	TTGTCAGTCA	TTGGATTGGTAACACGC	CCTT	
<i>G. latissima</i> 026	TCGAAACCTGCC	AACA	GCACGACCTG	TGAACCTGTTTAA	CACAT	TGACAAG	GGCGTG	TGGTCAGTCA	TTGGATTGGCAACACGC	CCTT	
<i>G. latissima</i> AF479789	TCGAAACCTGCC	AGCA	GCATGACCTG	TGAACCTGTTTAA	CACAC	TGACAAG	TGTGTG	TGCCCAGTCC	TTGGATTGGAGACATGC	ACTA	
<i>G. madruno</i> AJ509215	TCGAAACCTGCC	AGCA	GCACGACCTG	TGAACCTGTTTAA	CACAT	TGACCAG	TGTGTG	TGGCCAGTCC	TTGGATTGGCAACATGC	ACTA	
<i>G. maingayi</i> 098	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CACAT	TGACAAG	GGTGTG	TGGTCAGTCG	CTGGATTGGCAACACCC	CCTT	
<i>G. malaccensis</i> AB110810	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. malaccensis</i> AB110811	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. malaccensis</i> var. <i>malaccensis</i> BB04	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. malaccensis</i> var. <i>malaccensis</i> BB08	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. malaccensis</i> var. <i>malaccensis</i> BB09	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. malaccensis</i> var. <i>pseudomangostana</i> SP01	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> 096	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AB110807	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AB110808	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AB110809	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AB110810	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AB110811	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AF367215	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> AJ509214	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. mangostana</i> LAO	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACACGC	CCTT	
<i>G. megaphylla</i> 030	TCGAAACCTGCC	AGCA	GCACAACCTG	TGAACCTGTTTAA	CACAC	TGACAAG	TGTGTG	TGGCCAGTCC	TTGGATTGGAAACATGC	ACTA	
<i>G. megaphylla</i> 041	-----	ACCTGCC	AGCA	GCACGACCCG	TGAACCTGTTTAC	CACATT	CGACAAG	CGCGTG	TGGCCAGTCC	TCGGATTGGCAACATGC	CCTT
<i>G. nervosa</i> AF367222	TCGAAACCTGCC	AGCA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACAAG	CGCGTG	TGGCCAGTCC	TCGGATTGGCAACATGC	CCTT	
<i>G. nigrolineata</i> AF367216	TCGAAACCTGCC	AATA	GCACGACTCG	TGAACCTGTTTAA	CACAT	CGACAAG	GGTGTG	TGGTCAGTCA	TTGGATTGGCAACATA	CCTT	
<i>G. nitida</i> 119	TCGAAACCTGCC	AACA	GCATGACCCG	TGAACCTGTTTAA	CCCAT	TGACGAG	GGTGTG	TTGTTGGTCA	TTAGATTGGCGACACGC	CCTT	
<i>G. penangiana</i> 006	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGATGAG	GGTGTG	TTGTCATCA	CTAGATTGGCAACACGC	CCTT	
<i>G. penangiana</i> AF367226	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGATGAG	GGTGTG	TTGTCGATCA	CTAGATTGGCAACACGC	CCTT	
<i>G. penangiana</i> BB07	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGATGAG	GGTGTG	TTGTCGATCA	CTACATTGGCAACACGC	CCTT	
<i>G. picrorhiza</i> 039	TTGAAACCTACCA	AATA	GTTAGACTTG	CGAACCTGTTTTA	CACAT	TGACAAG	TGCGTG	TTGTCAGTCG	TTGACTTCTAACATGC	CCTT	
<i>G. porrecta</i> AB110812	TCGAAACCTGCC	AATA	GCACGACCCG	TGAACCTGTTTAA	CACAT	TGACAAG	GGTGTG	TGGTCAGTCA	TTGGATTGGCAACATGC	CCCT	
<i>G. prainiana</i> AF367218	TCGAAACCTGCC	AATA	GCACGACCCG	TGAACCTGTTTAA	CACAT	CAACAAG	TGTGTG	TGGTCAGTCT	TTGATTGGCAACACGC	CCTT	
<i>G. rigida</i> 029	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	AGACAAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCGACACGC	CCTT	
<i>G. rigida</i> 045	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACAAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCGACACGC	CCTT	
<i>G. rigida</i> 049	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACAAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCGACACGC	CCTT	
<i>G. rigida</i> 057	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	TGACAAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACATGC	CCTT	
<i>G. rostrata</i> 007	TCGAAACCTGCC	CACA	GCACCACACG	GAACCCGTTTAA	CACAT	TTGCAAG	TGTGTG	TGGTCAGTCG	TTGCATTGACAACATGC	CCTT	
<i>G. rostrata</i> 091	TCGAAACCTGCC	AACA	GCACGACACG	CGAACCTGTTTAT	CACAT	GCGCAAG	TGTGTG	TGGTCAGTTG	TTGGATTGACAACATGC	CCTT	
<i>G. rostrata</i> AB110813	TCGAAACCTGCC	AACA	GCACGACCCG	TGAACCTGTTTAA	CCCAT	CGACGAG	GGTGTG	TTGTCGGTCA	CTAGATTGGCAACATGC	CCTT	
<i>G. rostrata</i> AF367225	TCGAAACCTGCC	AACA	GCACGACACG	CGAACCTGTTTAT	CACAT	GCGCAAG	TGTGTG	TGGTCAGTTG	TTGGATTGACAACATGC	CCTT	

*G. rostrata* BL051  
*G. schomburgkiana* AB110814  
*G. scortechinii* AF367219  
*G. scortechinii* 012  
*G. scortechinii* 089  
*G. scortechinii* K04  
*G. sp.* AJ312607  
*G. sp.*B1 008  
*G. sp.*B2 011  
*G. sp.*D1 064  
*G. sp.*D1 070  
*G. sp.*D2 P01  
*G. sp.*D3 P02  
*G. sp.*D4 BL04  
*G. sp.*D5 BL052  
*G. sp.*D6 K12  
*G. spicata* 019  
*G. sp.*M1 025  
*G. sp.*M2 068  
*G. sp.*M3 072  
*G. sp.*M4 022  
*G. sp.*M5 069  
*G. sp.*M5 073  
*G. sp.*M6 093  
*G. sp.*M6 094  
*G. sp.*X1 052  
*G. sp.*X2 080  
*G. syzygifolia* AB110815  
*G. tinctoria* AB110817  
*G. trianaii* K02  
*G. urophylla* 001  
*G. urophylla* 081  
*G. vilersiana* AB110818  
*G. xanthochymus* AB110819  
*G. xanthochymus* 034  
*G. xanthochymus* 092  
*G. xanthochymus* BB03  
*G. xanthochymus*cf 032  
*Symphonia globulifera* AY682919  
*Tripetalum cymosum* AY145206  
*Clusia rosea*

```
TCGAAACCTGCC CACA GCACCACATGCGAACCCGTTTAA TACAT TTGCAAG TGTGTG TGGTCAGTCG TTGCATTGACAAATGC CCTT
TCGAAACTCGCTA AATA GCACAACCTGTAAGTCGTTTAA CACAT CGACAGA GGTGTG TGGTCAGTCA TTGGATTGGCAACATAC CCTT
TCGAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAT CGACAAG GGCGTG TTGTCAGTCA TTGGATTGGTAACACGC CCTT
-----CCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAT CGACAAG GGCGTG TCGTCAGTCA TTGGATTGGTAACACGC CCTT
TCGAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAT CGACAAG GGCGTG TCGTCAGTCA TTGGATTGGTAACACGC CCTT
TCAAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAT CGACAAG GGCGTG TGGTCAGTCA TTGGATTGGCAACACGC CCTT
TCAAAACCTGCC AGCA GCACGACCTGTGAAGTCGTTTAA CACAC TGACCAG TGTGTG TGGCCAGTCC TTGGATTGGCAACATGC ACTA
-----GCAA AATA GCACGACTCGTGAAGTCGTTTAA CACAT CGACAGA GGTGTG TGGTCAGTCA TTGGATTGGCCACACGC CCTT
TCGAAACTCGC A AATA GCACGACTCGTGAAGTCGTTTAA CACAT CGACAGA GGTGTG TGGTCAGTCA TTGGATTGGCCACCTGC CCTT
TCGAAACCTGCC AACA GCACGACACGCGAACCTGTTTAT CACAT GCGCAAG TGTGTG TGGTCAGTCA TTGGATTGGCAACATGC CCTT
TCGAAACCTGCC AACA GCACGACACGCGAACCTGTTTAT CACAT GCGCAAG TGTGTG TGGTCAGTCA TTGGATTGGCAACATGC CCTT
-----GTTTAA CACATGCGCAAGCGTGTGTG GTCAGTCC TCGTATTGACAAATGC CCTT
TCGAAACCTGCC GACA GCACGACGCGGAAACCCGTTTAA CACATGCGCAAGCGTGTGTG GTCAGTCC TCGGATTGACAAATGC CCTT
TCGAAACCTGCC AACA GCACGACACGCGAACCTGTTTAA CACA TGCCA G TGTGTG TGGTCAGTCA TTGGATTGACAAATGC CCTT
TCGTAACTGCC CACA GCACGACACGCGAACCTGTTTAA CACAT GTGCGAG TGTGTGTGTTGTCAGTCC TTGGATTGACGGCATGC CCTC
TCGAAACCTGCC AACA GCACGACACGCGAACCTGTTTAA CACAT GTGCA G TGTGTGTGTTGTCAGTCC TTGGATTGACGGCATGC CCTT
TCGAAACCTGCC AGCA GCACGACCTGTGAAGTCGTTTAC CACAT TTACAAG TGCGTG TGGTCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AACA GCACGACCTGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCA TTGGATTGGCAACACGC CCTT
TCGA CTGCC ACA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCA TTGGATTGGCAACACGC CCTT
TCGAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCA TTGGATTGGCAACACGC CCTT
TCGAAACCTGCC AACA GCATGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCA TTGGATTGGCAACACGC CCTT
TCGAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCA TTGGATTGGCAACACGC CCTT
TCGAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CACAC TGACAAG GGCGTG TGGTCAGTCA TTGAATTGGCAACACGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CACAT TGACAAG GGCGTG TGGCCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AACA GCACGACCCGTGAAGTCGTTTAA CCCAT TGACAAG GGTGTT TTGTCAGTCA CTAGATTGGCGACACGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CACAT TGACAAG GGCGTG TGGTCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AATA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGTGTT TGGTCAGTCC TCGGATTGGCAACACGC CCTT
TCGAAACCTGCCCAACA GCACGACCCGTGAAGTCGTTTAA CACAT CGACAAG GGTGTT TCGTCATTCA TTGGATTGGCAACACGC CCTT
TCGAAACCTGCCCAACA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TCGTCATTCA TTGGATTGGCAACACGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CACAT TGACAAG GGCGTG TGGCCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CACAT TGACAAG GGCGTG TGGCCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CACAT TGACAAG GGCGTG TGGCCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CACAT TGACAAG GGCGTG TGGTCAGTCC TCGGATTGGCAACATGC CCTT
-----CCGTTGAAGTCGTTTAA CCCAT TAC AG GGTGTG GTCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAA CACAT TGACAAG GGCGTG TGGTCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA GCACGACCTGTGAAGTCGTTTAA CATAT GGACAAG GGCGTG CGGCAATCCTCATCGGGTTGGAAACACGC CGTT
TCGAAACCTGCC AGCA GCACGACCCGTGAAGTCGTTTAC CGCAT TGACAAG GGCGTG TGGCCAGTCC TCGGATTGGCAACATGC CCTT
TCGAAACCTGCC AGCA ACACAACCCGTGAAGCCGTTTAT TGTAT CGACGGGGTTCG TTGTAATCC TTGGATTGACGACATTCCTCCGA
```

	110	120	130	140	150	160	170	180	190	200
<i>G. acuminata</i> AB110796	GTAAGTC	AAGGGGTG	CCCATTGCTCATGTAGCGTATGGCTCCTCAAGACG	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	TGTTG			
<i>G. acuticosta</i> BB01	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. atroviridis</i> AF367211	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATTG			
<i>G. atroviridis</i> AB110797	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATTG			
<i>G. balica</i> 021	GTAAGTC	AAGAGGTG	CCCATTGCTCATGTAGCGTATGGCTCCTCAAGACG	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	TGTTG			
<i>G. bancana</i> AF367212	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACA	TAACA	CACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATTG			
<i>G. cataractalis</i> 077	GTAAGTC	AAGAGGTG	CCCATTGCTCATGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. celebica</i> 002	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 003	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 005	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 009	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 023	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 027	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 028	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 033	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 038	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 043	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 054	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 059	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> 062	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> AB110816	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> AB110813	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> AB110814	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. celebica</i> AF367214	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCA			
<i>G. costata</i> BL03	GTGAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATACGGCTCCCAAGACG	CAACA	AACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATAG			
<i>G. costata</i> BL06	GTGAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATACGGCTCCCAAGACG	CAACA	AACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATAG			
<i>G. costata</i> AB110798	GTAAGTC	GAGGGGTG	CCAACGGCTCATGCAGCCTATGGCTCCTCAAGACAA	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	AATAA			
<i>G. cowa</i> 004	GTAAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	TAACA	CACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATTG			
<i>G. cowa</i> 013	GTAAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	TAACA	CACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATTG			
<i>G. cowa</i> AB110799	GTAAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	TAACA	CACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATTG			
<i>G. cowa</i> AF367216	GTAAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	TAACA	CACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TAATG			
<i>G. cymosa</i> 037	GTAAGTC	GAGGGGTG	CCAATTGCTCACGTGGCATTTGGCTCCTCAAGACG	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	TGTCG			
<i>G. cymosa</i> 042	GTAAGTC	GAGGGGTG	CCAATTGCTCACGTGGCATTTGGCTCCTCAAGACG	CAACA	AACCAA	TCCCGCGCGGGATGCGCAAGGAAT	TGTTT			
<i>G. diospyrifolia</i> var. <i>arborea</i> BB02	GTAAGTC	AAGGGGTG	CCCATTGCTCATGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. diospyrifolia</i> var. <i>arborea</i> BB05	GTAAGTC	AAGGGGTG	CCCATTGCTCATGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. diospyrifolia</i> var. <i>diospyrifolia</i> 078	GTAAGTC	AAGGGGTG	CCCATTGCTCATGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. diospyrifolia</i> var. <i>minor</i> AF36	GTAAGTC	AAGGGGTG	CCCAATGCTCATGTAGCATGTGGCTCCTCAAGATG	CAACA	AACCAA	TATCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. diospyrifolia</i> var. <i>minor</i> BB06	GTAAGTC	AAGGGGTG	CCCATTGCTCATGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. discoidea</i> SL01	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. discoidea</i> SL02	GTAAGTC	AAGAGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCGAGATG	CAACA	AACCAA	TCTCGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. dulcis</i> 050	GTAAGTC	AAGGGGTG	CCAATCGCTCATGTAGCATGTGGCTCCTCAAGACA	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	AATCG			
<i>G. dulcis</i> AB110800	GTAAGTC	GAGGGGTG	CCAATGGCTCATGCAGCCTTCGGCTCCTCAAGACA	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	AATCA			
<i>G. dulcis</i> AB110801	GTAAGTC	GAGGGGTG	CCAATGGCTCATGCAGCCTTCGGCTCCTCAAGACA	CAACA	AACCAA	TCCCGCGCGGGTTCGCGCAAGGAAC	AATCA			
<i>G. echinocarpa</i> 035	GTAAGTC	TAGGGGTG	CCCATTGCTCATGTAGCATGTGGCTCCTCAAGACG	CAACA	AACCAA	TTTGGGCGCGGGTTCGCGCAAGGAAC	TATCG			
<i>G. forbesii</i> AF367224	GTAAGTC	GAGGGGTG	CCCATTGCTCCTGTAGCATGTGGCTCCTCAAGACG	TAACA	CACCAA	TCCCGCGCGGGATGCGCAAGGAAC	TATTA			

*G. forbesii* AF67223  
*G. globulosa* AF367221  
*G. hanburyi* AB110802  
*G. laterifolia* 040  
*G. latissima* 026  
*G. latissima* AF479789  
*G. madruno* AJ509215  
*G. maingayi* 098  
*G. malaccensis* AB110810  
*G. malaccensis* AB110811  
*G. malaccensis* var. *malaccensis* BB04  
*G. malaccensis* var. *malaccensis* BB08  
*G. malaccensis* var. *malaccensis* BB09  
*G. malaccensis* var. *pseudomangostana* SP01  
*G. mangostana* 096  
*G. mangostana* AB110807  
*G. mangostana* AB110808  
*G. mangostana* AB110809  
*G. mangostana* AB110810  
*G. mangostana* AB110811  
*G. mangostana* AF367215  
*G. mangostana* AJ509214  
*G. mangostana* LAO  
*G. megaphylla* 030  
*G. megaphylla* 041  
*G. nervosa* AF367222  
*G. nigrolineata* AF367216  
*G. nitidall19*  
*G. penangiana*006  
*G. penangiana* AF367226  
*G. penangiana*BB07  
*G. picrorhiza*039  
*G. porrecta* AB110812  
*G. prainiana* AF367218  
*G. rigida* 029  
*G. rigida* 045  
*G. rigida* 049  
*G. rigida* 057  
*G. rostrata* 007  
*G. rostrata* 091  
*G. rostrata* AB110813  
*G. rostrata* AF367225  
*G. rostrata* BL051  
*G. schomburgkiana* AB110814  
*G. scortechinii* AF367219  
*G. scortechinii* 012  
*G. scortechinii* 089

GTAAGTC GAGGGGTG--CCCATTGCTCTTGTAGCACATGGC--CTGAAGACG TAATAACACCAA TCCCGGCGCGGGATGCGCCAAGGAAC--TATTG  
GTAAGTC GAGGGGTG--CCCATTGCTCTTGTAGCACATGGCTCCTCAAGACG TAACA-CACCAA TCCCGGCGCGGGATGCGCCAAGGAAC--TATTG  
GTAAGTC AAGGGGTG--CCCATCGCTCATGTAGCGTATGGCTCCTCAAGACG CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TGTTG  
GTAAGTC GAGGGGTG--CCCATCGCTCATGTAGCGTATGGCTCCTCAAGATG CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTG  
GTAAGTC GAGGGGTG--CCCATCGCTCATGTAGCGTATGGCTCCTCAAGACG CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TGTTG  
GTGAGTC GAGGGGTG--TCGATTGCTCATGCAGCAT-TGGCTCCTCAAGACA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAG--AATAA  
GTGAGTC GAGGGGTG--TCGATTGCTCATGCAGCAT-TGGCTCCTCAAGACA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAG--AATAA  
GTAAGTC GAGGGGTG--GCCGTGCTCTCGTAGCATATGGCTCCCAAGACG TAACA-AACCAA TCTCGGCGCGGGATGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCTCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCTCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCAATGGCTCATGCAGCCTTTGGCTCCTCAAGACAA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--AATCA  
GTAAGTC GAGGGGTG--CCAATCGCTCATGCAGC-TTTGGCTCCTCAAGACA CAACAACACCAAAT TCCCGGCGCGGGTTGCGCCAAGGAAC--AATCG  
GTAAGTC GAGGGGTG--CCCATTGCTCTTGTAGCACATGGCTCCTCAAGACG TAACA-CACCAA TCCCGGCGCGGGATGCGCCAAGGAAC--TAATG  
GTAAGTC AAGGGGTG--CCCATCGCTCCCGTAGCATGTGGCTCCTCGAGATG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATTGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATTGCTCCCGTAGCATGTGGCTCCTCGAGACG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATTGCTCCCGTAGCATGTGGCTCCTCGAGATG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TGTTG  
GTAAGTC GAGGGGTG--CCCATTGCTCTTGTAGCACATGGCTCCTCAAGACG TAATAACACCAA TCCCGGCGCGGGATGCGCCAAGGAAC--TATTG  
GTAAGTC GAGGGGTG--CCCATTGCTCATGCAGCATTTGGCTCCTCAAGACA GAAACA-AACCAA TACCGGCGCGGGTTGCGCCAAGGAAC--TATTG  
GTAAGTC AAGGGGTG--CCCATCGCTCCTGTAGCATGTGGCTCCTCGAGATG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCTGTAGCATGTGGCTCCTCGAGATG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC AAGGGGTG--CCCATCGCTCCTGTAGCATGTGGCTCCTCAAGATG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCG  
GTAAGTC GAGGGGTG--TTGAGTGCTCGCATGGCATTTGGCTCCTCGAGACA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTG  
GTAAGTC GAGGGGTG--TCGAGTGCTCGCTTGGCATTCGGCTCCTCGAGACA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTA  
GTAAGTC AAGAGGTG--CCCATCGCTCCTGTAGCATGTGGCTCCTCGAGATG CAACA-AACCAA TCTCGGCGCGGGTTGCGCCAAGGAAC--TATCA  
GTAAGTC GAGGGGTG--TCGAGTGCTCGCTTGGCATTCGGCTCCTCGAGACA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTA  
GTAAGTC GAGGGGTG--TCGAGTGCTCGCATGGCATTTGGCTCCTCGAGACA CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTG  
GTAAGTC GAGGGGTG--CCCATTGCTCTTGTAGCAGATGGCTCCTCAAGACG TAACA-CACCAA TCCCGGCGCGGGATGCGCCAAGGAAC--TATTG  
GTAAGTC GAGGGGTG--CCCATCGCTCATGTAGCGTATGGCTCCTCAAGACG CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTG  
GTAAGTC CAGGGGTG--CCCATCGCTCATGTAGCGTATGGCTCCTCAAGACG CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTG  
GTAAGTC CAGGGGTG--CCCATCGCTCATGTAGCGTATGGCTCCTCAAGACG CAACA-AACCAA TCCCGGCGCGGGTTGCGCCAAGGAAC--TATTG



	210	220	230	240	250	260	270	280	290	300
	..... ..... ..... ..... ..... ..... ..... ..... ..... ..... .....									
<i>G. acuminata</i> AB110796	TAAATGTGA	GGAACTCCTCTC	CGTCTT	GGAAACAATGCACGTGATGAGGGCTTGTT	---	CCTTTCAGTCGT	AAACAAAATGACTCTCGGCAACGG			
<i>G. acuticosta</i> BB01	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAAGTACGTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. atroviridis</i> AF367211	TAAATGTAAAGGAACTCCTCTC	CGTCTT	GGAAACAATGATGATGATGAGGGCTTGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGGCAACGG				
<i>G. atroviridis</i> AB110797	TAAATGTAAAGGAACTCCTCTC	CGTCTT	GGAAACAATGATGATGATGAGGGCTTGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGGCAACGG				
<i>G. balica</i> 021	AAAAATGTA	GGAACTCCTCCC	CGTCTT	GGAAACAATGCACGTGATGAGGGCTTGTT	---	CCTTTCAGTCGT	AAACAAAACGACTCTCGGCAACGG			
<i>G. bancana</i> AF367212	TAAATGTAAAGGAACTCCTCTC	CATCTT	GGAAACAATGAATGTGACGAGGGCTTGTT	---	CCTTTTAGTTGG	AAACAAAACGACTCTCGGCAACGG				
<i>G. cataractalis</i> 077	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTAGGTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. celebica</i> 002	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 003	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 005	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 009	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 023	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 027	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 028	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 033	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 038	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 043	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 054	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 059	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> 062	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> AB110816	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> AB110813	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> AB110814	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. celebica</i> AF367214	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. costata</i> BL03	TAAATGTAAAGGAACTCCTCTC	CATCTT	GGAAACAACGTATGTGACGAGGGCTTGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGCAACGG				
<i>G. costata</i> BL06	TAAATGTAAAGGAACTCCTCTC	CATCTT	GGAAACAACGTATGTGACGAGGGCTTGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGCAACGG				
<i>G. costata</i> AB110798	AAAACTCAA	GGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. cowa</i> 004	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAATGACTCTCGGCAACGG			
<i>G. cowa</i> 013	TAAATGTAAAGGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGGCAACGG				
<i>G. cowa</i> AB110799	TAAATGTAAAGGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGGCAACGG				
<i>G. cowa</i> AF367216	TAAATTTAAAGGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTTAGTCGT	AAACAAAACGACTCTCGGCAACGG				
<i>G. cymosa</i> 037	TAAATGTGA	GGAACTCCTCTC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	AAACAAAACGACTCTCGGCAACGG			
<i>G. cymosa</i> 042	TAAATGTGA	GGAACTCCTCTC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	AAACAAAACGACTCTCGGCAACGG			
<i>G. diospyrifolia</i> var. <i>arborea</i> BB02	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. diospyrifolia</i> var. <i>arborea</i> BB05	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. diospyrifolia</i> var. <i>diospyrifolia</i> 078	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. diospyrifolia</i> var. <i>minor</i> AF36	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. diospyrifolia</i> var. <i>minor</i> BB06	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. discoidea</i> SL01	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. discoidea</i> SL02	TAAATGTAA	GGAACTCCTCCC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			
<i>G. dulcis</i> 050	AAAAACGCAA	GGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTTAGTTGT	AAACAAAACGACTCTCGGCAACGG			
<i>G. dulcis</i> AB110800	AAAACTCAA	GGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTTAGTTGT	AAACAAAACGACTCTCGGCAACGG			
<i>G. dulcis</i> AB110801	AAAACTCAA	GGAACTCCTCTC	CATCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTTAGTTGT	AAACAAAACGACTCTCGGCAACGG			
<i>G. echinocarpa</i> 035	TAAATGTAA	GGAACTCCTCTC	CGTCTT	GGAAACAAGTACTTGATGAGGGCTGGTT	---	CCTTTCAGTCGT	TAACAAAACGACTCTCGGCAACGG			

*G. forbesii* AF367224 TAAATGTAAAAGGAACTCCTCTC-CATCTT GGAAACAATGAATGTGACGAGGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. forbesii* AF67223 TAAATGTAAAAGGAACTCCTCTC-CATCTT GGAAACAATGAATGTGACGAGGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. globulosa* AF367221 TAAATGTAAAAGGAACTCCTCTC-CATCTT GGAAACAATGAATGTGACGAGGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. hanburyi* AB110802 TAAATGTGA--GGAACCTCTCTC-CGTCTT GGAAACAATGCACGTGATGAGGGCTTGTT--CCTTCAGTCGT AAACAAAATGACTCTCGGCAACGG  
*G. laterifolia* 040 TAAATGTGA--GGAACCTCTCTC-CGTCTT GGAAACAATGCACGTGATGAGGGCTTGTT--TCTTCAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. latissima* 026 AAAATGTGA--GGAACCTCTATC-CGTCTT GGAAACAATGCACGTGATGAGGGCTTGTT--CCTTCAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. latissima* AF479789 AAAATATGG--GGAACCTCTCTC-CATCCT GGAAACAGTGTATGCGACGAGGGATTGTT--CCTTTTAGTCGT AAATAGAACGACTCTCGGCAACGG  
*G. madruno* AJ509215 AAAATATGA--GGAACCTCTCTC-CATCCT GGAAACAGTGTATGCGACGAGGGATTGTT--CCTTTTAGTCGT AAATAAAACGACTCTCGGCAACGG  
*G. maingayi* 098 TAAATGTAAAAGGAACTCCTCTC-CGTCTT GGAAACAGTGTACGTGACGAGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. malaccensis* AB110810 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. malaccensis* AB110811 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. malaccensis* var. *malaccensis* BB04 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. malaccensis* var. *malaccensis* BB08 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. malaccensis* var. *malaccensis* BB09 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. malaccensis* var. *pseudomangostana* SP01 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* 096 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AB110807 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AB110808 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AB110809 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AB110810 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AB110811 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AF367215 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. mangostana* AJ509214 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. mangostana* LAO TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. megaphylla*030 AAAATATGA--GGAACCTCTCTC-CATCCT GGAAACAGTGTATGCGACGAGGGATTGTT--CCTTTTAGTCGT AAATAGAACGACTCTCGGCAACGG  
*G. megaphylla*041 AAAACTCAA--GGAACCTCTCTC-CATCCT GGAAACAGTGTATGCGACGAGGGCTTGTT--CCTTTTAGTGT AAATCAAACGACTCTCGGCAACGG  
*G. nervosa*AF367222 AAAACGCAA--GGAACCTCTCTC-CATCCT GGAAACAGTGTATGCGACGAGGGCTTGTT--CCTTTTAGTGT AAACAAAACGACTCTCGGCAACGG  
*G. nigrolineata*AF367216 TAAATTTAAAAGGAACTCCTCTC-CATCTT GGAAACAATGAATGTGACGAGGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. nitida*119 TAAATGTAA--GGAACCTCTCTC-CGTCTT GGAAACAAGTACGTGATGAGGGCTGGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. penangiana*006 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCATT--CCTTCAGTTGTTAAACAAAACGACTCTCGGCAACGG  
*G. penangiana* AF367226 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCATT--CCTTCAGTTGTTAAACAAAACGACTCTCGGCAACGG  
*G. penangiana*BB07 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAATGTACGTGATGAGGGCTCATT--CCTTCAGTTGTTAAACAAAACGACTCTCGGCAACGG  
*G. picrorhiza*039 TAGAAGTAA--GGAACCTCTTTA-TGTCTT GGAAACAGTGTACATGATGAGGGCTTGTT--CCTTTTACTGGT AAACGAAATGACTCTCGGCAACGG  
*G. porrecta* AB110812 TAAATGTAAAAGGAACTCCTCTC-CATCTT GGAAACAATGAATGTGACGAGGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. prainiana* AF367218 TAAATGTGA--GGAACCTCTCTC-CGTCTT GGAAACAGATGACGCGATGAGGGTATGTC--CCTTTTAGTCAT AAACAAAACGACTCTCGGCAACGG  
*G. rigida* 029 TAAATGTAA--GGAACCTCTCCC-CGTCTTTGGAAACAAAGTACGTGATGAGGGCTGGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. rigida* 045 TAAATGTAA--GGAACCTCTCCC-CGTCTTTGGAAACAAAGTACGTGATGAGGGCTGGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. rigida* 049 TAAATGTAA--GGAACCTCTCCC-CGTCTTTGGAAACAAAGTACGTGATGAGGGCTGGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. rigida* 057 TAAATGTAA--GGAACCTCTCCC-CGTCTTTGGAAACAAAGTACGTGATGAGGGCTGGTT--CCTTCAGTCGTTAAACAAAACGACTCTCGGCAACGG  
*G. rostrata* 007 TAAA-GTGAAGGAACTCCTCTCCTGTCTT GGAAACAATGAACCGCAGGGCATGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. rostrata* 091 TAA--GTGAAGGAACTCCTCTCTCGTCTT GGAAACAATGAACCGTATGAGGGCATGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. rostrata* AB110813 TAAATGTAA--GGAACCTCTCCC-CGTCTT GGAAACAAGTACGTGATGAGGGCTGGTT--CCTTCAGTCGTTAAACAAAATGACTCTCGGCAACGG  
*G. rostrata* AF367225 TAAAGTAAA--GGAACCTCTCTCTCGTCTT GGAAACAATGAACCGTATGAGGGCATGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. rostrata* BL051 TAAA-GTGAAGGAACTCCTCTCCTGTCTT GGAAACAATGAACCGCAGGGCATGTT--CCTTTTAACTGT AAACAAAACGACTCTCGGCAACGG  
*G. schomburgkiana* AB110814 TAAATGTAAAAGGAACTCCTCTC-CATCTT GGAAACAATGAATGTGACGAGGGCTTGTT--CCTTTTAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. scortechinii* AF367219 TAAATGTGA--GGAACCTCTCTC-CGTCTT GGAAACAATGCACGTGATGAGGGCTTGTT--TCTTCAGTCGT AAACAAAACGACTCTCGGCAACGG  
*G. scortechinii* 012 TAAATGTGA--GGAACCTCTCTC-CGTCTT GGAAACAATGCACGTGATGAGGGCTTGTT--CCTTCAGTCGT AAACAAAACGACTCTCGGCAACGG









ITS2

	410	420	430	440	450	460	470	480	490	500			
	..... ..... ..... ..... ..... ..... ..... ..... ..... ..... .....												
<i>G. acuminata</i> AB110796	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ACTGT	TGCCAAAT	CCGAA	TTGTTA	TGGA	ATTTGGGCG	
<i>G. acuticosta</i> BB01	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAA	TTTCTA	CGGT	GTTTGGGCG	
<i>G. atroviridis</i> AF367211	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	GCCCAAAAC	CACAA	TTGTCA	GGGT	ATTTGGGTG	
<i>G. atroviridis</i> AB110797	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	GCCCAAAAC	CACAA	TTGTCC	GGGT	ATTTGGGCG	
<i>G. balica</i> 021	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAT	CCCAA	TTGTTA	TGGA	ATTTGGGCG	
<i>G. bancana</i> AF367212	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CCCAA	TTGCTG	TGGT	ATTTGGGCG	
<i>G. cataractalis</i> 077	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. celebica</i> 002	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 003	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 005	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 009	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	ACCAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 023	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 027	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 028	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 033	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 038	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 043	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 054	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 059	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> 062	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> AB110816	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	ACCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> AB110813	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> AB110814	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAT	AAATGT	TGCCAAAC	CCCAA	TTGTTA	TGGT	GTTTGGGCG	
<i>G. celebica</i> AF367214	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	ACCAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. costata</i> BL03	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CACAG	TTGCTA	TGGT	ATTTGGGAG	
<i>G. costata</i> BL06	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CACAG	TTGtTA	TGGT	ATTTGGGAG	
<i>G. costata</i> AB110798	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCAAA	TTATCC	TGGT	ATAAGGGCG	
<i>G. cowa</i> 004	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CCCAA	TTGTTG	TGGT	ATTTGGGCG	
<i>G. cowa</i> 013	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CCCAA	TTGTTG	TGGT	ATTTGGGCG	
<i>G. cowa</i> AB110799	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CCCAA	TTGTTG	TGGT	ATTTGGGCG	
<i>G. cowa</i> AF367216	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CCCAA	TTGCTG	TGGT	ATTTGGGCG	
<i>G. cymosa</i> 037	CCCAAAGCC	TTATGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCCAA	C	CCAAA	ATGCAA	GGGG	ATTTGGGCG
<i>G. cymosa</i> 042	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	ATTGTT	TGCCAAAC	CCAAA	ATGCAA	GGGA	ATTTGGGCG	
<i>G. diospyrifolia</i> var. <i>arborea</i> BB02	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCGAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. diospyrifolia</i> var. <i>arborea</i> BB05	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCGAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. diospyrifolia</i> var. <i>diospyrifolia</i> 078	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCGAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. diospyrifolia</i> var. <i>minor</i> AF367227	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. diospyrifolia</i> var. <i>minor</i> BB06	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. discoidea</i> SL01	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCATGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAT	TTTCTA	CGGT	GTTTGGGCG	
<i>G. discoidea</i> SL02	CCCAAAGCA	TTTTGG	TTGAGGGC	ACGTC	GCATGGG	TGTCACAC	AAATGT	TGCCAAAC	CCCAT	TTTCTA	CGGT	GTTTGGGCG	
<i>G. dulcis</i> 050	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGTT	TGCCAAAC	CCAGA	TTGTGG	TGGG	GTGAGGGCG	
<i>G. dulcis</i> AB110800	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGTT	TGCCAAAC	CCAAA	TTATCC	TGGG	ATAAGGGCG	
<i>G. dulcis</i> AB110801	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	AAATGTT	TGCCAAAC	CCAAA	TTATCC	TGGG	ATAAGGGCG	
<i>G. echinocarpa</i> 035	CCCAAAGCC	TTTTGG	TTGAGGGC	ACGTC	GCCTGGG	TGTCACAC	TGT	TGCCAAAT	TCCAA	TTGCTA	TGGG	ATATGGGCG	

<i>G. forbesii</i> AF367224	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAAC	CCCAA	TTG	TG	TGGT	GTTTGGGCG
<i>G. forbesii</i> AF67223	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAAC	CCCAA	TTGCTG	TG	TGGT	GTTTGGGCG
<i>G. globulosa</i> AF367221	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTG	TG	TGGT	ATTTGGGCG
<i>G. hanburyi</i> AB110802	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACACTGT	TGCCCAAAT	CCGAA	TTGTTA	TGGA	ATTTGGGCG	
<i>G. laterifolia</i> 040	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAT	CCAAA	TTGTTG	TGGA	ATTTGGGTTG	
<i>G. latissima</i> 026	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAAT	CCCAA	TTGTTA	TGGA	ATTTGGGCG	
<i>G. latissima</i> AF479789	CCCGAAGCC TTCAGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAG	CCCAA	CTGGTA	TTGG	ATTTGGGCG	
<i>G. madruno</i> AJ509215	CCCGAAGCC TTCAGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAG	CCCAA	CTGGTA	TTGG	ATTTGGGCG	
<i>G. maingayi</i> 098	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAACC	CCCAG	TTGCTTA	TGGT	ATTTGGGCG	
<i>G. malaccensis</i> AB110810	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. malaccensis</i> AB110811	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. malaccensis</i> var. <i>malaccensis</i> BB04	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. malaccensis</i> var. <i>malaccensis</i> BB08	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. malaccensis</i> var. <i>malaccensis</i> BB09	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. malaccensis</i> var. <i>pseudomangostana</i> SP01	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> 096	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AB110807	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AB110808	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AB110809	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AB110810	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AB110811	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AF367215	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> AJ509214	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. mangostana</i> LAO	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. megaphylla</i> 030	CCCGAAGCC TTCAGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	CTGGTA	TTGG	ATATGGGCG	
<i>G. megaphylla</i> 041	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCAAA	TTATCG	TGGG	ATAAGGGCG	
<i>G. nervosa</i> AF367222	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCAA	TTGTCG	TGGG	ATAAGGGCG	
<i>G. nigrolineata</i> AF367216	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTG	TGGT	ATTTGGGCG	
<i>G. nitida</i> 119	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. penangiana</i> 006	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCTAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. penangiana</i> AF367226	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCTAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. penangiana</i> BB07	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCTAA	TTGCTA	CGGT	GTTTGGGCG	
<i>G. picrorhiza</i> 039	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCGCACAATGTT	TGCCCAAAC	CCAAG	TCATAA	TGGG	ATTTGGGCG	
<i>G. porrecta</i> AB110812	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAAC	CCCAA	TTGCTG	TGGT	GTTTGGGCG	
<i>G. prainiana</i> AF367218	CCCAAAGCCTTA TGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAAC	CCAAA	TTGCAA	GGGG	ATTTGGGCG	
<i>G. rigida</i> 029	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	TGGT	GTTTGGGTTG	
<i>G. rigida</i> 045	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	TGGT	GTTTGGGTTG	
<i>G. rigida</i> 049	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	TGGT	GTTTGGGTTG	
<i>G. rigida</i> 057	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCCAA	TTGCTA	TGGT	GTTTGGGTTG	
<i>G. rostrata</i> 007	CCCGAAGCC TTTCCGGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	AGCCCAAAC	CCAAA	ACGCAAA	AGGGGATTT	GGGCA	
<i>G. rostrata</i> 091	CCCGAAGCC TCTCCGGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCAAA	ATGCAAA	TGGG	ATTTGGGCG	
<i>G. rostrata</i> AB110813	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	ACCAA	TTGCTA	TGGT	GTTTGGGCG	
<i>G. rostrata</i> AF367225	CCCGAAGCC TCTCCGGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCAAA	ATGCAAA	TGGG	ATTTGGGCG	
<i>G. rostrata</i> BL051	CCCGAAGCC TTTAGGGG	TTGAGGGCACGTCCTGGGTGTCACACAATGTT	TGCCCAAAC	CCAAA	ACGCAAA	AGGGGATTT	GGGCG	
<i>G. schomburgkiana</i> AB110814	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACATTGTT	TGCCCAAAC	CCCAA	TTGCTG	TGGT	ATTTGGGCG	
<i>G. scortechinii</i> AF367219	CCCAAAGCCTTTTGG	TTGAGGGCACGTCCTGGGTGTCACACACTAT	TGCCCAAAT	CCAAA	TTGTTA	TGGA	ATTTGGGTTG	
<i>G. scortechinii</i> 012	CCCAAAGCC TTT GG	TTGAGGGCACGTCCTGGGTGTCACACACTGT	AGCCCAAAT	CCAAA	T GTTA	TGGA	ATTTGGGCG	

*G. scortechinii* 089  
*G. scortechinii* K04  
*G. sp.* AJ312607  
*G. sp.*B1 008  
*G. sp.*B2 011  
*G. sp.*D1 064  
*G. sp.*D1 070  
*G. sp.*D2 P01  
*G. sp.*D3 P02  
*G. sp.*D4 BL04  
*G. sp.*D5 BL052  
*G. sp.*D6 K12  
*G. spicata* 019  
*G. sp.*M1 025  
*G. sp.*M2 068  
*G. sp.*M3 072  
*G. sp.*M4 022  
*G. sp.*M5 069  
*G. sp.*M5 073  
*G. sp.*M6 093  
*G. sp.*M6 094  
*G. sp.*X1 052  
*G. sp.*X2 080  
*G. syzygifolia* AB110815  
*G. tinctoria* AB110817  
*G. trianaii* K02  
*G. urophylla* 001  
*G. urophylla* 081  
*G. vilersiana* AB110818  
*G. xanthochymus* AB110819  
*G. xanthochymus* 034  
*G. xanthochymus* 092  
*G. xanthochymus* BB03  
*G. xanthochymus*cf 032  
*Symphonia globuliera* AY682919  
*Tripetalum cymosum* AY145206  
*Clusia rosea*

CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCAAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTAGGCG  
 CCCGAAGCC TTCAGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAG--CCCAA-----CTGGTA-TTGG-ATATGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACATTGTT--TGCCCAAAC--CCCAA-----TTGCTG-TGGT-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCGAAC--CCAAA-----ATGCAAAATGGG-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCGAAC--CCAAA-----ATGCAAAATGGG-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTGATA-TGGT-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----ATGCAAAATGGG-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----ATGCAAAATGGG-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----ATGCAAAATGGG-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----ATGCAAAATGGG-ATTTGGGCG  
 CCCGAAGCCTCTCGGGGTTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TTCCCAAC--CACAG-----TTGATA-TGGT-ATTTGGGAG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATATT--TGCCCAAAC--CCAAA-----TCGTTG-TGGG-ATAAGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACATTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACATTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCCAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTATCG-TGGG-ATAAGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTGTCG-TGGG-ACAAGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCCAA-----TTGCTA-TGGT-GTTGGGTG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTATCG-TGGG-ATAAGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTGTTA-TGGA-ATTTGGGCA  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACACTGT--TGCCCAAAT--CCAAA-----TTGTTA-TGGA-ATTTGGGCG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTATCG-TGGG-ATAAGGGCG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTATCG-TGGG-ATAAGGGCG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTGTCG-TGGG-ACAAGGGCG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTGTCG-TGGG-ACAAGGGCG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAGA-----TTGTTG-TGGG-ATAAGGGCG  
 CCCAAAGCCTTTCGG--TTGAGGGCACGTCTGCCTGGGTGTCACGCAATGT--CGCCAAATA--CCAAACGGCTACGAATTGCTACTGG--ATTTGGGCG  
 CCCAAGGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAATGT--TGCCCAAAC--CCAAA-----TTATCGTTGG--ACAAGGGCG  
 CCCAAAGCCTTTTGG--TTGAGGGCACGTCTGCCTGGGTGTCACACAAGT--CGCCCAA--G--CCCAA-----CATCCAGTGGG-AATAGGGCG

	510	520	530	540	550	560	570	580	590	600
<i>G. acuminata</i> AB110796	GATGTTGGTCTCCCGTGTGCCCTT	CAGTTGCGGTTGGCCATAAT	GCGAGCCTTGTGACGTTG	TGCACGAC	AATCGGTGGTTT	ATAGACCCCTC				
<i>G. acuticosta</i> BB01	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. atroviridis</i> AF367211	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. atroviridis</i> AB110797	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. balica</i> 021	GATGTTGGTCTCCCGTGTGCCCTT	CGGTTGCGGTTGGCCAAAA	ACGAGCCTTGTGACATTG	TGCACGAC	AATCGGTGGTTT	ATAGACCCCTAT				
<i>G. bancana</i> AF367212	GTTGTTGGCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. cataractalis</i> 077	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACCAGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. celebica</i> 002	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 003	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 005	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 009	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. celebica</i> 023	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACATGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 027	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 028	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 033	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 038	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 043	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 054	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 059	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> 062	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> AB110816	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. celebica</i> AB110813	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> AB110814	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. celebica</i> AF367214	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ATGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. costata</i> BL03	GATGTTGGTCTCCCGTGCGCCCTT	TGCTCGCGGTTGGCCAAAT	ACGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. costata</i> BL06	GATGTTGGTCTCCCGTGCGCCCTT	TGCTCGCGGTTGGCCAAAT	ACGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				
<i>G. costata</i> AB110798	GATGTTGGTCTCCCGTGTGCCATT	TGCTCGCGGTTGGCCAAAC	ACAAGCCTCTTTGATGTTG	TACACGGC	AATCGGTGGTTT	TAAGACCCCTCG				
<i>G. cowa</i> 004	GTTGTTGGCCTCCCGTGCGCCCTT	TGCTCGCGGTTGGCCAAAT	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. cowa</i> 013	GTTGTTGGCCTCCCGTGCGCCCTT	TGCTCGCGGTTGGCCAAAT	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. cowa</i> AB110799	GTTGTTGGCCTCCCGTGCGCCCTT	TGCTCGCGGTTGGCCAAAT	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. cowa</i> AF367216	GTTGTTGGCCTCCCGTGCGCCCTT	TGCTCGCGGTTGGCCAAAT	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. cymosa</i> 037	GATGTTGGTCTCCCGTGTGCCCTT	CGCCCGCGGTTGGCCAAAT	ACGAGCCTTGTGATGTCG	TACACGGC	AATCGGTGGTTT	TAAGACCCCTCG				
<i>G. cymosa</i> 042	GATGTTGGTCTCCCGTGTGCCCTT	CGCCCGCGGTTGGCCAAAT	ACGAGCCTTGTGATGTCG	TACATGGC	AATCGGTGGTTT	AAAGACCCCTCG				
<i>G. diospyrifolia</i> var. <i>arborea</i> BB02	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACATGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. diospyrifolia</i> var. <i>arborea</i> BB05	GATGTTGGTCTCCCGTGTGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACATGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. diospyrifolia</i> var. <i>diospyrifolia</i> 078	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	AGGAGCCTTGTGATGTTG	TACATGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. diospyrifolia</i> var. <i>minor</i> AF367227	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. diospyrifolia</i> var. <i>minor</i> BB06	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. discoidea</i> SL01	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. discoidea</i> SL02	GATGTTGGTCTCCCGTGCGCCCTT	GGCTCGCGGTTGGCCAAAA	ACGAGCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ACAGACCCCTCG				
<i>G. dulcis</i> 050	GATGTTGGTCTCCCGTGTGCGTT	TGCTCGCGGTTGGCCAAAC	ACGAGCCTCTCTGATGTCG	TACACGGC	AATCGGTGGTTT	AAAGACCCCTCG				
<i>G. dulcis</i> AB110800	GATGTTGGTCTCCCGTGTGCCATT	TGCTCGCGGTTGGCCAAAC	ACAAGCCTCTCTGATGTTG	TACACGGC	AATCGGTGGTTT	AAAGACCCCTCG				
<i>G. dulcis</i> AB110801	GATGTTGGTCTCCCGTGTGCCATT	TGCTCGCGGTTGGCCAAAC	ACAAGCCTCTCTGATGTTG	TACACGGC	AATCGGTGGTTT	AAAGACCCCTCG				
<i>G. echinocarpa</i> 035	GATGTTGGTCTCCCGTGTG	CACC	CGCTCGCGGTTGGCCAAAT	ACGAGCCTATGTTGATGTTG	TTCACGGC	AATCGGTGGTTT	ATAGACCCCTCG			
<i>G. forbesii</i> AF367224	GTTGTTGGCCTCCCATGCCGCCCTT	CGCTCGGTTGGCCAAAT	ATGTGCCCTTGTGATGTTG	TACACGGC	AATCGGTGGTTT	ATAGACCCCTCG				





\*\*\*\*\*

	610	620	630	640	650	660	670	680
	.... .... .... .... .... .... .... .... .... .... .... .... .... ....							
<i>G. acuminata</i> AB110796	GACAATGTC	GTGAAGTC	CTGTGCTATC	GGCGTTAATGGCCCCA	ATGCAA	TCATTGCATGATC	CACAGAA	???????
<i>G. acuticosta</i> BB01	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAATGGCCCCC	ATGCAT	TCTTTGCATGATC	GAAAAAGCGACCCC	
<i>G. atroviridis</i> AF367211	GATAGTGTC	GTGCACTC	TCGTGCGATT	GGCATTAAAGGCCCCA	ATGCAA	TTATTGCATGATC	TTCAAA	?????
<i>G. atroviridis</i> AB110797	GATAGTGTC	GTGCACTC	TCGTGCGATT	GGCATTAAAGGCCCCA	ATGCAA	TTATTGCATGATC	TTCAAA	???????
<i>G. balica</i> 021	GACAATGTC	GTGCACTC	CTGTGCTATC	GGTGTAAATGGCCCCA	ATGCAA	TCATTGCATGATC	CACAGAGCGACCCC	
<i>G. bancana</i> AF367212	GATAGTGTT	GTGCACTC	TCGTCTCATT	GGCATTAAATGGCCCCA	ATGCAA	TCATTGCATGATC	CGTAAACAA	???
<i>G. cataractalis</i> 077	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAAGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 002	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 003	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	CAA-GC-ACCCC	
<i>G. celebica</i> 005	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	CAAC-CAACCC?	
<i>G. celebica</i> 009	GACAATGTT	GTGCACTC	TCATCGCATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 023	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCC	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCTC	
<i>G. celebica</i> 027	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 028	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 033	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 038	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 043	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 054	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 059	GACAATGTT	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCA	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> 062	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. celebica</i> AB110816	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAA	???????
<i>G. celebica</i> AB110813	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAA	???????
<i>G. celebica</i> AB110814	GACAATGTC	GTGCACTC	TCATCGAATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAA	???????
<i>G. celebica</i> AF367214	GACAATGTT	GTGCACTC	TCATCGCATT	GGCATTAAATGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAA	???????
<i>G. costata</i> BL03	GACAATGTC	GTGCACTC	TCGTGCGATT	GGCATTAAATGGCCCCG	ATGCAA	TCATTGCATGATC	TCGAAGCGACCCC	
<i>G. costata</i> BL06	GACAATGTC	GTGCACTC	TCGTGCGATT	GGCATTAAATGGCCCCG	ATGCAA	TCATTGCATGATC	TCGAAGCGACCCC	
<i>G. costata</i> AB110798	GACAACGTC	GTGCACTC	TCGTGCGATT	GGCATTAAATGGCCCCA	AAGCAA	TCGTTGCATGCTC	ATAAA	???????
<i>G. cowa</i> 004	GATAGTGTT	GTGCACTC	TCGTCTCATT	GGCATTAAATGGCCCCA	ATGCAA	TCATTGCATGATC	TCGTAGCGACCCC	
<i>G. cowa</i> 013	GATAGTGTT	GTGCACTC	TCGTCTCATT	GGCATTAAATGGCCCCA	ATGCAA	TCATTGCATGATC	TCGTAGCGACCCC	
<i>G. cowa</i> AB110799	GATAGTGTT	GTGCACTC	TCGTCTCATT	GGCATTAAATGGCCCCA	ATGCAA	TCATTGCATGATC	TCGTAGCGACCCC	
<i>G. cowa</i> AF367216	GATAGTGTT	GTGCACTC	TCGTCTCATT	GGCATTAAATGGCCCCA	ATGCAA	TCATTGCATGATC	TCGTAGCGACCCC	
<i>G. cymosa</i> 037	GACAATGTC	GTGCACTC	GCGTCACCTT	GGCATT--TGGCCCCA	ATGCGG	T--TCGCATGATCTCAA	GCGACCCC	
<i>G. cymosa</i> 042	GACTGTGCC	GTGCACTC	TCGTGCGATT	GGCCCT--TGGCCCCA	ATGCGG	TCGCTGCATGATAT	TGAAGCGACCCC	
<i>G. diospyrifolia</i> var. <i>arborea</i> BB02	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAAGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. diospyrifolia</i> var. <i>arborea</i> BB05	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAAGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. diospyrifolia</i> var. <i>diospyrifolia</i> 078	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAAGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. diospyrifolia</i> var. <i>minor</i> AF367227	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAAGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGA	???????
<i>G. diospyrifolia</i> var. <i>minor</i> BB06	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAAGGCCCCA	ATGCAT	TCTTTGCATGATC	GCAAAGCGACCCC	
<i>G. discoidea</i> SL01	GACAATGTC	GTGCACTC	TCATAGCATT	GGCA	??			
<i>G. discoidea</i> SL02	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATT	??			
<i>G. dulcis</i> 050	AACAACGTC	GTGCACTC	TCGTGCGATT	GGCACCAGTACCCCCA	GAGCAA	TCGTTGCATGCTC	ATAA-GCGACCCC	
<i>G. dulcis</i> AB110800	GATAGCGTC	GTGCACTC	TCGTGCGATT	GGCATTAAATGGCCCCA	AAGCAA	TCGTTGCATGCTC	ATAAA	???????
<i>G. dulcis</i> AB110801	GATAGCGTC	GTGCACTC	TCGTGCGATT	GGCATTAAATGGCCCCA	AAGCAA	TCGTTGCATGCTC	ATAAA	???????
<i>G. echinocarpa</i> 035	GACAATGTC	GTGCACTC	TCATCGCATT	GGCATTAAATGGCCCCA	ATGCAA	TCATTGCA	GATCA	?????????????

*G. forbesii* AF367224 GATAGTGTT GTGCACTC TCGTCTCATT GGCATTAATGGCCCTA-ATGCAA TCATTGCATACTATCGTAAACAA???

*G. forbesii* AF67223 GATAGTGTT GTGCACTC TCGTCTCATT GGCATTAATGGCCCTA-ATGCAA TCATTGCATACTATCGTAAACAA??

*G. globulosa* AF367221 GATAGTGTT GTGCACTC TCGTCTCATT GGCATTAATGGCCCTA-ATGCAA TCATTGCATACTATCGTAA???????

*G. hanburyi* AB110802 GACAATGTC GTGAAGTC CTGTGCTATC GGCCTTAATGGCCCTA-ATGCAA TCATTGCATGATCAGAGAA???????

*G. laterifolia* 040 GACAATGTC GTGAAGTC CTATCGCATC GGCCTTAATGGCCCTA-ATGCAA TCATTGCATGATCAGAAAGCGACCC?

*G. latissima* 026 GACAATGTC GTGCACTC CTGTGCGATC GGTGTTAATGGCCCTA-ATGCAA TCATTGCATGATCAGAGCGACCC

*G. latissima* AF479789 GACAATGTC GTGCACTCCTCATCGCTTT GGCACCAATGGGCCA-AAGCAAATCATTGCAAGCTCAG???????????

*G. madruno* AJ509215 GACAATGTC GTGCACTC TCATCGCTTT GGCACCAATGGGCCA-AAGCAAATCATTGCAAGCTCAGAAAGCGACCC

*G. maingayi* 098 GACAATGTC GTGCACTC TCTTCTCATT GGCCTTAATGGCCCTG-ATGCAA TCATTGCATGATCAGATCGACCC

*G. malaccensis* AB110810 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. malaccensis* AB110811 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. malaccensis* var. *malaccensis* BB04 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. malaccensis* var. *malaccensis* BB08 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. malaccensis* var. *malaccensis* BB09 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. malaccensis* var. *pseudomangostana* SP01 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. mangostana* 096 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. mangostana* AB110807 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. mangostana* AB110808 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. mangostana* AB110809 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. mangostana* AB110810 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. mangostana* AB110811 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. mangostana* AF367215 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. mangostana* AJ509214 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCATAGCGACCC

*G. mangostana* LAO GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCT-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. megaphylla* 030 GACAATGTC GTGCACTC TCATCGGTTT GGCACCAATGGGCCA-AAGCAAATCATTGCAAGCTCAGAAAGCGACCC

*G. megaphylla* 041 GACGACGTC GTGCACTC TCGTCACATT GGCACATGATACCCCA-AAGCAA TCGTTGCATGCTCATAA GCGACCC

*G. nervosa* AF367222 GACAACGTC GTGCACTC TCGTCACATT GGCACATTAACCCCA-A-GCAA TCGTTGCATGCTCATAAACA?????

*G. nigrolineata* AF367216 GATAGTGTT GTGCACTC TCGTCTCATT GGCATTAATGGCCCTA-ATGCAA TCATTGCATACTATCGTAA-ACAA???

*G. nitida* 119 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. penangiana* 006 GACAATGTT GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATTGCAAAAGCGACCC

*G. penangiana* AF367226 GACAATGTT GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATTGCAAAA???????

*G. penangiana* BB07 GACAATGTT GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAAGCGACCC

*G. picrorhiza* 039 GATAGTGTC GTGCACTC TTATCACCTT GGCCTCTCTG-CCCCA-ATGCAA TCATTGCATGATCTTGAAGTGACCC

*G. porrecta* AB110812 GATAGTGTT GTGCACTC TCGTCTCATT GGCATTAATGGCCCTA-ATGCAA TCATTGCATACTATCGTAA???????

*G. prainiana* AF367218 GACAATGCT GTGCACTC TTGTACATC GGCCT- -TGGCCCA-ATGCAAATGA-TGCATGATCCCAACACAA???

*G. rigida* 029 GACAATGTC GTGTACTC TCATCGCACT GGCATTAACGGCCTCA-ATGCAT TCTTTACATGATCGCAAAAGCGACCC

*G. rigida* 045 GACAATGTC GTGTACTC TCATCGCACT GGCATTAACGGCCTCA-ATGCAT TCTTTACATGATCGCAAAAGCGACCC

*G. rigida* 049 GACAATGTC GTGTACTC TCATCGCACT GGCATTAACGGCCTCC-ATGCAT TCTTTACATGATCGCAAAAGCGACCC

*G. rigida* 057 GACAATGTC GTGTACTC TCATCGCACT GGCATTAACGGCCTCC-ATGCAT TCTTTACATGATCGCAAAAGCGACCC

*G. rostrata* 007 GACAATGTC GTGCACTC ATGTCACGCC GGAGCTAATGGCCCTA-ATGCAA TCATTGCATGATATCGAAAGCGACCC

*G. rostrata* 091 GACAATGTC GTGCGCTC ATGTCACATC GGTGCTAATGGCCCTA-ATGCAA TCATTGCATGATATCGAAAGCGACCC

*G. rostrata* AB110813 GACAATGTC GTGCACTC TCATCGCATT GGCATTAATGGCCCTA-ATGCAT TCTTTGCATGATCGCAAAA???????

*G. rostrata* AF367225 GACAATGTC GTGCGCTC ATGTCACATC GGTGCTAATGGCCCTA-ATGCAA TCATTGCATGATATGAAA???????

*G. rostrata* BL051  
*G. schomburgkiana* AB110814  
*G. scortechinii* AF367219  
*G. scortechinii* 012  
*G. scortechinii* 089  
*G. scortechinii* K04  
*G. sp.* AJ312607  
*G. sp.* B1 008  
*G. sp.* B2 011  
*G. sp.* D1 064  
*G. sp.* D1 070  
*G. sp.* D2 P01  
*G. sp.* D3 P02  
*G. sp.* D4 BL04  
*G. sp.* D5 BL052  
*G. sp.* D6 K12  
*G. spicata* 019  
*G. sp.* M1 025  
*G. sp.* M2 068  
*G. sp.* M3 072  
*G. sp.* M4 022  
*G. sp.* M5 069  
*G. sp.* M5 073  
*G. sp.* M6 093  
*G. sp.* M6 094  
*G. sp.* X1 052  
*G. sp.* X2 080  
*G. syzygifolia* AB110815  
*G. tinctoria* AB110817  
*G. trianaii* K02  
*G. urophylla* 001  
*G. urophylla* 081  
*G. vilersiana* AB110818  
*G. xanthochymus* AB110819  
*G. xanthochymus* 034  
*G. xanthochymus* 092  
*G. xanthochymus* BB03  
*G. Xanthochymus cf.* 032  
*Symphonia globulifera* AY682919  
*Tripetalum cymosum* AY145206  
*Clusia rosea*

GACAATGTC-GTGCCTC-ATGTCACGTC GGAGCTAATGGCCTTA-ATGCAA-TCATTGCATTATATCGAAGCGACCCC  
GATAGTGT-GTGCAC-TCGTCTCACT GGCATTAATGGCCCTA-ATGCAA-TCATCGCATACTATCATAA???????  
GACAATGTC-GTGAAC-CTATCGCATC GCGTAAATGGCCCCA-ATGCAA-TCATTGCATGATCACAAAA???????  
GACAATGTC-GTGAAC-CTATCGCATC GCGTAAATGGCCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGAAC-CTATCGCATC GCGTAAATGGCCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGCAC-TCATCGCTTT GGCACCAATGGACCCA-AAGCAAA TCATTGCAAGCTCACAAAGCGACCCC  
GATAGTGT-GTGCAC-TCGTCTCACT GGCATTAATGGCCCTG-ATGCAA-TCATTGCATACTATCGTAGCGACCCC  
GATAGTGT-GTGCAC-TCGTCTCACT GGCATTAATGGCCCTA-ATGCAA-TCATTGCATACTATCGTAGCGACCCC  
GACGATGTC-GTGCCTCATGTCCACATC GGTGCTAATGGCCCTC-ATGCAA-TCATTGCATCATATTGAAGCAACCCC  
GACGATGTC-GTGCCTCATGTCCAC-ATC GGTGCTAATGGCCCTA-ATGCAA-TCATTGCATGATATTGAAGCGACCCC  
GACGATGCC-GTGCCTC-ATGTCACATC GGTGGTAGTGGCCCG-ATGCAG-TCATTGCATGAT????????????  
GACAATGCC-GTGTGTC-ATGTCACCTC GGTGCTAATGGCCCTA-ATGCAA-TCATTGCATGATATTAAAGCGACCCC  
GACAATGTC-GTGTGTC-ATGTCACGTC GGTGGTGGTGGCCCTG-ATGCAA-TCATTGCA-CATAT????????????  
GATAATGTC-GTGCCTC-GTGTCCATC GGTGCTAATGGCCCTA-ATGCAA-TCATTGCATGATATTGAAGCGACCCC  
GACAATGTC-GTGCAC-TCGTCCGATT GGCATCAATGGCCCG-ATGCAA-TCATTGCATGATCTCGAAGCGACCCC  
GACAACGTC-GTGATTC-TCGTCCACATT GGCAGTAATGCCCCCA-GAGCAA-TCGTTGCATGCTCATAAAGCGACCTC  
GACAATGTC-GTGCAC-CTGTCCGATC GGTGTTAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGCAC-CTGTCCGATC GGTGTTAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGCAC-CTGTCCGATC GGTGTTAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGCAC-CTGTCCGATC GGTGTTAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGCAC-CTGTCCGATC GGTGTTAATGGCCCTA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGCAC-CTGTCCGATC GGGGTT-ATGGCCC--ATGCAA-TCATTGCATGATCACAGA????????  
GACAATGTC-GTGCAC-CTGTCCGATC GCGTAAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACGACGTC-GTGCAC-TCGTCCACATT GGCATAATACCCCA-AAGCAA-TCGTTGCATGCTCATAAAGCGACCCC  
GACAATGTC-GTGTACT-TCATCGCACT GGCATTAACGGCTCA-ATGCAT-TCATTGCATGATCGCAAAA????????  
GACAACGTC-GTGCAC-TCGTCCACATT GGCATAATACCCCA-AAGCAA-TCGTTGCATGCTCATAAA????????  
GACAATGTC-GTGCAC-TCGTCTCATT GCGTAAATGGCCCA-ATGCAA-TCATTGCATGATCTTGATGCGACCCC  
GACAATGTC-GTGAAC-CTGTCCGATC GCGTAAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAGCGACCCC  
GACAATGTC-GTGAAC-CTGTCCGATC GCGTAAATGGCCCA-ATGCAA-TCATTGCATGATCACAGAA????????  
GACAACGTC-GTGCAC-TCGTCCACATT GGCATAATACCCCA-AAGCAA-TCGTTGCATGCTCATAAA????????  
GACAACGTC-GTGCAC-TCGTCCACATT GGCATAATACCCCA-AAGCAA-TCGTTGCATGCTCATAAA????????  
GACAATGTC-GTGCAC-TCGTCCACATT GGCATAATACCCCA-AAGCAA-TCGTTGCATGCTCATAAAGCGACCCC  
GACAATGTC-GTGCAC-TCGTCCACATT GGCATAATACCCCA-AAGCAA-TCGTTGCATGCTCATAAAGCGACCCC  
GACAACGTC-GTGATTC-TCGTCCACATT GGCATAATACCCCA-GAGCAA-TCGTTGCATGCTCATAA????????  
GAAATATGTC-GTGCAC-TCGTCCCTC GGCATGACGTCTTCA-GGGCA-TCATAACGCGACCCAG????????  
GACGACGTC-GTGCCTC??  
GACATCGTC-GTGCCTC-CTGTCTCTTG-GTACCAGCGCCCGA-AAAAGCATAACTGCTTGCTCCTAATGCGACCCC

APPENDIX VI

Matrix of *trnS-trnG* sequences and binary gap coded characters. Asterisks indicate excluded characters in the phylogenetic analysis.

\*\*\*\*\*

	10	20	30	40	50	60	70	80	90	100											
<i>G_acuticosta</i> BB01	.... ....	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT
<i>G_balica</i> 021	TGTC	TATGA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_cataractalis</i> 077	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_celebica</i> 002	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATT	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_celebica</i> 003	????	????	????	????	????	????	????	????	????	????	TA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	CAAT	TAT
<i>G_celebica</i> 009	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATT	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_celebica</i> 023	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_celebica</i> 027	GGTC	TATAA	ATAACCC	TATT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTC	AAT	TAT	AT	
<i>G_celebica</i> 033	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATT	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_celebica</i> 059	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_celebica</i> 28	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_costata</i> BL03	TCTT	ATCAC	AATAAT	TACCC	TGTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_cymosa</i> 037	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	CGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_cymosa</i> 042	TGTC	TATAA	ATAACCC	TATT	GTC	GAGT	AAGAGT	ATG	GGATA	AGAT	CCTAAA	AGAAT	CATA	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_diospyrifolia</i> var. <i>minor</i> BB06	GGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_diospyrifolia</i> var. <i>diospyrifolia</i> 078	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_discoidea</i> SL01	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_dulcis</i> 050	GGTC	TATAA	ATAACCC	TATT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_forbesii</i> 024	GGAT	TATCG	ATT	ACCC	TATT	GTC	GAGT	AAGAGT	GTATA	AGAT	CCAAAA	AGAAT	CATC	----	ATCAAAA	TTATAGT	GTT	AAT	TAT	AT	
<i>G_lateriflora</i> 040	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGAG	AGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_magostana</i> LAO	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_malaccensis</i> var. <i>malaccensis</i> BB08	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_malaccensis</i> var. <i>malaccensis</i> BB09	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_malaccensis</i> var. <i>pseudomangostana</i> SP01	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	TTCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_mangostana</i> 096	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_megaphylla</i> 041	GGTC	TATAA	ATAACCC	TTTT	GTC	GAGG	AAGAGT	ATGGA	GGATA	GGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_nitida</i> 119	AGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_penangiana</i> 006	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_picrorhiza</i> 039	TGTC	TATAA	ATAACCC	TATT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_rigida</i> 029	???	TATAG	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_rigida</i> 045	GGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_rigida</i> 049	????	????	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_rigida</i> 057	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_rostrata</i> 007	????	????	????	????	????	????	????	????	????	????	????	????	????	????	ATCAT	CGA	CGAAAT	CACAA	TTACAGT	GTT	AAT
<i>G_rostrata</i> 09091	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATAGG	ATACCT	AAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_scortechinii</i> 089	????	????	????	????	????	????	????	????	????	????	????	????	????	????	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_sp.</i> B1 011	????	????	????	????	????	????	????	????	????	????	????	????	????	????	ATCAAAA	TTATAGT	GTC	AAT	TAT	AT	
<i>G_sp.</i> D1 064	GGAC	TATAG	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	GGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_sp.</i> D1 07071	????	????	????	????	????	????	????	????	????	????	????	????	????	????	ATCCAAA	TTACT	TGTT	AAT	TAT	AT	
<i>G_sp.</i> D3 P02	TGTC	TATAA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	GGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	
<i>G_sp.</i> M4 022	GGTC	TATGA	ATAACCC	TTTT	GTC	GAGT	AAGAGT	ATGGA	GGATA	AGAT	CCTAAA	AGAAT	CATC	----	ATCAAAA	TTACAGT	GTT	AAT	TAT	AT	



```

G_rigida 057 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_rostrata 007 -TGTATCTCTT-CAGGGAACCCAACGAAATTCG-AAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_rostrata 09091 -TGTATCTCTT-CAGGGAACCCAACGAGATTCG-AAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_scortechinii 089 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. B1 011 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. D1 064 -TGTATCTCTT-CAGGGAACCCAACGAGATTCG-AAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. D1 07071 -TGTATCTCTG-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. D3 P02 -TGTATCTCTT-CAGGGAACCCAACGAGATTCG-AAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. M4 022 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. M5 069 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. M6 093 -TGTATCTCTT-CAGGGAACCCAAC- AATTC-AAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. M6 094 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. X1 052 -TGTATCTCTT-CCGGGAACCCAACGAGATTCGAAAAGGAAGACTCATTAAATGTTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_sp. X2 080 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCATTAAATGTTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_trianii K02 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_urophylla 001 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCA-----TTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA
G_urophylla 081 ?????????????????????????????????????????????????????????????????????????????????????????????????????????????
G_xanthochymus cf. 032 -TGTATCTCTT-CAGGGAACCCAACGAGATTCGAAAAGGAAGACTCATTAAATGTTAATTGAATTCATATCATTTCGTATAGAATTTGATTCTCTATCA

```

```

*****
      210      220      230      240      250      260      270      280      290      300
      |.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|
G_acuticosta BB01 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_balica 021 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_cataractalis 077 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 002 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 003 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 009 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 023 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 027 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 033 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 059 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_celebica 28 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_costata BL03 TACTCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_cymosa 037 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_cymosa 042 TACGCGTGGAAATAGTTCTAC-----TTAA-AATTG-----TTTTGA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_diospyrifolia var. minor BB06 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_diospyrifoliavardios 078 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_discoidea SL01 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_dulcis 050 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_forbesii 024 TACGCGTGGAAATAGTTCTACTTAGAAATGTTTTTCTACTTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_lateriflora 040 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_magostana LAO TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_malaccensis var. malaccensis BB08 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_malaccensis var. malaccensis BB09 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA
G_malaccensis var. pseudomangostana SP01 TACGCGTGGAAATAGTTCTAC-----TTAG-AATTG-----TTTTTA-TTTC AATAAGAACTCGTTTTGTTTTGAATCA

```

*G\_mangostana* 096  
*G\_megaphylla* 041  
*G\_nitida* 119  
*G\_penangiana* 006  
*G\_picrorhiza* 039  
*G\_rigida* 029  
*G\_rigida* 045  
*G\_rigida* 049  
*G\_rigida* 057  
*G\_rostrata* 007  
*G\_rostrata* 09091  
*G\_scortechinii* 089  
*G\_sp.* B1 011  
*G\_sp.* D1 064  
*G\_sp.* D1 07071  
*G\_sp.* D3 P02  
*G\_sp.* M4 022  
*G\_sp.* M5 069  
*G\_sp.* M6 093  
*G\_sp.* M6 094  
*G\_sp.* X1 052  
*G\_sp.* X2 080  
*G\_trianii* K02  
*G\_urophylla* 001  
*G\_urophylla* 081  
*G\_xanthochymuscf* 032

```

TACGCGTGGAAATAGTTCTAC -----
TACACGTGGAAATAGTTCTAC ----- TTAG AATTG ----- TTTTTC -TTTCAATAAGAACTCGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC -----
TACGCGTGGAAATAGTTCTAC -----
TACGCGTGGAAATAGTTCTAC ----- TTAG AATTG ----- TTTT TA -TTTCAATAAGAACTCGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC ----- TTAGAATTGTTTTTTTTTTTTTTTTTTTTTTCACATTA AAAA ACTCTTTTGGTTTTAAATTC A
TACGCGTGGAAATAGTTCTAC ----- TTAGAAATTG ----- TTTTTT -----
TACGCGTGGAAATAGTTCTAC ----- TTATGAAATTG ----- TTTTTTTTTTCAATAAGAACTCGTTTTGTTTTGATTCA
TACGCGTGGAAATAGTTCTAC ----- TTAG AATTGTTTTTTTTTTTTTTTTTTTTTCAATTA AAAA CCCCTTTTGGTTTTGATTCA
TACGCGTGGAAATAGTTCTAC TTAGAA ----- TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGTGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGTGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGTGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGTGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACACGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGCGTGGAAATAGTTCTACTTAGAATTG ----- G AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
TACGTGTGGAAATAGTTCTAC TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA
????????????????????????????????????????????????????????????????????????????????????????????????????????????????
TACACGTGGAAATAGTTCTAC ----- TTAG AATTG ----- TTTTTC -TTTCAATA ----- CGTTTTGTTTTGAATCA

```

310            320            330            340            350            360            370            380            390            400  
 .....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....

*G\_acuticosta* BB01  
*G\_balica* 021  
*G\_cataractalis* 077  
*G\_celebica* 002  
*G\_celebica* 003  
*G\_celebica* 009  
*G\_celebica* 023  
*G\_celebica* 027  
*G\_celebica* 033  
*G\_celebica* 059  
*G\_celebica* 28  
*G\_costata* BL03  
*G\_cymosa* 037  
*G\_cymosa* 042  
*G\_diospyrifolia* var. *minor* BB06

```

AAAAGAGAA TTTT -GTTGACCCCAATTGGTTGGAAC -AAAAAGGTATCGGTGGGTTTCGTGGAAATAGAAAGGGACTTTTC
-----
-----
-----
-----
-----
-----
-----
-----
-----
-----
C AAAGAGAA TTTT -GTTGACCCCAATTGGTTGGAAC -AAAAAGGTATCAGTGGGTTTCGTGGAAATAGAAAGGGACTTTTC
AAAAGAGAA TTTT -GTTGACCCCAATTGGTTGGAAC -AAAAAGGTATCGGTGGGTTTCGTGGAAATAGAAAGGGACTTTTC
AAAAGAGAA TTTT -GTTGACTCCAATTGGTTGGAAC -AAAAAGGTATCGGTGGGTTTGTGGAAATATAAAGGGACTTTTC
-----

```







*G\_sp.* M6 094  
*G\_sp.* X1 052  
*G\_sp.* X2 080  
*G\_trianii* K02  
*G\_urophylla* 001  
*G\_urophylla* 081  
*G\_xanthochymus* cf. 032

```

TCAAATCGAAGAAGTTTTCTTATTTTTTATATCTTTATAGCAATCAATATAAAATATATCTTTATAGCAATCAATATAAAATCA
-----
TCAAATCGAAGAAGTTTTCTTATTTTTTA                    TATCTTATAGCAATCAATATAAAATCA
TCAAATCGAAGAAGTTTTCTTATTTTTTA                    TATCTTATAGCAATCAATATAAAATCA
TCAAATCGAAGAAGTTTTCTTATTTTTTA                    TATCTTATAGCAATCAATATAAAATCA
-----
  
```

*G\_acuticosta* BB01  
*G\_balica* 021  
*G\_cataractalis* 077  
*G\_celebica* 002  
*G\_celebica* 003  
*G\_celebica* 009  
*G\_celebica* 023  
*G\_celebica* 027  
*G\_celebica* 033  
*G\_celebica* 059  
*G\_celebica* 28  
*G\_costata* BL03  
*G\_cymosa* 037  
*G\_cymosa* 042  
*G\_diospyrifolia* var. *minor* BB06  
*G\_diospyrifolia* var. *diospyrifolia* 078  
*G\_discoidea* SL01  
*G\_dulcis* 050  
*G\_forbesii* 024  
*G\_lateriflora* 040  
*G\_magostana* LAO  
*G\_malaccensis* var. *malaccensis* BB08  
*G\_malaccensis* var. *malaccensis* BB09  
*G\_malaccensis* var. *pseudomangostana* SP01  
*G\_magostana* 096  
*G\_megaphylla* 041  
*G\_nitida* 119  
*G\_penangiana* 006  
*G\_picrorhiza* 039  
*G\_rigida* 029  
*G\_rigida* 045  
*G\_rigida* 049  
*G\_rigida* 057

```

          610      620      630      640      650      660      670      680      690      700
...|...|...|...|...|...|...|...|...|...|...|...|...|...|...|...|...|...|...|...|
-----
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
-----
                    AATATTGAGTTAGAGATATCTTTTT - AACCCCTAACTTTACTTTTA
                    AATATTCAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTTACTTTATTTAT
                    AATATTCAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTTATTTAT
-----
                    AATATTGAGTAGGAGATATCTTTTT - AACCCCTAACTTTACTTT
                    AATATTGAGTTAGAGATATCTTTTT - AACCCCTAACTTTACTTT
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
-----
AAGATATAAAAAATATTGATTAGTTGATTGGAAATTGAGTCCGGAGATGCTTTTTTAAACCCCTAACTTTACTTT
-----
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
                    AATATTGAGTTGGAGATATCTTTTT - AACCCCTAACTTTACTTT
-----
  
```





<i>G_diospyrifolia</i> var. <i>diospyrifolia</i> 078	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_discoidea</i> SL01	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_dulcis</i> 050	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_forbesii</i> 024	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_lateriflora</i> 040	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_magostana</i> LAO	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_malaccensis</i> var. <i>malaccensis</i> BB08	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_malaccensis</i> var. <i>malaccensis</i> BB09	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_malaccensis</i> var. <i>pseudomangostana</i> SP01	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_mangostana</i> 096	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_megaphylla</i> 041	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_nitida</i> 119	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_penangiana</i> 006	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_picrorhiza</i> 039	TTAGATCTAAAT GAAT	CCCTAAA	TAGATGAAATATGCTAATTTT	ATCTAGAAA
<i>G_rigida</i> 029	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	CATCTAGAAA
<i>G_rigida</i> 045	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	CATCTAGAAA
<i>G_rigida</i> 049	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	CATCTAGAAA
<i>G_rigida</i> 057	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	CATCTAGAAA
<i>G_rostrata</i> 007	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	ATCTAGAAA
<i>G_rostrata</i> 09091	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	ATCTAGAAA
<i>G_scortechinii</i> 089	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCTAATTTT	ATCTAGAAA
<i>G_sp.</i> B1 011	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_sp.</i> D1 064	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAAGATGCTTATTTT	ATCTAGAAA
<i>G_sp.</i> D1 07071				
<i>G_sp.</i> D3 P02	TAAAATTTAAAGGAAAAATTATTAGGGTAAGG	TTTAAAGAATTCTAT	TTTAAAGGAAATTTTAAATTTT	TAGGGGAAAAATGGGCAAAATTTT
<i>G_sp.</i> M4 022	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_sp.</i> M5 069	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_sp.</i> M6 093	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_sp.</i> M6 094	TAAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATCC	ATCTTTCTAGATGAAAAATGCATATTTT	ATCTAGA
<i>G_sp.</i> X1 052	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_sp.</i> X2 080	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATTAGCTAATTTT	ATCTAGAAA
<i>G_trianii</i> K02	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_urophylla</i> 001	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_urophylla</i> 081	TTAGATCTACAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA
<i>G_xanthochymus</i> cf. 032	TTAGATCTAAAT GAATATTATTTA GGTACG	TTTAAAGAATTCTAT	ACTTTCTAGATGAAATATGCATATTTT	ATCTAGAAA

910 920 930 940 950 960 970 980 990 1000

<i>G_acuticosta</i> BB01	GTATAGA-TATAGAACTAGAAAATTTTTGTCTTTTCTATTTTCATTTT	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_balica</i> 021	GTATAGA-TATAGAACTAGAAAATTTTTGTCTTTTCTATTTTCATTTT	TATATCTATATTTT	AAATTTAAATTTTGA
<i>G_cataractalis</i> 077	GTATAGA-TATAGAACTAGAAAATTTTTGTCTTTTCTATTTTCATTTT	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_celebica</i> 002	GTATAGA-TATAGTAACCTAGAAAATTTTTGTCTTTTCTATTTTCATTTT	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_celebica</i> 003	GAAATGA-TATAGTATAACCTAGAAAATTTTTGTCTTTTCTATTTTCATTTT	TATATCTATATTTA	AAATTTAAATTTTA

<i>G_celebica</i> 009	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_celebica</i> 023	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_celebica</i> 027	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATT A	AAATTTAAATTTTA
<i>G_celebica</i> 033	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_celebica</i> 059	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_celebica</i> 28	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_costata</i> BL03	GTATAGA-TATATACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G_cymosa</i> 037	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	GAATTTCAATTTGA
<i>G_cymosa</i> 042	GTATAGA-TATAGTAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	GAATTTCAATTTTA
<i>G_diospyrifolia</i> var. <i>minor</i> BB06	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_diospyrifolia</i> var. <i>diospyrifolia</i> 078	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_discoides</i> SL01	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_dulcis</i> 050	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC		
<i>G_forbesii</i> 024	GTATAGA-TATAGAACTAGAAAACTTTTGTCTTTCTATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G_lateriflora</i> 040	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G_mangostana</i> LAO	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTCAATTTTA
<i>G_malaccensis</i> var. <i>malaccensis</i> BB08	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_malaccensis</i> var. <i>malaccensis</i> BB09	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_malaccensis</i> var. <i>pseudomangostana</i> SP01	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_mangostana</i> 096	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTCAATTTTA
<i>G_megaphylla</i> 041	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	GATATCTATATTTTC	AAATTTAAATTTGAA
<i>G_nitida</i> 119	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_penangiana</i> 006	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTCACATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_picrorhiza</i> 039	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G_rigida</i> 029	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTT	AAATTTAAATTTTA
<i>G_rigida</i> 045	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTT	AAATTTAAATTTTA
<i>G_rigida</i> 049	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTT	AAATTTAAATTTTA
<i>G_rigida</i> 057	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTT	AAATTTAAATTTTA
<i>G_rostrata</i> 007	GTATAGA-TATAGAACTAGAAAAATTTT		AA
<i>G_rostrata</i> 09091	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	AAATATAGATATAGATTTAAATATAGATATAGAAATGAAAAATAGAA	
<i>G_scortechinii</i> 089	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGT
<i>G.sp.</i> B1 011	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G.sp.</i> D1 064	GTATAGA-TATAGAACTAGAAAAATTTT		A
<i>G.sp.</i> D1 07071			
<i>G.sp.</i> D3 P02	GGGAAAAAAAAAGAAAAAAAAAAAAATTTTTTTTTTTTTTTTCATTTT	TAAATATAGATATAGAAATGAAAAATTTAA	
<i>G.sp.</i> M4 022	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G.sp.</i> M5 069	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G.sp.</i> M6 093	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G.sp.</i> M6 094			
<i>G.sp.</i> X1 052	GTATAGA-TATATACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	GATATCTATATTTTC	AAATTTAAATTTGAA
<i>G.sp.</i> X2 080	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	GATATCTATATTTTC	AAATTTAAATTTGAA
<i>G trianaii</i> K02	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTA	AAATTTAAATTTTA
<i>G urophylla</i> 001	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G urophylla</i> 081	GTATAGA-TATAGAACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	TATATCTATATTTTC	AAATTTAAATTTGA
<i>G_xanthochymus</i> cf. 032	GTATAGA-TATATACTAGAAAAATTTTGTCTTTCTATTTTCATTTTC	GATATCTATATTTTC	AAATTTAAATTTGAA

	1010	1020	1030	1040	1050	1060	1070	1080	1090	1100
<i>G_acuticosta</i> BB01	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_balica</i> 021	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	TGTCATATATTTAAATT	----			
<i>G_cataractalis</i> 077	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 002	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 003	AA-TATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 009	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 023	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 027	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 033	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 059	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_celebica</i> 28	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_costata</i> BL03	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_cymosa</i> 037	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_cymosa</i> 042	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_diospyrifolia</i> var. <i>minor</i> BB06	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_diospyrifolia</i> var. <i>diospyrifolia</i> 078	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_discoidea</i> SL01	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_dulcis</i> 050	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
<i>G_forbesii</i> 024	AA-CATTTAATCTGTTA	----	TATTC	TATATAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_lateriflora</i> 040	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_magostana</i> LAO	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_malaccensis</i> var. <i>malaccensis</i> BB08	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_malaccensis</i> var. <i>malaccensis</i> BB09	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_malaccensis</i> var. <i>pseudomangostana</i> SP01	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_mangostana</i> 096	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_megaphylla</i> 041	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_nitida</i> 119	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_penangiana</i> 006	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_picrorhiza</i> 039	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_rigida</i> 029	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_rigida</i> 045	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_rigida</i> 049	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_rigida</i> 057	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_rostrata</i> 007	AA-CATTTAGTCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_rostrata</i> 09091	AA-CATTTAGTCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G_scortechinii</i> 089	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G.sp.</i> B1 011	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G.sp.</i> D1 064	AA-CATTTAGTCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G.sp.</i> D1 07071	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
<i>G.sp.</i> D3 P02	AA-CATTTAGTCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G.sp.</i> M4 022	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	TGTCATATATTTAAATT	----			
<i>G.sp.</i> M5 069	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		
<i>G.sp.</i> M6 093	AA-CATTTAATCTGTTA	----	TATTC	TATACAATATGTTATATATTAATAATTAATATA	TTTATA	-----	TGTCATATATTTAAATT	----		







*G. discoidea* SL01 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. dulcis* 050 ?01110011111110001010-011-00100000100100010-000001001-11-0-11010101101011????????????????????  
*G. forbesii* 024 001110011111110001-0-0011-000001001001000000000-1-1-11-11-1-1-11-1-1-111-1-1-11-1-----  
*G. lateriflora* 040 001110001111110001-1-010101100000100100010-0000-1-1-11-1-1-1-11-1-11-1-11-1-1-1-1-----1-  
*G. magostana* LAO3 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. malaccensis* var *malaccensis* BB08 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. malaccensis* var *malaccensis* BB09 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. malaccensis* var *pseudomangostana* P01 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. mangostana* 096 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. megaphylla* 041 001110011111110000-1-011-00100000100100010-00011-----1-1-1-11-1-11-1-111-1-1-1-1-----1-  
*G. nitida* 119 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. penangiana* 006 001110011111110001-1-----11-1-1-11-1-11-1-111-1-1-1-1-----  
*G. picrorhiza* 039 001110011111010001-1-010101100001-00000010-0001-1-1-11-11-11-111-1-111-1-1-1-----  
*G. rigida* 029 ??1110011111110001-1-00000000000000100010-0000-1-1-11-11-1-11-1-1-111-1-1-1-1-----1-  
*G. rigida* 045 001110011111110001-1-001-00-1-----1-1-0110-1-1-11-11-1-11-1-1-111-1-1-1-1-----1-  
*G. rigida* 049 ??1110011111110001-1-001-00000000111000010-0000-1-1-11-11-1-11-1-1-111-1-1-1-1-----1-  
*G. rigida* 057 001110011111110001-1-010000000000111000010-0000-11-1-11-11-1-1-11-1-1-111-1-1-1-1-----1-  
*G. rostrata* 007 ?????????011100110001011-00101000100100010-000101000011-0-1101010110101110-----1101001-1-1-000-1  
*G. rostrata* 09091 001110010111110011-1-011-001010001001000010001-1-111-11-1-1-11-1-11-1-11-1-1-1-----  
*G. scortechinii* 089 ??????????11110001010-011-00100000100100010-000001001-11-0-1101010110101100001-011101001-1-1-00010  
*G. spB1* 011 ??????????111110001010-011-00100010100100010-000001001-101000100101011000110010-0110011-1-1-000-  
*G. spD1* 064 001110011111110011-1-01-10101000100100010-0001-1-111-11-1-1-11-1-11-1-11-1-1-1-----1  
*G. spD1* 07071 ????110000011110001-1-011-00101000100101010-0001????????????????????????????????????  
*G. spD3* P02 001110011111110011-1-011-0010100010010000001001-1-111-11-1-----1-1-11-1-1-1-1-----  
*G. spM4* 022 001110011111110001-1-011-00100000100100010-0000-1-1-1-11-11-1-11-1-11-1-11-1-1-----1-  
*G. spM5* 069 001110011111110001-1-011-00100000100100010-0000-1-1-11-11-1-11-1-11-1-11-1-1-----1-  
*G. spM6* 093 00111001011111101010-010100100000100110010-00000111-11-0-1101010110101100001-011101001-011-00100  
*G. spM6* 094 ????10011111110001010-011-00100000100100010-000000001-11-0-1-11010111101????????????????  
*G. spX1* 052 001110001111110000-1-011-00100000100100010-00011-----1-11-1-1-11-1-11-1-111-1-1-1-1-----1-  
*G. spX2* 080 001110001111110000-1-011-00100000100100010-00011-----1-11-1-1-11-1-11-1-1-1-1-1-----1-  
*G. trianaii* K02 001110011111110001-0-0111-00100000100100010-0000-1-1-1-----11-1-1-11-1-11-1-111-1-1-1-1-----1-  
*G. urophylla* 001 001110011111110001-1-011-00100000100100010-0000-1-1-11-11-1-11-1-11-1-11-1-1-----1-  
*G. urophylla* 081 ???01001-11-0-1101010110101100001-011111001-1-1-01-00  
*G. xanthochymus* cf. 032 ?01110011111110000-1-011-00100000100100010-00011-----1-11-1-1-11-1-11-1-111-1-1-1-1-----1-

\* \* \*\*\*\*\*

110

....|....|....|....

*G. acuticosta* BB01 -----1-----1?????  
*G. balica* 021 -----1?????  
*G. cataractalis* 077 -----1-----1?????  
*G. celebica* 002 0000000100?????????  
*G. celebica* 003 -----1-----1?????  
*G. celebica* 009 -----1-----1?????  
*G. celebica* 023 -----1-----1?????  
*G. celebica* 027 ??????????????????

<i>G. celebica</i> 033	-----1-----1?????
<i>G. celebica</i> 059	-----1-----1?????
<i>G. celebica</i> 28	-----1-----1?????
<i>G. costata</i> BL03	--1-----1?????
<i>G. cymosa</i> 037	---1-----1-1?????
<i>G. cymosa</i> 042	---1-----1?????
<i>G. diospyrifolia</i> var <i>minor</i> BB06	-----1-----1?????
<i>G. diospyrifolia</i> var <i>diospyrifo</i>	-----1-----1?????
<i>G. discoidea</i> SL01	-----1-----1-1???
<i>G. dulcis</i> 050	????????????????
<i>G. forbesii</i> 024	-1-----1?????
<i>G. lateriflora</i> 040	-----1?????
<i>G. magostana</i> LAO3	-----1-----1?????
<i>G. malaccensis</i> var <i>malaccensis</i> BB08	-----1-----1?????
<i>G. malaccensis</i> var <i>malaccensis</i> BB09	-----1-----1?????
<i>G. malaccensis</i> var <i>pseudomangostana</i> SP01	-----1-----1?????
<i>G. mangostana</i> 096	-----1-----1?????
<i>G. megaphylla</i> 041	-----1?????
<i>G. nitida</i> 119	-----1-----1?????
<i>G. penangiana</i> 006	-----1-----1-1-1
<i>G. picrorhiza</i> 039	1-----1?????
<i>G. rigida</i> 029	-----1?????
<i>G. rigida</i> 045	-----1?????
<i>G. rigida</i> 049	-----1?????
<i>G. rigida</i> 057	-----1?????
<i>G. rostrata</i> 007	000--0--00????????
<i>G. rostrata</i> 09091	-----1????????
<i>G. scortechinii</i> 089	0000--0--0????????
<i>G. sp.</i> B1 011	100--0--0????????
<i>G. sp.</i> D1 064	-----1????
<i>G. sp.</i> D1 07071	????????????????
<i>G. sp.</i> D3 P02	-----1????????
<i>G. sp.</i> M4 022	-----1????????
<i>G. sp.</i> M5 069	--1-----1?????
<i>G. sp.</i> M6 093	00000-0-00????????
<i>G. sp.</i> M6 094	????????????????
<i>G. sp.</i> X1 052	-----1?????
<i>G. sp.</i> X2 080	-----1-1???
<i>G. trianaii</i> K02	-----1?????
<i>G. urophylla</i> 001	-----1?????
<i>G. urophylla</i> 081	00000-0-000-1000??
<i>G. xanthochymus</i> cf 032	-----1?????

















*G. diospyrifolia* var *minor* BB06 TGCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. discoidea*\_SL01 TGCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. malaccensis* var. *malaccensis* BB08 TACCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. malaccensis* var. *pseudomangostana* SP01 TGCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. malaccensis* BB09 TGCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. mangostana* LAO TACCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. megaphylla* 041 TGTCCTTATTACTTT-----GTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----TGGA  
*G. penangiana* 006 TGCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. rigida* 029 TGCCCTTATTACTTTACTCT-----ACAGCGTATATCTACGGAACTAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. rigida* 045 ???  
*G. rigida* 049 TGCCCTTATTACTTTACTCT-----ACAGCGTATATCTACGGAACTAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. rigida* 057 TGCCCTTATTACTTTACTCT-----ACAGTGTATATCTACGGAACTAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. rostrata* 007 ???  
*G. spD2* P02 TGCACTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----  
*G. spM4* 022 ???  
*G. spX2* 080 ???  
*G. trianaii* K02 TGCCCTTATTACTTTACTCT-----AAAGTGTATATCTACGGAAATAGCAATACAATATAGCACCCGCCTTCTCGCACAACGCAGCGCGAA-----

\*\*\*\*\*

1310 1320 1330 1340 1350 1360 1370 1380 1390

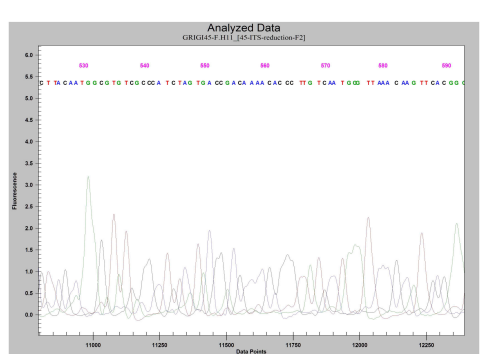
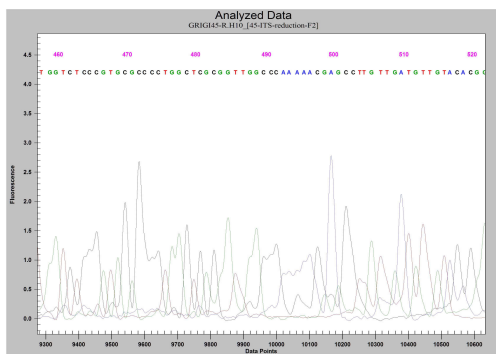
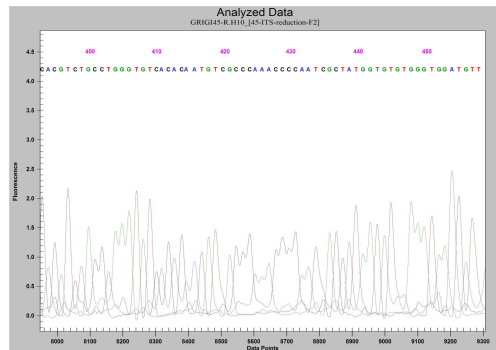
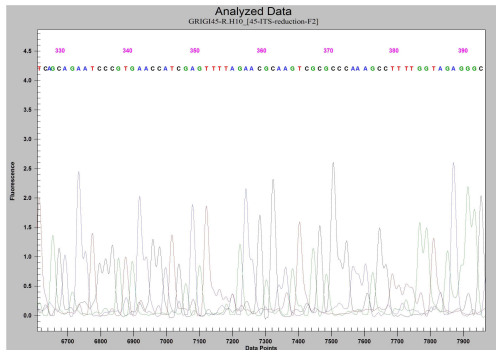
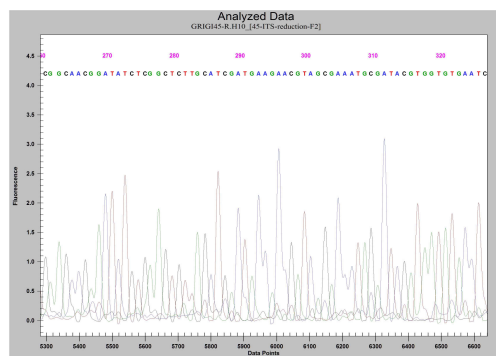
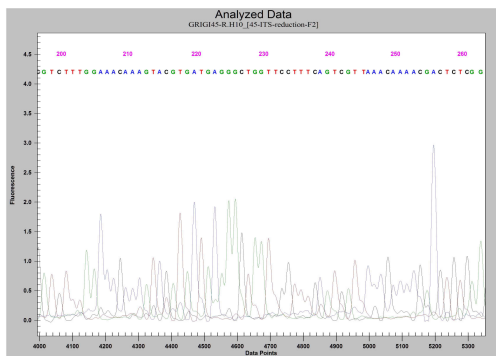
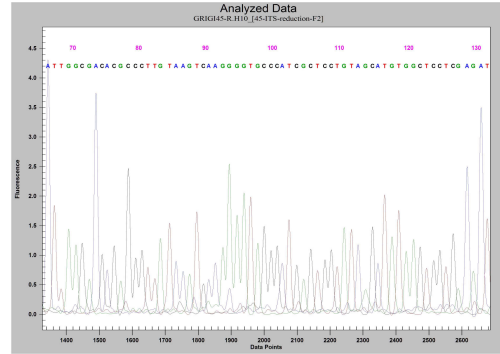
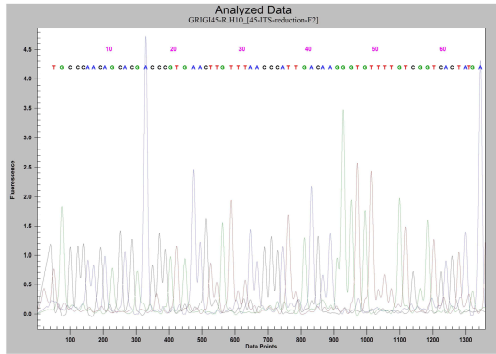
1400

....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|  
*G. acuticosta* BB01 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0001  
*G. cataractalis* 077 -----TGGAATCGAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????????????0100  
*G. celebica* 002 -----GGGAA-TCAAAAGGAAGG??  
*G. celebica* 023 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0000  
*G. celebica* 033 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATG??0101  
*G. cymosa* 037 ???0000  
*G. diospyrifolia* var *dumosa* 078 ???1000  
*G. diospyrifolia* var *minor* BB06 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTTATATA????????1000  
*G. discoidea*\_SL01 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0000  
*G. malaccensis* var. *malaccensis* BB08 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0000  
*G. malaccensis* var. *pseudomangostana* SP01 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0001  
*G. malaccensis* BB09 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTTATATTAA????????0  
*G. mangostana* LAO -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0000  
*G. megaphylla* 041 TAGAAAGAAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????????0000  
*G. penangiana* 006 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTTTTTCATTAATTG1000  
*G. rigida* 029 -----CGGAAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTTTTTC????????1000  
*G. rigida* 045 ???0001  
*G. rigida* 049 -----CGGAAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTTTCATATTA????????  
*G. rigida* 057 -----CGGAAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????????0  
*G. rostrata* 007 ???0000  
*G. spD2* P02 -----TGGAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0000  
*G. spM4* 022 ???0000  
*G. spX2* 080 ???0000  
*G. trianaii* K02 -----TGGAATCAAAGAATGGAAATAGAAAGAAAGGGTTTGAATGAAATTTTAAGAAATACGTATGCTGTACACTTTTT????????????0000



## APPENDIX VIII

### Electropherogram of *Garcinia rigida* Miq. (accession 045) upstream sequence of ITS.



# APPENDIX IX

## Electropherogram of *Garcinia mangostana* L. (accession 096) upstream sequence of ITS.

